

REFERENCES

REFERENCES

- Anonymous, (1987). **The Wealth of India: Raw materials.** New Delhi: Publication and Information Directorate, Council of Scientific and Industrial Research.
- Ashbee, H. R. and Evans, E. G. (2002). Immunology of diseases associated with *Malassezia* species. **Clinical Microbiology**, 15(1), 21-57.
- Attele, A. S., Wu, J. A. and Yuan, C. S. (1999). Ginseng pharmacology. Multiple constituents and multiple actions. **Biochemical Pharmacology**, 58(11), 1685-1693.
- Avijgan, M., Mahboubi, M., Darabi, M., Saadat, M., Sarikhani, S. and Nazilla, K. (2010). Overview on *Echinophora platyloba*, a synergistic antifungal agent candidate. **Journal of Yeast and Fungal Research**, 1(5), 88-94.
- Bharatiya Vidya Bhavan's Swami Prakashananda Ayurveda Research Center. (1992). **Selected Medicinal Plants of India.** Bombay: Chemexcil.
- Bhattacharya, S. K., Bhattacharya, A. and Chakrabarti, A. (2000). Adaptogenic activity of Siotone, a polyherbal formulation of Ayurvedic rasayanas. **Indian Journal of Experimental Biology**, 38(2), 119-28.
- Bhattacharya, A., Murugandam, A. V., Kumar, V. and Bhattacharya, S. K. (2002). Effect of polyherbal formulation, EuMil, on neurochemical perturbations induced by chronic stress. **Indian Journal of Experimental Biology**, 40(10), 1161-1163.
- Bopana, N. and Saxena, S. (2007). *Asparagus racemosus*-Ethnopharmacological evaluation and conservation needs. **Journal of Ethnopharmacology**, 110(1), 1-15.
- Cragg, G. M. and Newman, D. J. (2001). Natural product drug discovery in the next millennium. **Pharmaceutical Biology**, 39, 8-17.
- Cutsem, J. van, Gerven, F. van, Fransen, J., Schrooten, P. and Janssen, P. A. (1990). The *in vitro* antifungal activity of ketoconazole, zinc pyrithione, and selenium sulfide against *Pityrosporum* and their efficacy as a shampoo in the treatment of experimental pityrosporosis in guinea pigs. **Journal of the American Academy of Dermatology**, 22(6), 993-998.

- Dange, P. S., Kanitkar, U. K. and Pendse, G. S. (1969). Amylase and lipase activities in the root of *Asparagus racemosus*. **Planta Medica**, 17(4), 393-395.
- Dhuley, J. N. (1997). Effect of some Indian herbs on macrophage functions in ochratoxin A treated mice. **Journal of Ethnopharmacology**, 58(1), 15-20.
- Dinan, L., Savchenko, T. and Whiting, P. (2001). Phytoecdysteroids in the genus *Asparagus* (Asparagaceae). **Phytochemistry**, 56(6), 569-576.
- Diwanay, S., Chitre, D. and Patwardhan, B. (2004). Immunoprotection by botanical drugs in cancer chemotherapy. **Journal of Ethnopharmacology**, 90(1), 49-55.
- Elisa, B., Giuliano, B., Vincenzo, A., Behzad, Z., Ebrahim S., Felice, S., et al. (2007). Saponins from *Allium minutiflorum* with antifungal activity. **Phytochemistry**, 68(5), 596-603.
- Faergemann, J., Hersle, K. and Nordin, P. (1997). Pityriasis versicolor: clinical experience with Lamisil cream and Lamisil DermGel. **Dermatology**, 194(1), 19-21.
- Farnsworth, N. R., Akerele, O., Bingel, A. S., Soejarto, D. D., and Guo, Z. (1985). Medicinal plants in therapy. **Bulletin of the World Health Organization**, 63(6), 965-981.
- Farthing, C. F., Staughton, R. C. D. and Rowland, Payne, C. M. E. (1985). Skin disease in homosexual patients with Acquired Immune Deficiency Syndrome (AIDS) and lesser forms of human T cell Leukaemia Virus (HTLV III) disease. **Clinical and Experimental Dermatology**, 10(1), 3-12.
- Francis, G., Kerem, Z., Makkar, P. S. H. and Becker, K. (2002). The biological action of saponins in animal systems: a review. **British Journal of Nutrition**, 88(6), 587-605.
- Gautam, M., Diwanay, S., Gairola, S., Shinde, Y., Patki, P. and Patwardhan, B. (2004). Immunoadjuvant potential of *Asparagus racemosus* aqueous extract in experimental system. **Journal of Ethnopharmacology**, 91(2-3), 251-255.
- Gemmer, C. M., DeAngelis, Y. M., Theelen, B., Boekhout, T. and Dawson, T. L. (2002). Fast, noninvasive method for molecular detection and differentiation of *Malassezia* yeast species on human skin and application for the method to

- dandruff microbiology. **Journal of Clinical Microbiology**, 40(9), 3350-3357.
- Ghumare, B. C., Vadlamudi, V. P. and Rajurkar, S. R. (2004). Effect of *Asparagus racemosus* on growth and development of testes in wistar rats. **Aryavaidyan**, 18(1), 45-48.
- Gupta, A. K., (2002). Butenafine: an update of its use in superficial mycoses. **Skin Therapy Letter**, 7(7), 1-5.
- Gupta, A. K., Batra, R., Bluhm, R. and Faergemann, J. (2003a). Pityriasis versicolor. **Dermatology Clinical**, 21(3), 413-429.
- Gupta, A. K., Bluhm, R., Cooper, E. A., Summerbell, R. C. and Batra, R. (2003b). Seborrheic dermatitis. **Dermatology Clinical**, 21(3), 401-412.
- Gupta, A. K., Roma, B., Robyn, B., Teun, B. and Thomas, L. D. (2004). Skin diseases associated with *Malassezia* species. **Journal of the American Academy of Dermatology**, 51(5), 785-798.
- Hayes, P. Y., Jahidin, A. H., Lehmann, R., Penman, K., Kitching, W. and De Voss, J. J. (2007). Steroidal saponins from the roots of *Asparagus racemosus*. **Phytochemistry**, 69(3), 796-804.
- Joglekar, G.V., Ahuja, R. H. A. and Balwani, J. H. (1967). Galactogogue effect of *Asparagus racemosus*. **Indian Medical Journal**, 61(7), 165.
- Johnson, B. A. and Nunley, J. R. (2000). Treatment of seborrheic dermatitis. **American Family Physician**, 61(9), 2703-2704.
- Kamat, J. P., Boloor, K. K., Devasagayam, T. P. and Venkatachalam, S. R. (2000). Antioxidant properties of *Asparagus racemosus* against damage induced by gamma-radiation in rat liver mitochondria. **Journal of Ethnopharmacology**, 71(3), 425-435.
- Khanna, A. K., Chander, R. and Kapoor, N. K. (1991). Hypolipidaemic activity of Abana in rats. **Fitoterapia (Italy)**, 62(3), 271-275.
- Kinnunen, T. and Koskela, M. (1991). Antibacterial and antifungal properties of propylene glycol, hexylene glycol, and 1,3-butyleneglycol *in vitro*. **Acta Dermato-Venereologica**, 71(2), 148-150.

- Kulkarni, S. K. and Verma, A. (1993). Protective effect of Mentat (BR-16A) A herbal preparation, on alcohol abstinence-induced anxiety and convulsions. **Indian Journal of Experimental Biology**, 31(5), 435-442.
- Lalitha, T. and Venkataraman, L. V. (1991). Antifungal activity and mode of action of saponins from *Madhuca butyracea* Macb. **Indian Journal of Experimental Biology**, 29(6), 558-562.
- Ling, M. R. (2001) Topical tacrolimus and pimecrolimus: future directions. **Seminars in Cutaneous Medicine and Surgery**, 20(4), 268-274.
- Luccaa, A. J., Bland, J. M., Boue, S., Vigo, C. B., Cleveland, T. E. and Walsh, T. J. (2006). Synergism of CAY-1 with amphotericin B and itraconazole. **Cancer Chemotherapy**, 52(6), 285-287.
- Mandal, S. C., Kumar, C. K. A., Lakshmi, M., Sinha, S., Murugesan, T., Saha, B. P. and Pal, M. (2000a). Antitussive effect of *Asparagus racemosus* root against sulphur dioxide-induced cough in mice. **Fitoterapia**, 71(6), 686-689.
- Mandal, S. C., Nandy, A., Pal, M. and Saha, B. P. (2000b). Evaluation of antibacterial activity of *Asparagus racemosus* Willd. root. **Phytotherapy Research**, 14(2), 118-119.
- Marc, S., Tomofumi, M., Marie, A. and Laeaille, D. (2007). Steroidal saponins from *Asparagus acutifolius*. **Phytochemistry**, 68(20), 2554-2562.
- Marston, A., Gafner, F., Dossaji, S. F. and Hostettmann, K. (1988). Fungicidal and molluscicidal saponins from *Dolichos kilimandscharicus*. **Phytochem**, 27(5), 1325-1326.
- McGinley, K. J. and Leyden, J. J. (1982). Antifungal activity of dermatological shampoos. **Archives Dermatology Research**, 272, 339-342.
- Miyakoshi, M., Tamura, Y., Masuda, H., Mizutani, K., Tanaka, O., Ikeda, T., et al. (2000). Antiyeast steroidal saponins from *Yucca schidigera* (Mohave yucca), a new anti-food-deteriorating agent. **Journal of Natural Products**, 63(3), 332-338.
- Mshvildadze, V., Favel, A., Delmas, F., Elias, R., Faure, R., Decanosidze, Q., et al. (2000). Antifungal and antiprotozoal activities of saponins from *Hedera colchica*. **Pharmazie**, 55(4), 325-326.

- Nakagawa, H., Etoh, T., Yokota, Y., Ikeda, F., Hatano, K., Teratani, N., et al. (1996). Tacrolimus has antifungal activities against *Malassezia furfur* isolated from healthy adults and patients with atopic dermatitis. **Clinical Drug Investigation**, 12, 245-250.
- Nanal, B. P., Sharma, B. N., Ranade, S. S. and Nande, C. V. (1974). Clinical study of Shatavari (*Asparagus racemosus*). **Journal of Research in Indian Medicine**, 9, 23-29.
- Oleszek, W. A. (2002). Chromatographic determination of plant saponins. **Journal of Chromatography A**, 967(1), 147-162.
- Pandey, S. K., Sahay, A., Pandey, R. S. and Tripathi, Y. B. (2005). Effect of *Asparagus racemosus* (Shatavari) rhizome on mammary gland and genital organs of pregnant rat. **Phytotherapy Research**, 19(8), 721-724.
- Parihar, M. S. and Hemnani, T. (2004). Experimental excitotoxicity provokes oxidative damage in mice brain and attenuation by extract of *Asparagus racemosus*. **Journal of Neural Transmission**, 111(1), 1-12.
- Patel, A. B. and Kanitkar, U. K. (1969). *Asparagus racemosus* willd - form bordi, as a galactogogue, in buffaloes. **Indian Veterinary Journal**, 46(8), 718-721.
- Polacheck, I., Levy, M., Guizie, M., Zehavi, U., Naim, M. and Evron, R. (1991). Mode of action of the antimycotic agent G2 isolated from alfalfa roots. **Zenfralbl Bakteriol**, 275, 504-512.
- Price, K. R., Johnson, I. T. and Fenwick, G. R. (1987). The chemistry and biological significance of saponins in foods and feedstuffs. **Critical Reviews in Food Science and Nutrition**, 26(1), 27-135.
- Reamyindee, N., Phrompittayarat, W., Putalun W., Tanaka, H. and Ingkaninan, K. (2011). Determination of steroid saponins in *Asparagus racemosus* roots using ELISA method with monoclonal antibody against Shatavarin IV. In **Proceedings of the Third International Conference on Natural Products for Health and Beauty** (p.162). Bangkok: Rangsit University.
- Rege, N. N. and Dahanukar, S. A. (1993). Quantitation of microbicidal activity of mononuclear phagocytes: an *in vitro* technique. **Journal of Postgraduate Medicine**, 39(1), 22-25.

- Rege, N. N., Nazareth, H. M., Isaac, A., Karandikar, S. M. and Dahanukar, S. A. (1989). Immunotherapeutic modulation of intraperitoneal adhesions by *Asparagus racemosus*. **Journal of Postgraduate Medicine**, 35(4), 199-203.
- Rege, N. N., Thatte, U. M. and Dahanukar, S. A. (1999). Adaptogenic properties of six rasayana herbs used in Ayurvedic medicine. **Phytotherapy Research**, 13(4), 275-291.
- Rao, A. R. (1981). Inhibitory action of *Asparagus racemosus* on DMBA-induced mammary carcinogenesis in rats. **International Journal of Cancer**, 28(5), 607-610.
- Sabnis, P. B., Gaitonde, B. B. and Jetmalani, M. (1968). Effects of alcoholic extract of *Asparagus racemosus* on mammary glands of rats. **Indian Journal of Experimental Biology**, 6, 55-57.
- Saxena, V. K. and Chourasia, S. (2001). A new isoflavone from the roots of *Asparagus racemosus*. **Fitoterapia**, 72(3), 307-309.
- Sekine, T., Fukasawa, N., Kashiwagi, Y., Ruangrungsi, N. and Murakoshi, I. (1994). Structure of asparagamine A, a novel polycyclic alkaloid from *Asparagus racemosus*. **Chemical and Pharmaceutical Bulletin**, 42(6), 1360-1362.
- Sekine, T., Fukasawa, N., Murakoshi, I. and Ruangrungsi, N. (1997). A 9,10-dihydrophenanthrene from *Asparagus racemosus*. **Phytochemistry**, 44(4), 763-764.
- Shakila, R. J., Vasundhara T. S. and Rao, D. V. (1996). Inhibitory effect of spices on *in vitro* histamine production and histidine decarboxylase activity of *Morganella morganii* and on the biogenic amine formation in mackerel stored at 30 degrees C. **Zeitschrift für Lebensmittel-Untersuchung und-Forschung**, 203(1), 71-76.
- Sharma, S., Ramji, S., Kumari, S. and Bapna, J. S. (1996). Randomized controlled trial of *Asparagus racemosus* (shatavari) as lactagogue in lactational inadequacy. **Indian Paediatrics**, 33(8), 675-677.
- Shimoyamada, M., Suzuki, M., Sonta, H., Maruyama, M. and Okubo, K. (1990). Antifungal activity of the saponin fraction obtained from *Asparagus officinalis* L. and its active principle. **Agricultural Biological Chemistry**, 54(10), 2553-2557.

- Smith, K. J., Skelton, H. G, Yeager, J., Ledsky, R., McCarthy, W., Baxter, D., et al., (1994). Cutaneous findings in HIV-1 positive patients: a 42-month prospective study. **Journal of the American Academy of Dermatology**, 31(5), 746-754.
- Sparg, S. G., Light, M. E. and Staden, J. van. (2004). Biological activities and distribution of plant saponins. **Journal of Ethnopharmacology**, 94(2-3), 219-243.
- Takechi, M., Matsunami, S., Nishizawa, J., Uno, C. and Tanaka, Y. (1999). Haemolytic and antifungal activities of saponins or anti-ATPase and antiviral activities of cardiac glycosides. **Planta Medica**, 65(6), 585-586.
- Takeo, K. and Nakai, E. (1986). Mode of cell growth of *Malassezia (Pityrosporum)* as revealed by using plasma membrane configuration as natural makers. **Canadian Journal of Microbiology**, 32, 389-394.
- Thatte, U. M., and Dahanukar, S. A. (1988). Comparative study of immunomodulating activity of Indian medicinal plants, lithium carbonate and glucan. **Methods and Findings in Experimental and Clinical Pharmacology**, 10(10), 639-644.
- Uma, B., Prabhakar, K. and Rajendran, S. (2009). Anticandidal activity of *Asparagus racemosus*. **Indian Journal of Pharmaceutical Sciences**, 71(3), 342-343.
- Uchida, K., Nishiyama, Y., Tanaka, T. and Yamaguchi, H. (2003). *In vitro* activity of novel imidazole antifungal agent NND-502 against *Malassezia* species. **International Journal of Antimicrobial Agents**, 21(3), 234-238.
- Vardy, D. A., Zvulunov, A., Tchetov, T., Biton A. and Rosenman, D. (2000). A double-blind, placebo-controlled trial of a ciclopirox olamine 1% shampoo for the treatment of scalp seborrheic dermatitis. **Journal of Dermatological Treatment**, 11(2), 73-77.
- Visavadiya, N. P. and Narasimhacharya A. V. R. L. (2005). Hypolipidemic and antioxidant activities of *Asparagus racemosus* in hypercholesterolemic rats. **Indian Journal of Pharmacology**, 37(6), 376-380.
- Warner, R. R., Schwartz, J. R., Boissy, Y. and Dawson, T. L. (2001). Dandruff has an altered stratum corneum ultrastructure that is improved with zinc pyrithione

- shampoo. **Journal of the American Academy of Dermatology**, 45(6), 897-903.
- Warren, R., Schwartz, J. R., Sanders, L. M. and Juneja, P. S. (2003). Attenuation of surfactant-induced interleukin-1alpha expression by zinc pyrithione. **Exogenous Dermatology**, 2, 23-27.
- Warren, L. (2008). **Review of Medical Microbiology and Immunology**. California: McGraw-Hill Medical.
- Wiboonpun, N., Phuwapraisirisan, P. and Tip-pyang, S. (2004). Identification of antioxidant compound from *Asparagus racemosus*. **Phytotherapy Research**, 18(9), 771-773.
- Zhang, J. D., Zheng, X., Cao, Y. B., Chen, H. S., Yan, L., An, M. M., et. al. (2006). Antifungal activities and action mechanisms of compounds from *Tribulus terrestris* L. **Journal of Ethnopharmacology**, 103(1), 76-84.

APPENDIX

APPENDIX A PERCENTAGE OF YIELD

The percent yield of *A. racemosus* roots extract is calculated by using this equation:

$$\boxed{\text{% yield} = [\text{dried extract weight (g)} / \text{plant material sample weight (g)}] \times 100}$$

1. % yield of AR 1-H extract

$$\begin{aligned}\text{% yield} &= [1.03 \text{ (g)} / 1.8 \text{ (Kg)}] \times 100 \\ &= 0.06\end{aligned}$$

2. % yield of AR 1-E extract

$$\begin{aligned}\text{% yield} &= [115.35 \text{ (g)} / 1.8 \text{ (Kg)}] \times 100 \\ &= 6.41\end{aligned}$$

3. % yield of AR 1-A extract

$$\begin{aligned}\text{% yield} &= [87.65 \text{ (g)} / 1.8 \text{ (Kg)}] \times 100 \\ &= 4.87\end{aligned}$$

4. % yield of AR 1-B extract

$$\begin{aligned}\text{% yield} &= [34.72 \text{ (g)} / 1.8 \text{ (Kg)}] \times 100 \\ &= 1.93\end{aligned}$$

5. % yield of AR 1-S extract

$$\begin{aligned}\text{% yield} &= [87.45 \text{ (g)} / 1.8 \text{ (Kg)}] \times 100 \\ &= 4.86\end{aligned}$$

6. % yield of AR 2-H extract

$$\begin{aligned}\text{% yield} &= [0.46 \text{ (g)} / 1.8 \text{ (Kg)}] \times 100 \\ &= 0.05\end{aligned}$$

7. % yield of AR 2-E extract

$$\begin{aligned}\text{% yield} &= [57.29 \text{ (g)} / 1 \text{ (Kg)}] \times 100 \\ &= 5.73\end{aligned}$$

8. % yield of AR 2-A extract

$$\begin{aligned}\text{% yield} &= [55.12 \text{ (g)} / 1 \text{ (Kg)}] \times 100 \\ &= 5.51\end{aligned}$$

9. % yield of AR 2-B extract

$$\begin{aligned}\% \text{ yield} &= [23.37 \text{ (g)} / 1 \text{ (Kg)}] \times 100 \\ &= 2.34\end{aligned}$$

10. % yield of AR 2-S extract

$$\begin{aligned}\% \text{ yield} &= [72.85 \text{ (g)} / 1 \text{ (Kg)}] \times 100 \\ &= 7.29\end{aligned}$$

11. % yield of AR 3-H extract

$$\begin{aligned}\% \text{ yield} &= [7.43 \text{ (g)} / 4.8 \text{ (Kg)}] \times 100 \\ &= 0.15\end{aligned}$$

12. % yield of AR 3-E extract

$$\begin{aligned}\% \text{ yield} &= [525.07 \text{ (g)} / 4.8 \text{ (Kg)}] \times 100 \\ &= 10.94\end{aligned}$$

13. % yield of AR 3-A extract

$$\begin{aligned}\% \text{ yield} &= [247.16 \text{ (g)} / 4.8 \text{ (Kg)}] \times 100 \\ &= 5.15\end{aligned}$$

14. % yield of AR 3-B extract

$$\begin{aligned}\% \text{ yield} &= [115.54 \text{ (g)} / 4.8 \text{ (Kg)}] \times 100 \\ &= 2.41\end{aligned}$$

15. % yield of AR 3-S extract

$$\begin{aligned}\% \text{ yield} &= [215.56 \text{ (g)} / 4.8 \text{ (Kg)}] \times 100 \\ &= 4.49\end{aligned}$$

16. % yield of AR 3-En extract

$$\begin{aligned}\% \text{ yield} &= [32.4 \text{ (g)} / 150 \text{ (g)}] \times 100 \\ &= 1.22\end{aligned}$$

APPENDIX B PREPARATION OF STERILE MEDIA FOR YEAST

1. Sabouraud dextrose agar (Becton dickinson)

Pancreatic digest of casein	5.0	g
Peptic digest of animal tissue	5.0	g
Dextrose	40	g
Agar	15	g

Suspend the mixture 65 g in 1 lit of water, and autoclave at 121 °C 15 minute. Cool the sterile media to <50°C before pipette media 20 ml in to sterile plastic disc (10 cm). Dry plates 1 days at room temperature, or 30 min with the lids slightly off at 37°C or in a laminar flow hood. Store dried plates wrapped at 4°C.

2. Sabouraud dextrose agar (Becton dickinson) + 1% olive oil

Pancreatic digest of casein	5.0	g
Peptic digest of animal tissue	5.0	g
Dextrose	40	g
Agar	15	g
Olive oil	1%	v/v

Suspend the mixture 65 g in 1 lit of water add 1% v/v of olive oil, and autoclave at 121 °C 15 minute. Cool the sterile media to <50°C before pipette media 20 ml in to sterile plastic disc (10 cm). Dry plates 1 days at room temperature, or 30 min with the lids slightly off at 37°C or in a laminar flow hood. Store dried plates wrapped at 4°C.

APPENDIX C HEMACYTOMETER CALCULATION

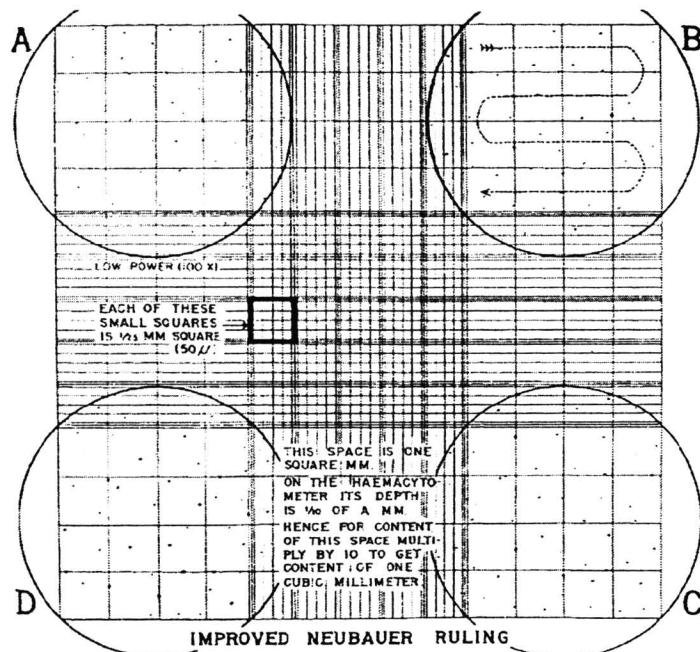


Figure 21 Hemacytometer counting chamber (yeast cells) Areas marked A, B, C, and D are used to count yeast cells

- (1) Cover the coglass on to the counting chamber.
- (2) Place the forefinger over the top (short end) of the pipet, hold the pipet at a 45° angle, and touch the pipet tip to the junction of the cover glass and the counting chamber.
- (3) Allow the yeast suspension to flow under the cover glass until the chamber is completely charged. Similarly, fill the opposite chamber of the hemacytometer.
- (4) Count the yeast cells in the four 1 sq mm corner areas corresponding to those marked A, B, C, and D of Figure 25 in each of two chambers.
- (5) Count all the yeast cells lying within the square and those touching the upper and right-hand center lines. The yeast cells that touch the left-hand and bottom lines are not to be counted. In each of the four areas, conduct the count as indicated by the "snake-like" line in figure 25. A variation of more than 10 cells between any of the four areas counted or a variation of more than 20 cells between sides of the hemacytometer indicate uneven distribution and require that the procedure be repeated.

Calculation

(1) The depth of the counting chamber is 0.1 mm and the area counted is 4 sq mm (4 squares are counted, each with an area of 1.0 sq mm therefore, 4×1.0 sq mm = a total of 4 sq mm). The volume counted is: area \times depth = volume. Four sq mm \times 0.1 mm = 0.4 cu mm.

(2) The formula is as follows:

$$\text{Yeast cells (cells/mL)} = \frac{\text{Average number of chambers (2)yeast counted} \times \text{dilution factor}}{\text{Volume (0.4)}}$$

APPENDIX D

The chemical stability of ethanolic extracts of *A. racemosus*.

The data were expressed as the mean with a standard deviation (SD).

Statistical data were analyzed by paired-sample t test. The significance criterion for the correlation measurements was set at 0.05

The statistical chemical stability of ethanolic extracts of *A. racemosus*.

Paired Samples Statistics

		Mean	N	Std. Deviation	Std. Error Mean
Pair 1	AR3E0Day	6.2210	3	.51260	.29595
	AR3E30Day	5.3075	3	.13406	.07740
Pair 2	AR1E0Day	2.0601	3	.25411	.14671
	AR1E30Day	1.6037	3	.64497	.37238

Paired Samples Correlations

		N	Correlation	Sig.
Pair 1	AR3E0Day & AR3E30Day	3	-.267	.828
Pair 2	AR1E0Day & AR1E30Day	3	-.972	.151

Paired Samples Test

	Paired Differences					t	df	Sig. (2-tailed)			
				95% Confidence Interval of the Difference							
	Mean	Std. Deviation	Std. Error Mean	Lower	Upper						
Pair 1	AR3E0Day - AR3E30Day	.91346	.56341	.32528	-.48613	2.31304	2	.107			
Pair 2	AR1E0Day - AR1E30Day	.45638	.89397	.51614	-1.76437	2.67713	2	.470			

The calculation of percent remaining of *A. racemosus* extract

The percent remaining of *A. racemosus* extract can be calculated by using the equation;

$$\boxed{\text{Percent remaining} = \frac{C_T}{C_I} \times 100}$$

where

C_I is the percent saponin equivalent to shatavarin IV of *A. racemosus* extract before stability test.

C_T is the percent saponin equivalent to shatavarin IV of *A. racemosus* extract after stability test.

If initiation, the % saponin equivalent to shatavarin IV of *A. racemosus* extract at normal condition and % saponin of extract at stress condition was 2.06 mg and 1.60 mg, respectively, therefore, the percent remaining of % saponin in *A. racemosus* extract was calculated as follows:

$$\begin{aligned}\text{Percent remaining} &= \frac{1.60}{2.06} \times 100 \\ &= 77.67\end{aligned}$$

BIOGRAPHY



BIOGRAPHY

Name-Surname	Churanya Onlom
Date of Birth	March 7, 1987
Address	142/1 Wungphikul Subdistrict Wungthong District, Phitsanulok Province 65130
Education Background	
2008	B.S. (Cosmetic sciences), Naresuan University, Phitsanulok.
Publication	
Onlom, C., Khanthawong, S., Waranuch, N. and Ingkaninan, K. (2011). Improvement of the bioactivity of <i>Asparagus racemosus</i> root extract by saponin-enriched extraction method. In Proceedings of the Third International Conference on Natural Products for Health and Beauty (p. 232-236). Bangkok: Rangsit University.	
Onlom, C., Khanthawong, S., Waranuch, N. and Ingkaninan, K. (2011). Evaluation of antifungal activity of <i>Asparagus racemosus</i> Willd. root extracts. In PERCH-CIC CONGRESS VII (p. 235). Bangkok: Center of excellence for innovation in chemistry.	
Petty patent	
Ingkaninan, K., Onlom, C., Waranuch, N., Phrompittayarat, W., Khanthawong, S. (2011). Extraction method for saponin enrichment from <i>Asparagus racemosus</i>. Apply for a Petty Patent, Department of Intellectual Property. (Form PI/PD/PP/001-A) No. 1103000243.	

