

REFERENCES



REFERENCES

- [1] Barton, B. A., Herbert, B., Hauskins, B.L. and Jansen, C. R. (2000). Juvenile pallid (*Scaphirhynchus albus*) and hybrid pallidshovelnose (*S.albusplatorynchus*) sturgeons exhibit low physiological responses to acute handling and severe confinement. **Comparative Biochemistry and Physiology, Part A**, 126, 125–134.
- [2] Barton, B. A. and Iwama, G. K. (1991). Physiological changes in fish from stress in aquaculture with emphasis on the response and effects of corticosteroids. **Annual Review of Fish Diseases**, 10, 3-26.
- [3] Hoskonen, P. and Pirhonen, J. (2006). Effects of repeated handling, with or without anaesthesia, on feed intake and growth in juvenile rainbow trout, *Oncorhynchus mykiss* (Walbaum). **Aquaculture Research**, 37, 409-415.
- [4] Mgbenka, B. O. and Ejiofor, E. N. (1999). Effects of Extracts of Dried Leaves of *Erythrophleum suaveolens* as Anesthetics on Clariid Catfish. **Journal of Applied Aquaculture**, 8(4), 73-80.
- [5] Roubach R., Gomes L.C. and Val A.L. (2001). Safest level of tricainemethanesulfonate (MS-222) to induce anesthesia in *juveniles matrinxá*, *Bryconcephalus*. **Acta Amazonica**, 31, 159-163.
- [6] Issacs, G. (1983). Permanent local anaesthesia and anhidrosis after clove oil spillage. **Lancet**, 1, 882-883.
- [7] Food and Drug Administration (2002). Status of clove oil and eugenol for anesthesia of fish. **Guidance for Industry**, 150, 4.
- [8] Keene, J.L., Noakes, D.L.G., Moccia, R.D. and Soto C.G. (1998). The efficacy of clove oil as an anaesthetic for rainbow trout, *Oncorhynchus mykiss* (Walbaum). **Aquaculture Research**, 29, 89-101.
- [9] Wagner, E., Arndt, R. and Hilton, B. (2002). Physiological stress responses, egg survival and sperm motility of rainbow trout broodstock anesthetized with clove oil, tricaine methanesulfonate or carbon dioxide. **Aquaculture Research**, 21(1-4), 353-366.

- [10] Karapmar, M. and Aktug, S.E. (1987). Inhibition of food borne pathogens by thymal, eugenol, menthol and anethode. **International Journal of Food Microbiology**, 4, 161-166.
- [11] Hussain, M.M.A., Wada, S., Hatai, K. and Yamamoto, A. (2000). Antimycotic activity of eugenol against selected water molds. **Journal of Aquatic Animal Health**, 12 (3), 224-229.
- [12] Mulcahy D. M. (2010). L. G. Ross and B. Ross: Anaesthetic and sedative techniques for aquatic animals (Third Edition). **Reviews in Fish Biology and Fisheries**, 20, 139–140.
- [13] Ah-Lim, T., Graham, P. and Richard, J. K. (1992). Prostaglandin H Synthase kinetics of tyrosyl radical formation and of cyclooxygenase catalysis. **The Journal of Biological Chemistry**, 267, 17753-17759 (5).
- [14] Dewhirst, F.E. and Goodson, J.M. (1974). Prostaglandin synthase inhibition by eugenol, guaiacol and other dental medicament. **Journal of Dental Research**, 53, 104.
- [15] Thompson, D. and Eling, T. (1989). Mechanism of inhibition of prostaglandin H synthase by eugenol and other phenolic peroxidase substrates. **Molecular Pharmacology**, 36, 809-817.
- [16] Pongprayoon, U., Baekstrom, P., Jacobsson, U., Lindstrom, M. and Bohlin, L. (1991). Compounds inhibiting prostaglandin synthesis isolated from *Ipomoea pes-caprae*. **Planta Medical**, 57, 515-518.
- [17] Hunn, J.B. and Allen, J.L. (1974). Movement of drugs across the gills of fishes. **Annual Review of Pharmacology**, 14, 47-55.
- [18] Schoettger, R. A. and Julin, A. M. (1967). Efficacy of MS-222 as an anesthetic on four salmonids. U.S. Fish and Wildlife Service. **Investigations in fish control**, 13, 1-15.
- [19] Mattson, N. S. and Riple, T. H. (1989). Metomidate, a better anesthetic for cod (*Gadus morhua*) in comparison with benzocaine, MS-222, chlorobutanol and phenoxyethanol. **Aquaculture**, 83, 89-94.
- [20] Bolasina, S. N. (2006). Cortisol and hematological response in Brazilian codling, *Urophycis brasiliensis* (Pisces, Phycidae) subjected to anesthetic treatment. **Aquacult International**, 14, 569–575.

- [21] Keene, J.L., Noakes, D.L.G., Moccia, R.D., Soto, C.G. (1998). The efficacy of clove oil as an anaesthetic for rainbow trout, *Oncorhynchus mykiss* (Walbaum). **Aquaculture Research**, 29, 89-101.
- [22] Fernandes, M. N. and Mazon, A. F. (2003). Environmental pollution and fish gill morphology. In: Val, A.L. and Kapoor, B.G. (Eds.), *Fish Adaptations*, **Science Publishers**, 9, 203-231.
- [23] Mcknight, I. M. A. (1966). Hematological study on the mountain white fish. **Popium Willasemi South**, 23, 45-64.
- [24] Sudagara, M., Mohammadizarejabada, A., Mazandarania, R. and Pooralimotlagha, S. (2009). The efficacy of clove powder as an anesthetic and its effects on hematological parameters on Roach (*Rutilus rutilus*). **Medwell Publishing**, 1, 1-5 (1).
- [25] Pirhonen, J. and Schreck, C.B. (2003). Effects of anaesthesia with MS-222, clove oil and CO₂ on feed intake and plasma cortisol in steelhead trout (*Oncorhynchus mykiss*). **Aquaculture Research**, 220, 507-514.
- [26] Kildea, M. A., Allan, G. L. and Kearney, R. E. (2004). Accumulation and clearance of the anaesthetic clove oil and AQUI-STM from the edible tissue of silver perch (*Bidyanus bidyanus*). **Aquaculture**, 232, 256-277.
- [27] Hussain, M.M.A., Wada, S., Hatai, K. and Yamamoto A. (2000). Antimycotic activity of eugenol against selected water molds. **Journal of Aquatic Animal Health**, 12(3), 224-229.
- [28] Zaikov, A., Iliev, I. and Hubenova, T. (2008). Induction and recovery from anaesthesia in Pike (*Esox Lucius L.*) exposed to clove oil. **Bulgarian Journal of Agricultural Science**, 14(2), 165-170.
- [29] Sandorfy, C. (2004). Hydrogen bonding and anaesthesia. **Journal of Molecular Structure**, 708, 3-5.
- [30] Sonesson, C., Lin, C. H., Hansson, L., Waters, N., Svensson, K., Carlsson, A., Smith, M. W. and Wikstron, H. (1994). Substituted (S)-phenylpiperidines and rigid congeners as preferential dopamine autoreceptor antagonists: synthesis and structure-activity relationships. **Journal Medicinal Chemistry**, 37, 2735-2753.

- [31] Bolton, J. L., Comeau, E. and Vukonmanovic, V. (1995). The influence of 4-alkyl substituents on the formation and reactivity of 2-methoxy-quinone methides: evidence that extended conjugation dramatically stabilizes the quinone methide formed from eugenol. **Chemico-Biological Interactions**, 95, 279-290.
- [32] Cheng, X., Prehm, M., Das, K. M., Kain, J., Baumeister, U., Diele, S., Leine, D., Blume, A. and Tschierske, C. (2003). Calamitic Bolaamphiphiles with (Semi) Perfluorinated Lateral Chains: Polyphilic Block Molecules with New Liquid Crystalline Phase Structures. **Journal of the American Chemical Society**, 125, 10977-10996.
- [33] Someya, H., Ohmiya, H., Yorimitsu, H. and Oshima, K. (2008). Silver-catalyzed benzylolation and allylation reaction of tertiary and secondary alkyl halides with Grignard reagents. **Organic Letters**, 10, 969-971.
- [34] Ricci, J., Poulain-Martini, S. and Dunach, E. (2009). Catalytic Friedel-Crafts allylation using Zn(II) triflimidate. *Comptes Rendus Chimie*, **Published by the French Academy of Sciences**, 12, 916-921.
- [35] Fang, Z., Zhou, G. C., Zheng, S. L., He, G. L., Li, J. L. and Bei, D. (2007). Lithium chloride-catalyzed selective demethylation of aryl methyl ethers under microwave irradiation. **Journal of Molecular Catalysis A: Chemical**, 274, 16-23.
- [36] Zuo, L., Yao, S., Wang, W. and Duan, W. (2008). An efficient method for demethylation of aryl methyl ethers. **Tetrahedron letters**, 49, 4054-4056.
- [37] Augusto, J., Rodrigues, R., Pedro de Oliveira Filho, A. and Paulo Moran, J. S. (1999). Regioselectivity of the nitration of phenol by acetyl nitrate adsorbed on silica gel. **Tetrahedron**, 55, 6733-6738.
- [38] Kwak, J. H., In, J. K., Lee, M. S., Choi, E. H., Lee, H., Hong, J. T., Yun, Y. P., Lee, S. J., Seo, S. Y., Suh, Y. G. and Jung, J. K. (2008). Concise synthesis of obovatol: Chemoselective ortho-bromination of phenol and survey of Cu-catalyzed diaryl ether couplings. **Archives of Pharmacal Research**, 31(12), 1559-1563.
- [39] Chen, C. M. and Liu, Y. C. (2009). A concise synthesis of honokiol. **Tetrahedron Letters**, 50, 1151-1152.

- [40] Bu, X., Jing, H., Wang, L., Chang, T., Jin, L. and Liang, Y. (2006). Organic base catalyzed O-alkylation of phenols under solvent-free condition. **Journal of Molecular Catalysis A: Chemical**, 259, 121–124.
- [41] Mizuno, M. and Yamano, M. (2005). A new practical one-pot conversion of phenols to anilines. *Organic Letters*, 7 (17), 3629-3631.
- [42] Allen, C. F. H. and Gates, J. W. (1955). **Organic Syntheses**, 3 (25), 418-449.
- [43] Yadav, J. S., Reddy, S. B. V., Borkar, P. and Reddy, J. (2009). Addition of aryl cuprates to azides: a novel approach for the synthesis of unsymmetrical diaryl amines. **Tetrahedron letters**, 50, 6642-6645.
- [44] Banno, T., Hayakawa, Y. and Umeno, M. (2002). Some application of the Grignard cross-coupling reaction in the industrial field. **Organo metallic chemistry**, 653, 288-291.
- [45] Nagano, T. and Hayashi, T. (2005). Iron-catalyzed oxidative homo-coupling of aryl grignard reagents. **Organic Letters**, 7 (3), 491-493.
- [46] Cahiez, G., Chaboche, C., Duplais, C. and Moyeux, A. (2009). A new efficient catalytic system for the chemoselective cobalt-catalyzed cross-coupling of aryl Grignard reagents with primary and secondary alkyl bromides. **Organic Letters**, 11 (2), 277-280.
- [47] Wilhelm, H. and Wessjohann, L. A. (2006). An efficient synthesis of the phytoestrogen 8-prenylnaringenin from xanthohumol by a novel demethylation process. **Tetrahedron**, 62, 6961-6966.
- [48] Bernini, R., Barontini, M., Mosesso, P., Pepe, G., Willfor, S. M., Sjöholm, R. E., Eklundb, P. C. and Saladino, R. (2009). A selective de-O-methylation of guaiacyl lignans to corresponding catechol derivatives by 2-iodoxybenzoic acid (IBX). The role of the catechol moiety on the toxicity of lignans. **Organic and Biomolecular Chemistry**, 7, 2367–2377.
- [49] Jin-mei, Z., Yue-qing, L. and Wei-jie, Z. Demethylating Reaction of Methyl Ethers. **The Proceedings of the 3rd International Conference on Functional Molecules**. China: State Key Laboratory of Fine Chemicals, Dalian University of Technology, Dalian 116012.

- [50] Kwak, J. H., Cho, Y. A., Jang, J. Y., Seo, S. Y., Lee, H., Hong, J. T., Han, S. B., Lee, K., Kwak, Y. S. and Jung, J. K. (2011). Expedient synthesis of 4-*O*-methylhonokiol *via* Suzuki-Miyaura cross-coupling. **Tetrahedron**, 67(48), 1-4.
- [51] Zuo, L., Yao, S., Wang, W. and Duan, W. (2008). An efficient method for demethylation of aryl methyl ethers. **Tetrahedron letters**, 49, 4054-4056.
- [52] Mizuno, M. and Yamano, M. (2005). A new practical one-pot conversion of phenols to anilines. **Organic Letters**, 7(17), 3629-3631.
- [53] Fujisaki, S., Eguchi, H. and Omura, A. (1993). Regioselective bromination of phenol. **Organic Chemistry**, 66, 6-16.
- [54] Bovonsombat, P., Ali, R. and Khan, C. (2010). Facile *p*-toluenesulfonic acid-promoted para-selective monobromination and chlorination of phenol and analogues. **Tetrahedron**, 66, 6928-6935.
- [55] Majetich, G., Hicks, R. and Reister, S. (1997). Electrophilic aromatic bromination using bromodimethylsulfonium bromide generated in situ. **Organic Chemistry**, 62, 4321-4326.
- [56] Courtney, A. (2004). CH 334 Organic Chemistry, **Lecture Notes on the Hydrogenation of Alkenes**. Retrieved April 2, 2012 from <http://www.wou.edu/las/physci/ch334/lecture/lect16.htm>.
- [57] Lew, J. (2011). ChemWiki: The Dynamic Chemistry Textbook. **Catalytic Hydrogenation of Alkenes**. Retrieved April 1, 2012 from http://chemwiki.ucdavis.edu/Organic_Chemistry/Hydrocarbons/Alkenes/Catalytic_Hydrogenation.
- [58] Kwak, J. H., In, J. K., Lee, M. S., Choi, E., H., , Lee, H., Hong, J. T., Yun, Y. P., Lee, S. J., Seo, S. Y., Suh, Y. G. and Jung, J. K. (2008). Concise synthesis of obovatol: chemoselective ortho-bromination of phenol and survey of Cu-catalyzed diaryl ether couplings. **Archives of Pharmacal Research** 31 (12), 1559-1563.
- [59] Dwyer, C. L. and Holzapfel, C. W. (1998). The nitration of electron-rich aromatics. **Tetrahedron**, 54, 7843-7848.

- [60] Fujikawa, N., Ohta, T., Yamaguchi, T., Fukuda, T., Ishibashi, F. and Iwao, M. (2006). Total synthesis of lamellarins D, L, and N. **Tetrahedron**, 62(4), 594-604.
- [61] David, A. L., Vieira-Coelho, M. A., Benes, J., Alves, P. C., Borges, N., Freitas, A. P. and Soares-da-Silva, P. (2002). Synthesis of 1-(3,4-Dihydroxy-5-nitrophenyl)-2-phenyl-ethanone and derivatives as potent and long-acting peripheral Inhibitors of catechol-*O*-methyltransferase. **American Chemical Society**, 45, 685-695.

APPENDIX

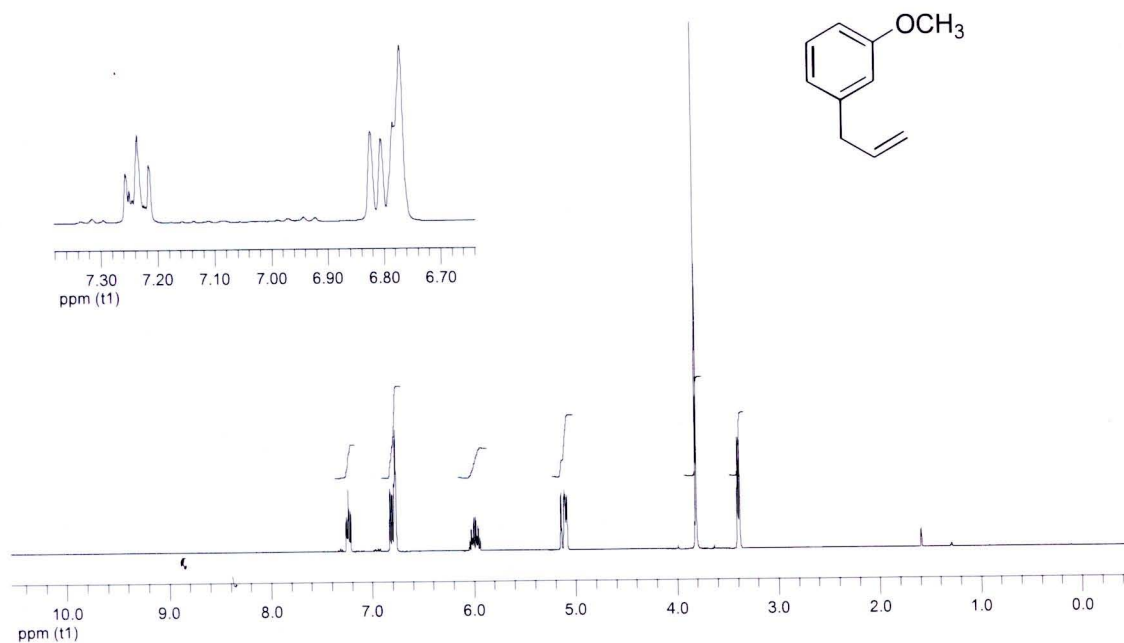


Figure 62 ^1H NMR spectrum of 3-allylanisole (1)

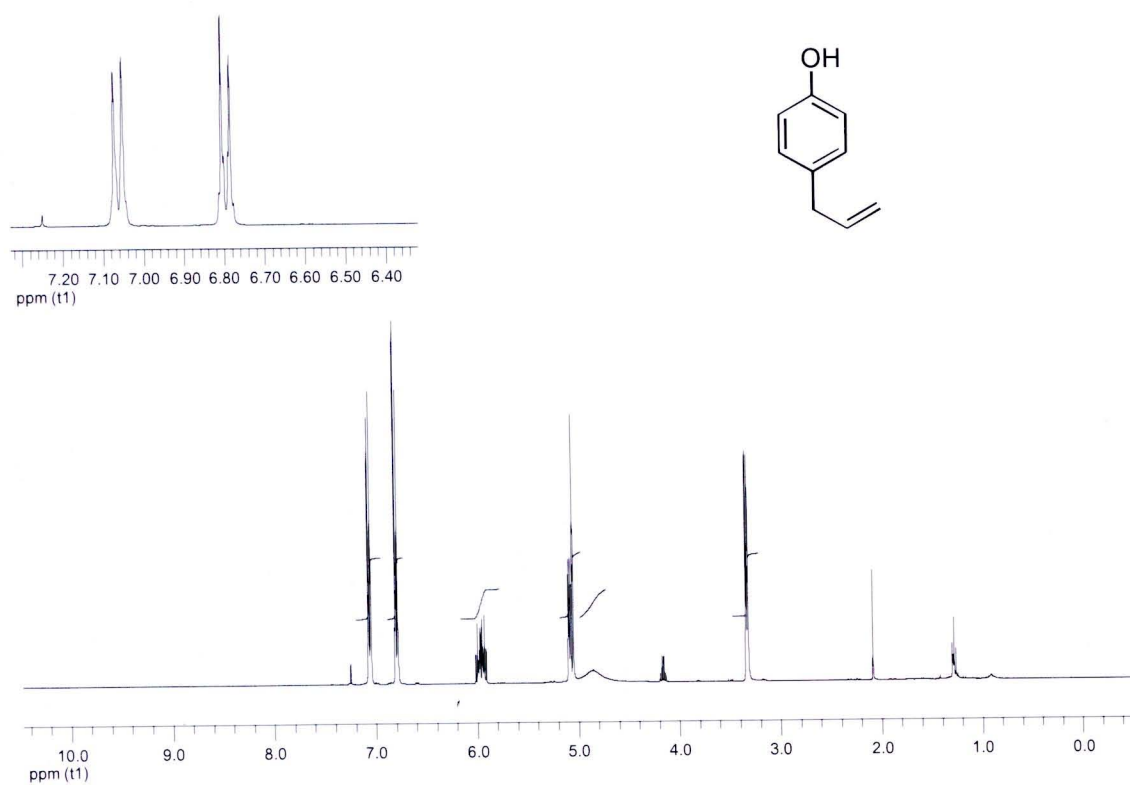


Figure 63 ^1H NMR spectrum of 4-allylphenol (2)

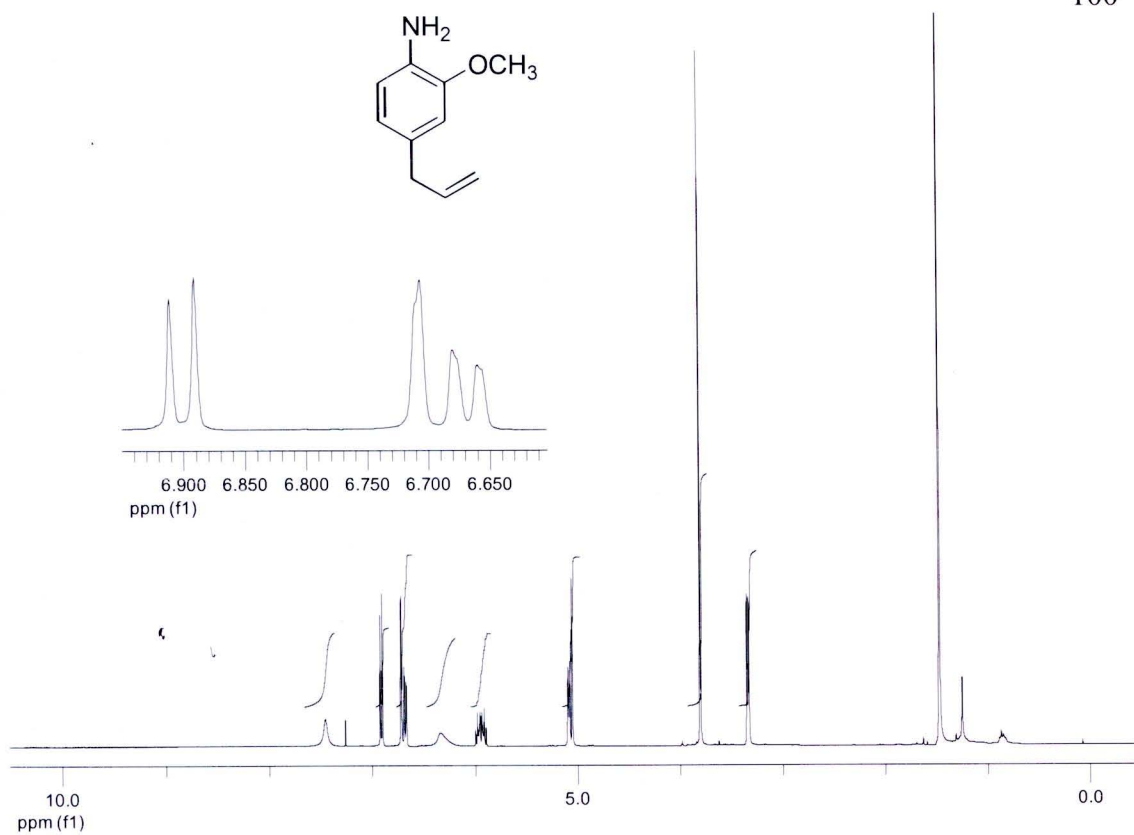


Figure 64 ^1H NMR spectrum of 4-allyl-2-methoxybenzenamine (4)

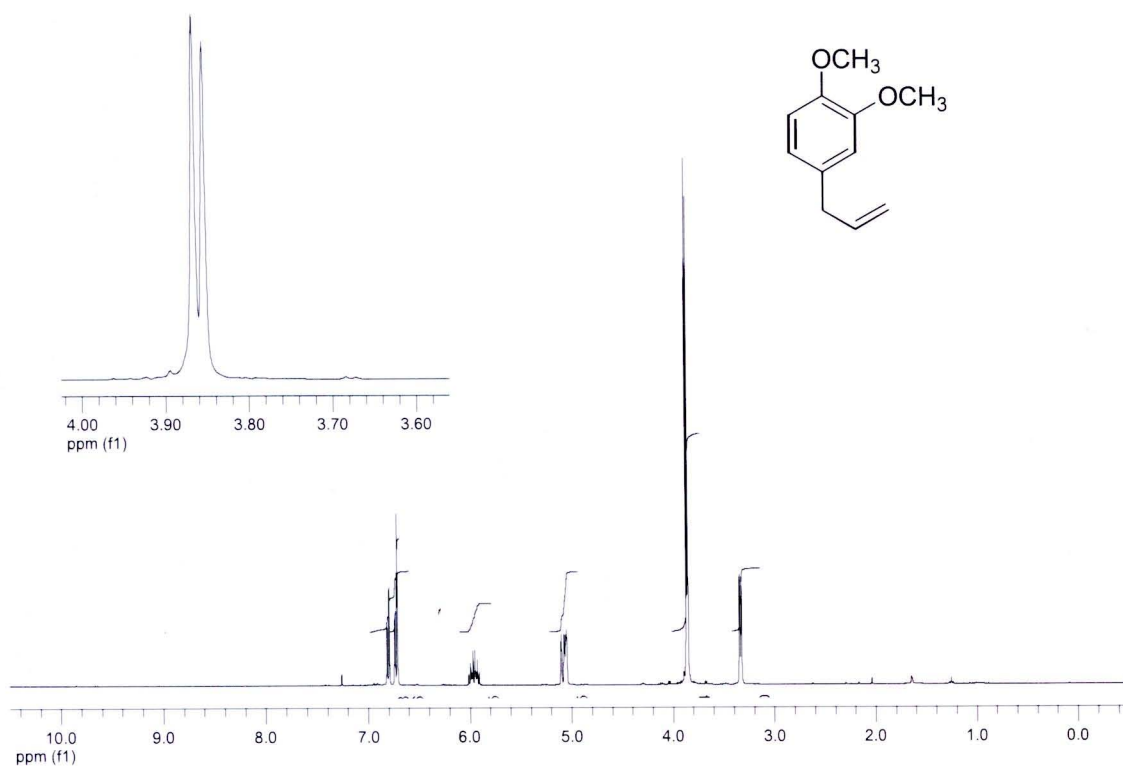


Figure 65 ^1H NMR spectrum of methyleugenol (5)

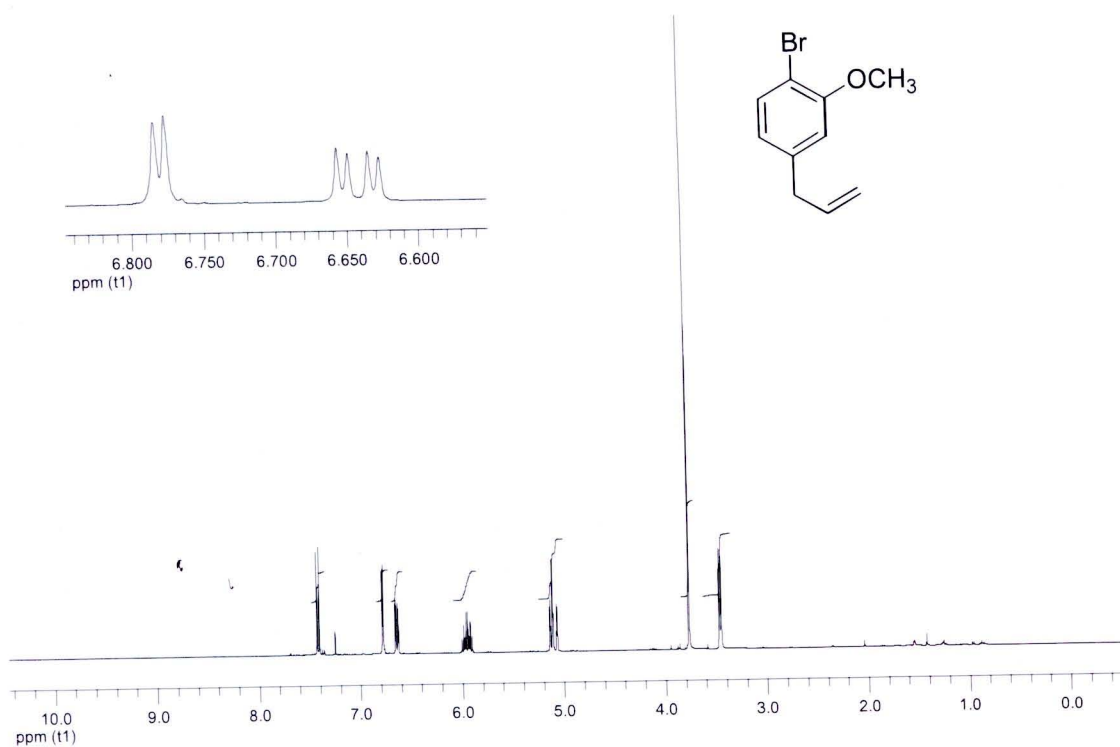


Figure 66 ^1H NMR spectrum of 4-allyl-1-bromo-2-methoxybenzene (6)

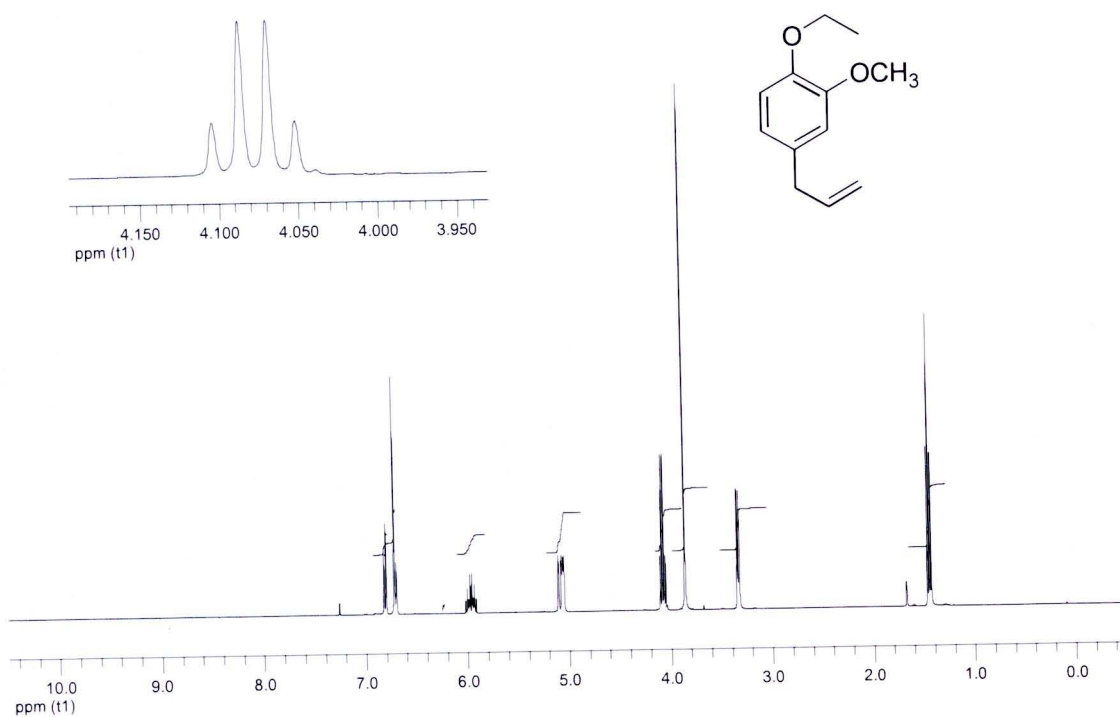


Figure 67 ^1H NMR spectrum of ethyleugenol (7)

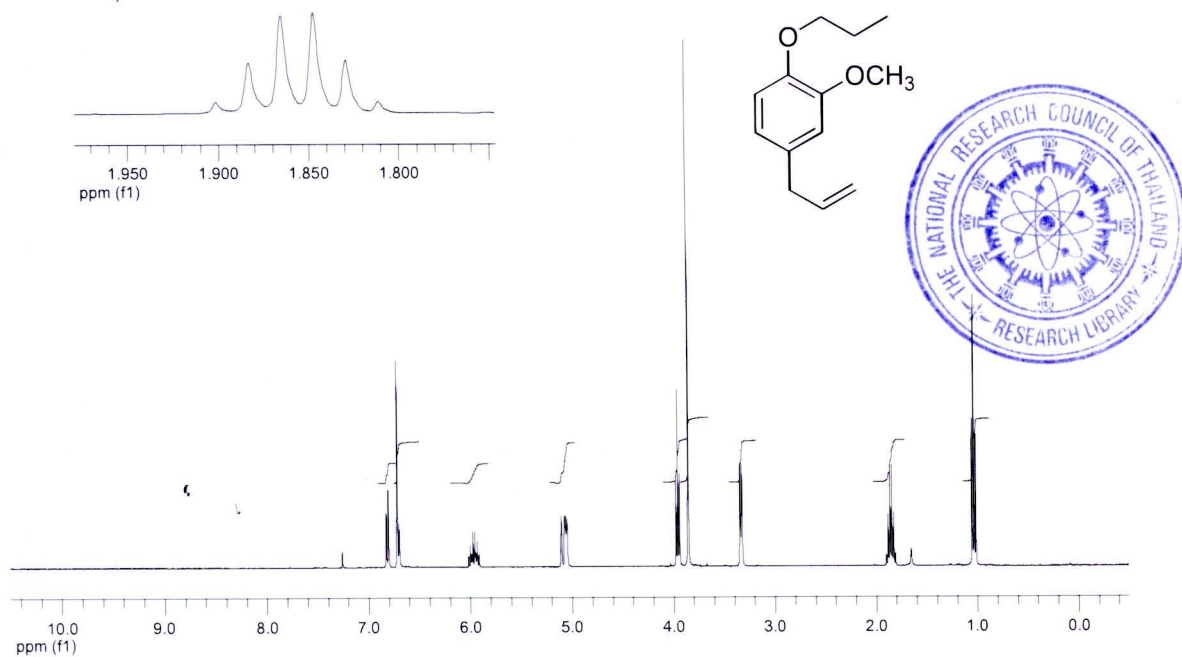


Figure 68 ^1H NMR spectrum of propyleugenol (8)

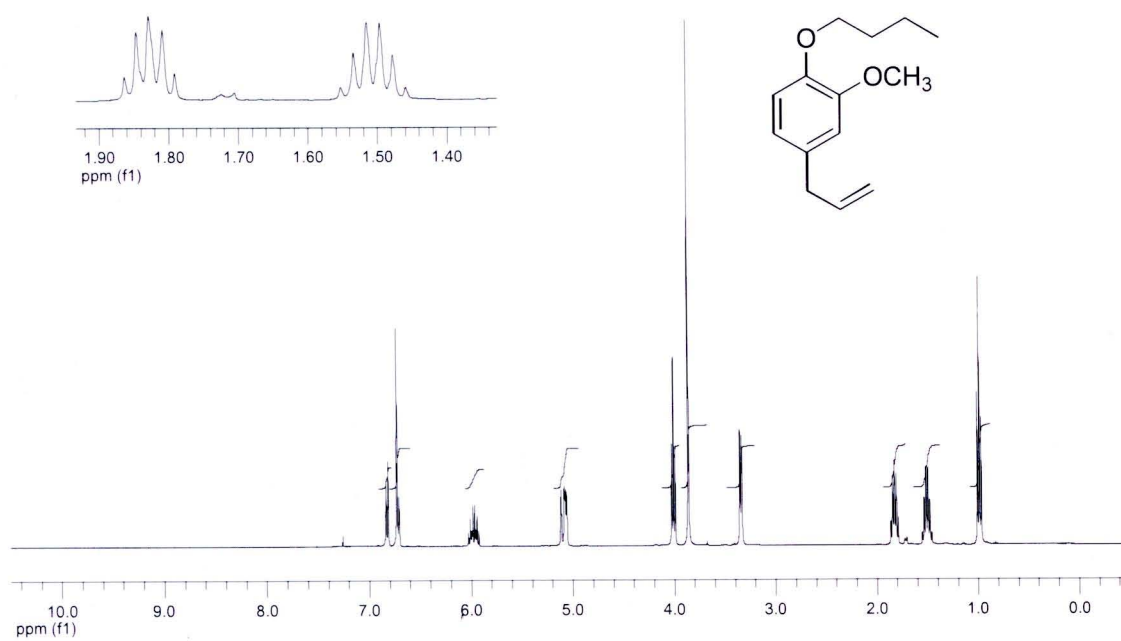


Figure 69 ^1H NMR spectrum of butyleugenol (9)

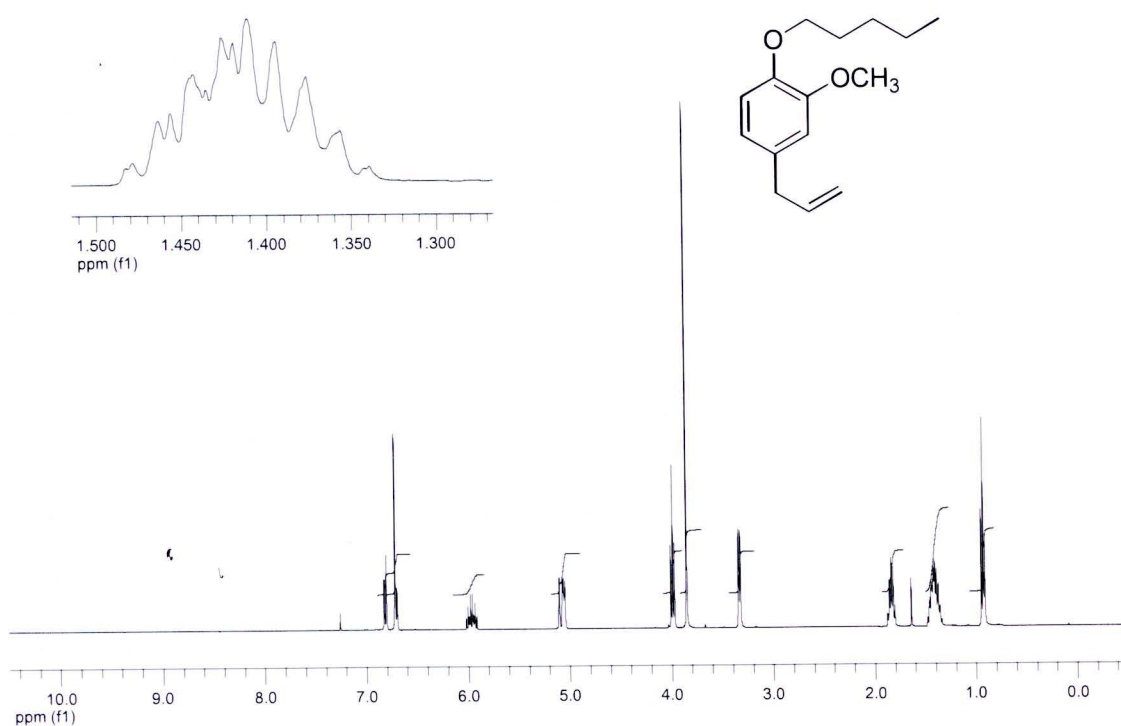


Figure 70 ^1H NMR spectrum of pentyleugenol (10)

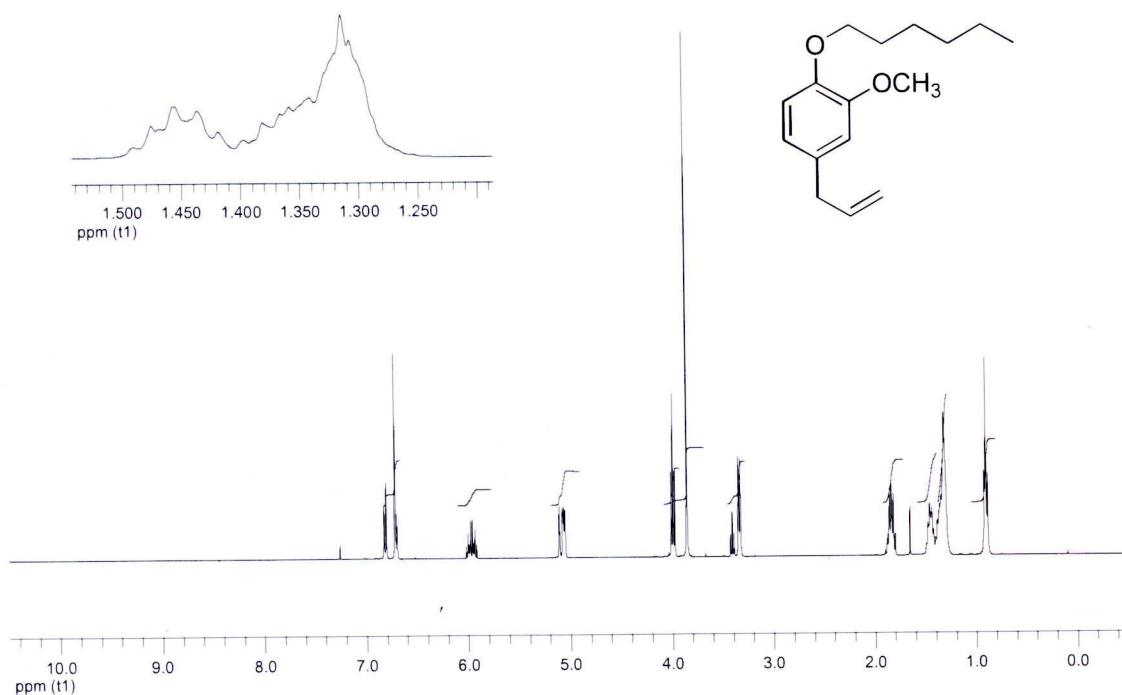


Figure 71 ^1H NMR spectrum of hexyleugenol (11)

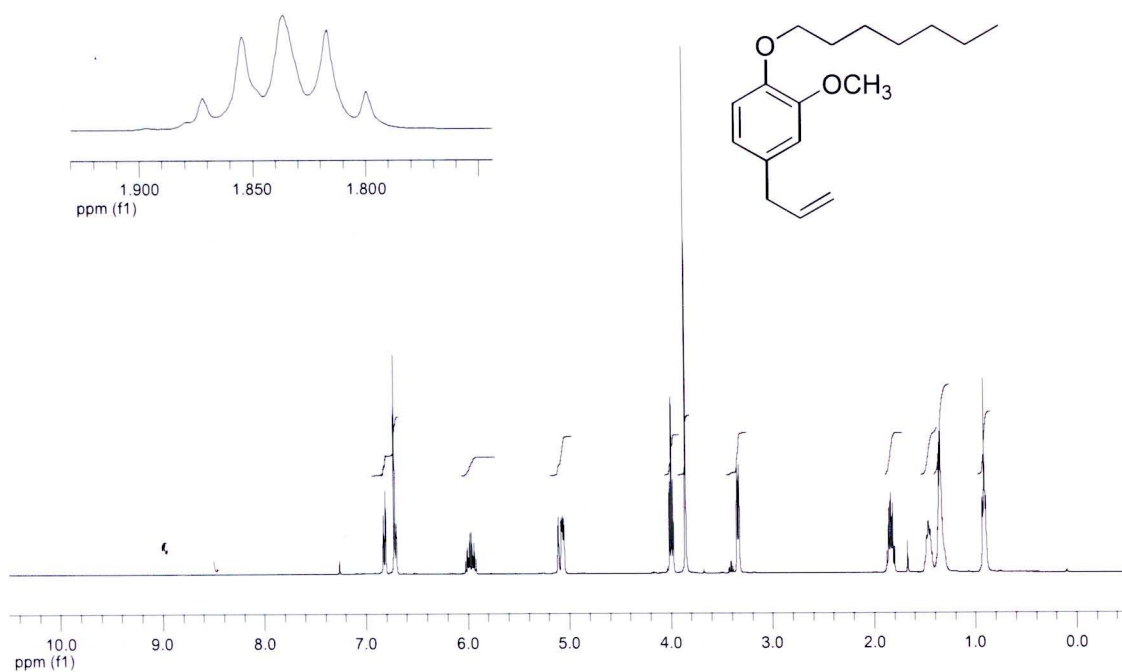


Figure 72 ^1H NMR spectrum of heptyleugenol (12)

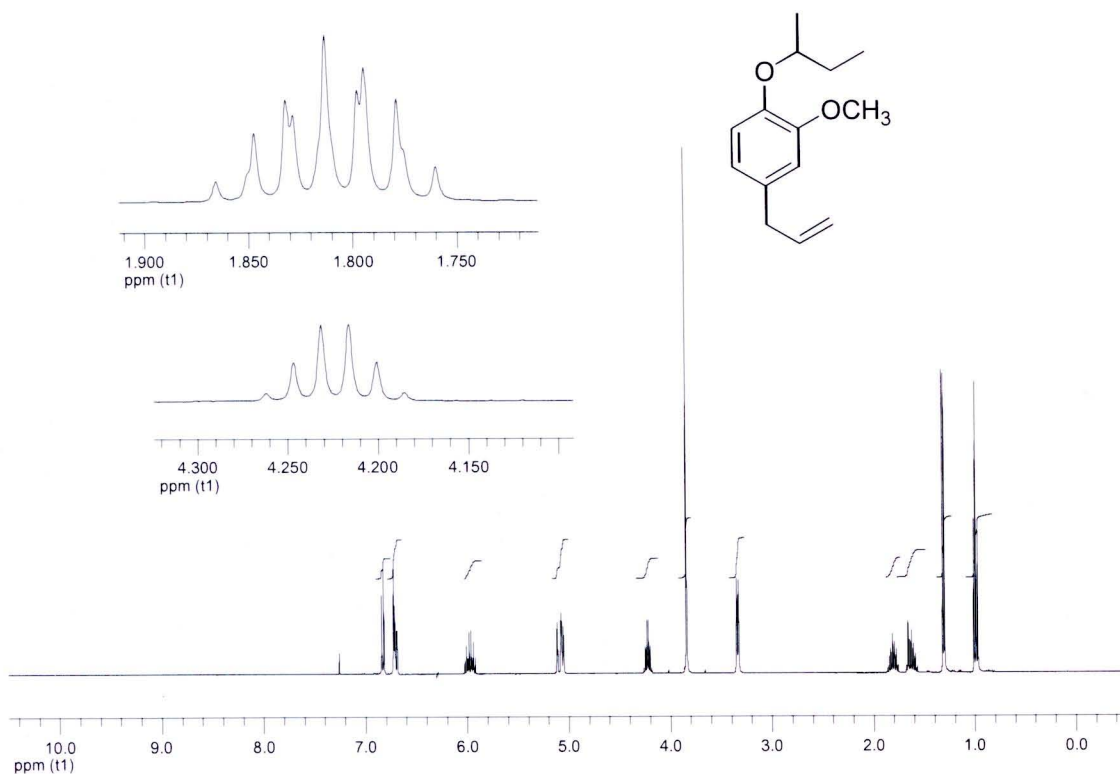


Figure 73 ^1H NMR spectrum of *sec*-butyleugenol (13)

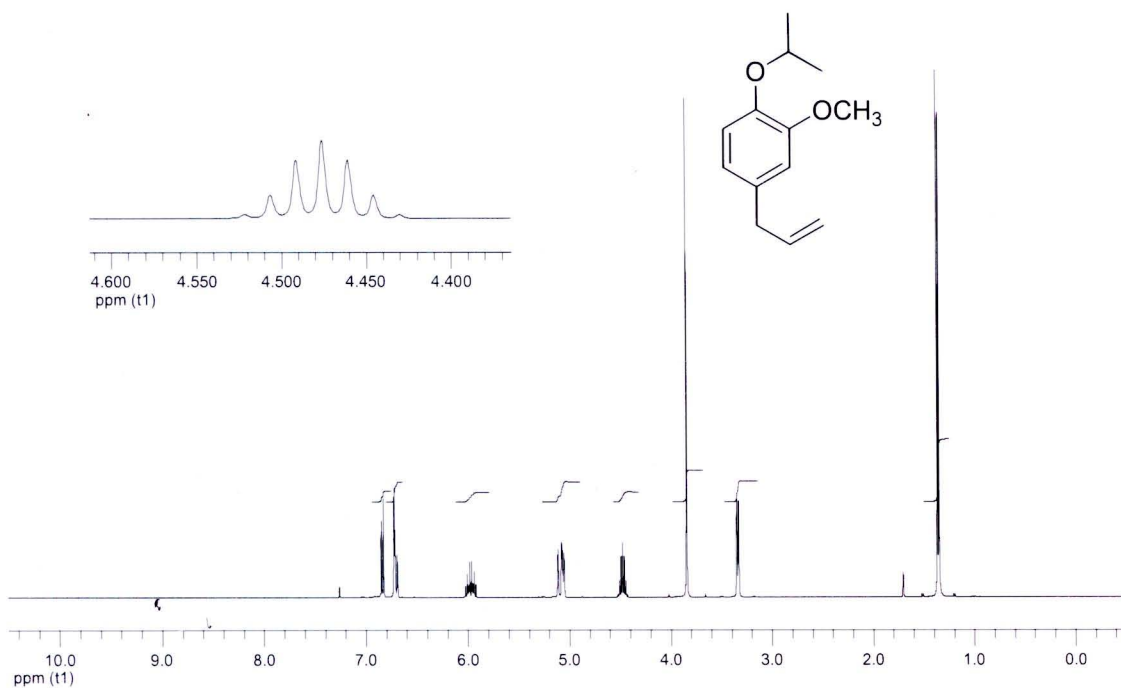


Figure 74 ^1H NMR spectrum of *iso*-propyleugenol (14)

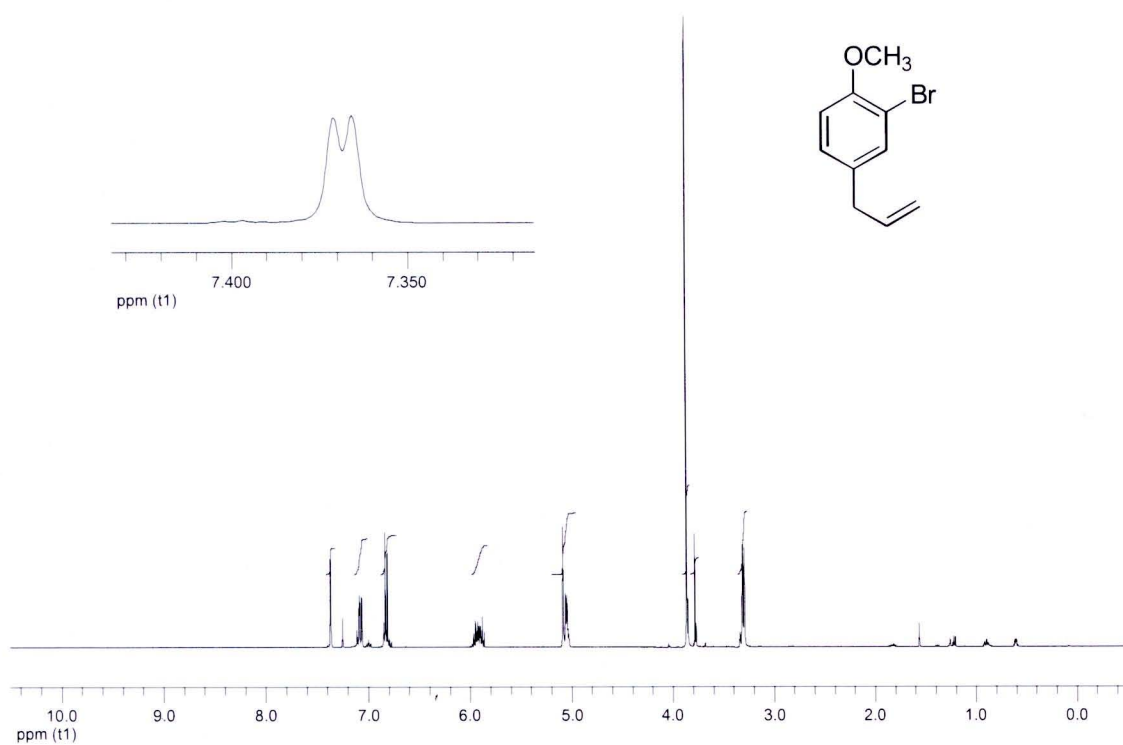


Figure 75 ^1H NMR spectrum of 4-allyl-2-bromo-1-methoxybenzene (15)

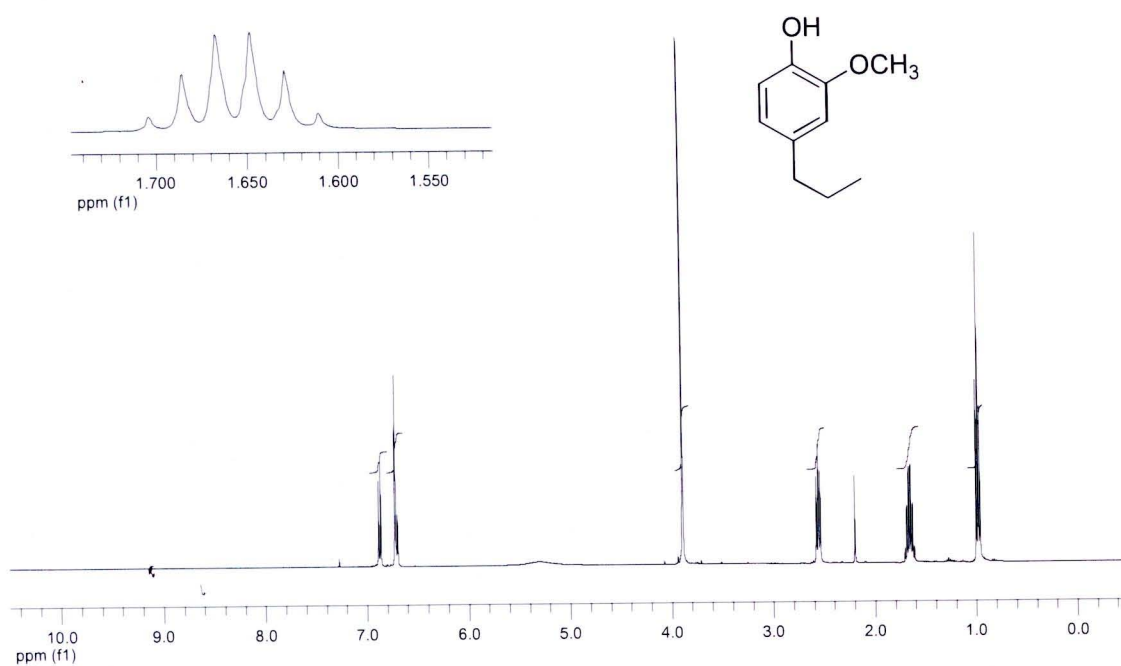


Figure 76 ^1H NMR spectrum of 2-methoxy-4-propylphenol (PMP) (17)

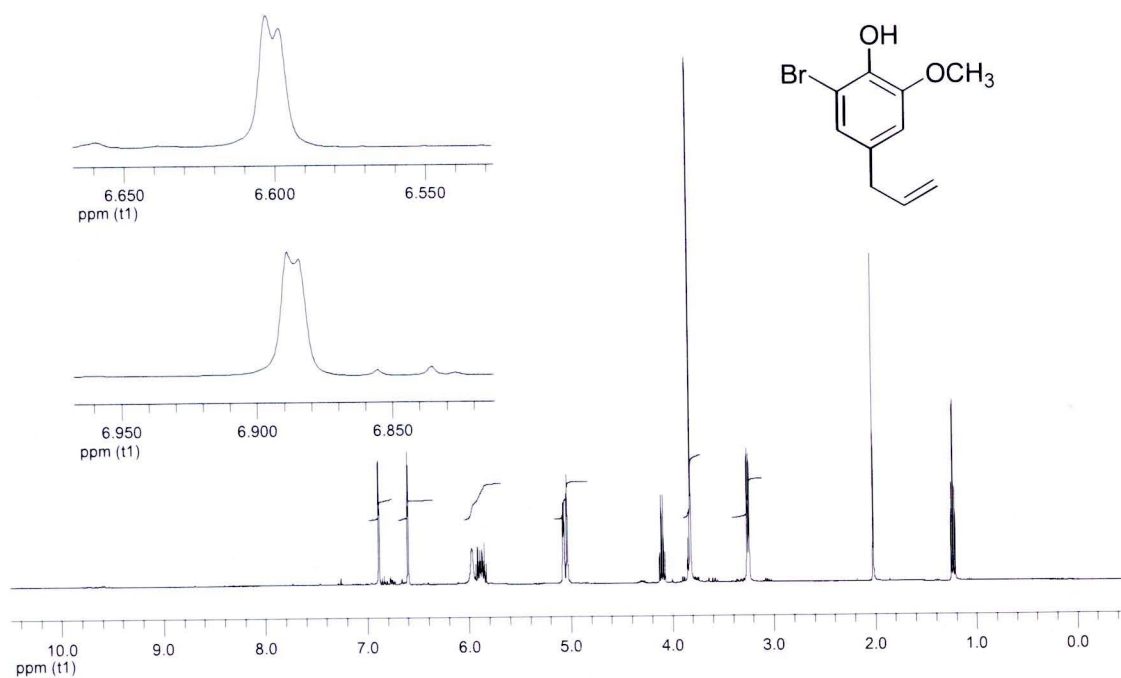


Figure 77 ^1H NMR spectrum of bromoeugenol (18)

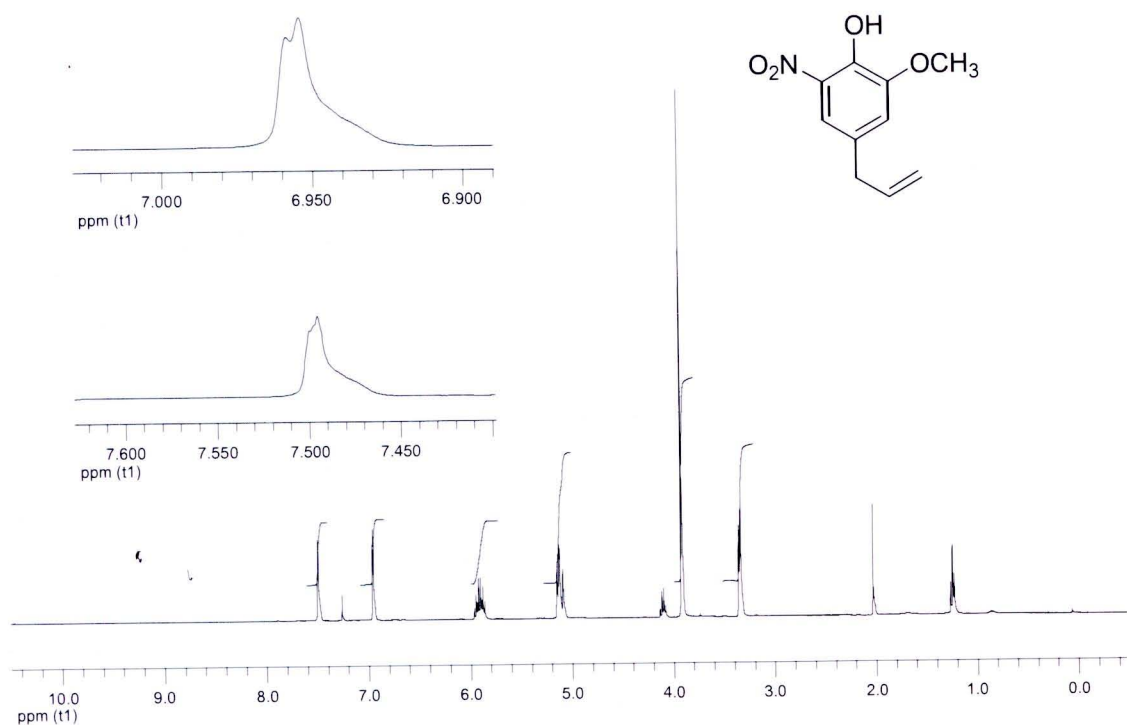


Figure 78 ^1H NMR spectrum of nitro Eugenol (19)

Table 20 Introduced time, recovery time and survival rate in post larvae

L. vannamei

Compound (derivative)	Concentration (ppm)	Introduce time (min)	Recovery time (min)	Survival rate (%)
1	25	-	-	100
	50	2:51	4:00	93.3
2	25	11:26	4:16	90.0
	50	5:03	3:56	96.6
3	25	-	-	100
	50	-	-	100
4	25	-	-	100
	50	-	-	100
5	25	-	-	100
	50	16:40	3:00	96.6
6	25	7:44	7:54	80.0
	50	6:40	28:25	80.3
7	25	-	-	100
	50	16:04	8:19	96.6
8	25	7:59	4:23	100
	50	4:37	3:24	100
9	25	17:29	12:07	90.0
	50	7:27	2:18	100
10	25	17:53	11:01	93.3
	50	13:06	8:23	90.0
11	25	-	-	100
	50	-	-	100
12	25	39:43	18:31	83.3
	50	28:24	8:39	96.6
13	25	26:04	19:55	96.6
	50	11:17	6:34	90.0

Table 20 (Cont.)

Compound (derivative)	Concentration (ppm)	Introduce time (min)	Recovery time (min)	Survival rate (%)
14	25	16:56	0:51	100
	50	7:15	4:18	100
15	25	6:18	7:34	100
	50	4:45	6:59	100
16	25	11:57	3:53	86.6
	50	3:44	6:13	96.6
17	25	7:20	6:45	100
	50	2:53	5:59	100
18	25	32:13	7:27	100
	50	27:19	6:47	93.3
19	25	-	-	100
	50	-	-	100
20	25	3:37	4:12	86.6
	50	3:19	5:30	100

Table 21 Introduced time, recovery time and survival rate adult *L. vannamei*

Compound (derivative)	Concentration (ppm)	Introduce time (min)	Recovery time (min)	Survival rate (%)
1	25	3:27	8:02	100
	50	1:30	4:37	93.3
2	25	-	-	90.0
	50	8:06	2:30	96.6
3	25	-	-	100
	50	-	-	100
4	25	-	-	100
	50	-	-	100
5	25	17:35	36:03	80.0
	50	11:36	35:25	100
6	25	3:08	5:06	83.3
	50	2:20	4:14	75.0
7	25	6:11	6:11	90.0
	50	5:30	35:25	80.0
8	25	4:27	38:58	100
	50	4:36	40:24	90.0
9	25	4:31	13:55	30.0
	50	4:21	11:31	100
10	25	5:23	10:31	20.0
	50	4:34	10:39	40.0
11	25	6:34	13:32	100
	50	5:34	14:26	10.0
12	25	7:26	11:44	80.0
	50	6:23	12:03	50.0
13	25	5:22	10:34	80.0
	50	4:12	12:28	80.0

Table 21 (Cont.)

Compound (derivative)	Concentration (ppm)	Introduce time (min)	Recovery time (min)	Survival rate (%)
14	25	5:19	37:03	100
	50	8:31	8:09	10.0
15	25	-	-	-
	50	-	-	-
16	25	-	-	100
	50	18:20	6:09	100
17	25	10:16	30:25	80.0
	50	11:11	25:39	80.0
18	25	-	-	100
	50	-	-	93.3
19	25	-	-	100
	50	-	-	100
20	25	17:02	4:14	100
	50	6:04	4:17	100

Table 22 Introduced time, recovery time and survival rate in post larvae
L. calcarifer

Compound (derivative)	Concentration (ppm)	Introduce time (min)	Recovery time (min)	Survival rate (%)
1	25	2:41	0:24	100
	50	1:20	0:10	100
2	25	3:38	0:38	60.0
	50	2:42	0	0
3	25	-	-	100
	50	-	-	100
4	25	-	-	100
	50	6:23	2:43	93.3
5	25	1:16	5:23	93.3
	50	0:44	6:40	96.6
6	25	8:01	8:05	13.3
	50	5:20	3:15	13.3
7	25	6:05	7:59	96.6
	50	2:56	9:11	93.3
8	25	3:12	14:07	96.6
	50	3:00	6:29	50.0
9	25	18:51	3:50	73.3
	50	10:29	1:32	46.6
10	25	13:28	0	0
	50	8:39	0	0
11	25	-	-	100
	50	-	-	100
12	25	-	-	100
	50	-	-	100
13	25	8:05	0	0
	50	6:28	0	0

Table 22 (Cont.)

Compound (derivative)	Concentration (ppm)	Introduce time (min)	Recovery time (min)	Survival rate (%)
14	25	12:48	15:33	100
	50	6:39	17:24	80.0
15	25	10:49	51:09	20.0
	50	5:20	34:43	26.6
16	25	0:55	2:47	100
	50	0:35	3:45	100
17	25	8:48	6:20	96.6
	50	4:38	6:29	100
18	25	10:41	2:16	100
	50	3:32	2:31	100
19	25	-	-	100
	50	-	-	100
20	25	1:07	4:07	100
	50	0:36	3:34	100

Table 23 Introduced time, recovery time and survival rate in post larvae*C. macrocephalus*

Compound (derivative)	Concentration (ppm)	Introduce time (min)	Recovery time (min)	Survival rate (%)
5	25	3:13	29:17	100
	50	3:05	34:40	94
7	25	8:15	10:17	96
	50	7:08	10:50	86
8	25	3:06	37:17	100
	50	2:10	43:50	100
9	25	3:13	9:17	92
	50	3:01	10:00	94
10	25	9:00	7:33	95
	50	8:11	10:16	70
11	25	8:11	10:33	100
	50	7:10	10:50	90
12	25	8:01	12:50	90
	50	7:10	13:20	90
13	25	4:11	9:50	100
	50	4:08	11:00	86
14	25	10:11	4:67	100
	50	8:10	9:50	90
17	25	12:10	9:00	86
	50	9:13	12:34	70
20	25	9:08	14:23	86
	50	7:17	17:16	75

Table 24 Introduced time, recovery time and survival rate adult *C.macrocephalus*

Compound (derivative)	Concentration (ppm)	Introduce time (min)	Recovery time (min)	Survival rate (%)
5	25	4:50	25:10	100
	50	4:30	32:20	100
7	25	10:10	10:20	90
	50	7:20	10:50	86
8	25	4:27	31:00	100
	50	3:20	40:50	100
9	25	4:50	7:10	92
	50	4:30	5:40	88
10	25	8:30	7:40	80
	50	7:50	9:10	92
11	25	8:33	8:10	100
	50	7:17	7:00	80
12	25	9:50	12:20	90
	50	7:34	12:10	86
13	25	5:20	6:40	100
	50	4:20	7:50	90
14	25	12:20	5:60	90
	50	9:20	7:50	90
17	25	13:40	9:30	80
	50	11:17	12:20	70
20	25	10:20	10:30	81
	50	8:34	12:40	70

BIOGRAPHY

BIOGRAPHY

Name-Surname

Thitiphong Khamkhen

Date of Birth

July 15, 1986

Address

26/3 Village No. 9, Huai Ruam sub-district, Nongbua
district, Nakhon Sawan 60110 Thailand

Education Background

2008

B.S. (Chemistry), University of Phayao, Thailand



