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APPENDICES

APPENDIX A

Determination of %DQ of *N,N,N*-trimethylammonium chitosan chloride (TMC)

The %DQ of TMCs were determined from the relative ratios between the signal at 3.10 ppm can be assigned to three methyl groups of quaternary ammonium group and the signal of H-1 (δ 4.38-5.34 ppm), H-2 (δ 2.95 ppm), H-2',3,4,5,6,6' (δ 3.25-4.30 ppm), H-2,2',3,4,5,6,6' (δ 2.95 ppm and 3.25-4.30 ppm), or $-NHCOCH_3$ (δ 1.84 ppm) of chitosan analyzed by 1H NMR. The position of those signals in the TMC spectra is shown in Figure A-1. %DQ, %DS_{*N(CH*₃)₂, %DS_{*NHCH*₃, and %DS (total) are quaternization, *N,N*-dimethylation, *N*-methylation, and a total degree of *N*-methylation of chitosan were determined from 1H NMR as shown in Tables A-1 to A-5.}}

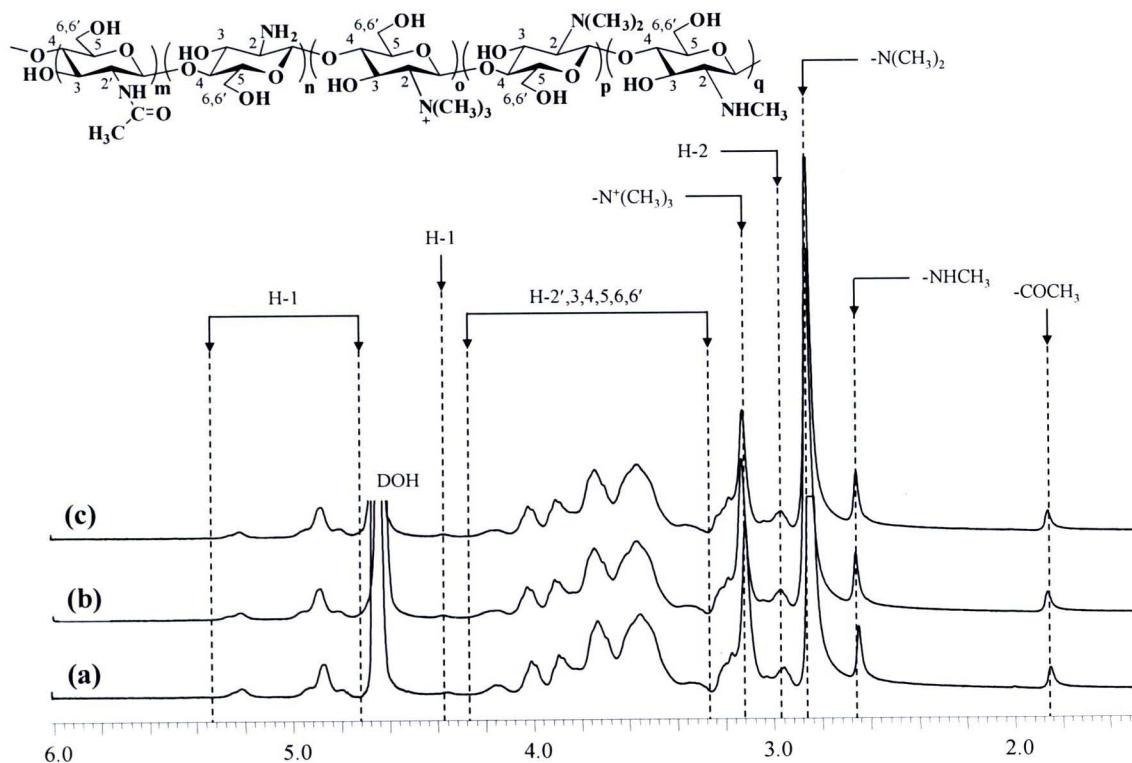


Figure A-1 1H NMR spectra of synthesized TMCs using (a) 4, (b) 5, and (c) 6 equivalents of CH_3I in comparison with the number of NH_2 in chitosan (solvent: $D_2O/TFA, 25^\circ C$).

Table A-1 Degree of quaternization (% DQ^a) was determined from the relative ratio between $-N^+(CH_3)_3$ and H-1.

Sample	n	Integration				% DQ^a (1)	average ± SD (2)	% $DS_{N(CH_3)_2}$ (1)	average ± SD (2)	% DS_{NHCH_3} (3)	average ± SD (1)+(2)+(3)	% DS (<i>total</i>) (1)+(2)+(3)
		N [†] (CH ₃) ₃	N(CH ₃) ₂	NHCH ₃	H-1							
TMC _{CH₃14eq}	1	100	220.94	44.82	36.30	30.6	101.4		13.7		145.8	
	2	100	215.98	42.04	60.39	18.4	22.6 ± 6.9	59.6	73.0 ± 24.7	7.7	9.5 ± 3.6	85.7
TMC _{CH₃15eq}	3	100	205.65	38.3	59.25	18.8		57.8		7.2		83.8
	1	100	228.75	50.12	49.42	22.5	77.1		11.3		110.9	
TMC _{CH₃16eq}	2	100	245.20	51.72	33.29	33.4	28.3 ± 5.5	122.8	100.5 ± 22.8	17.3	16.4 ± 4.7	173.4
	3	100	233.79	70.90	38.36	29.0		101.6		20.5		151.1
TMC _{CH₃16eq}	1	100	253.62	7.38	77.74	14.3		54.4		1.1		69.7
	2	100	265.82	19.50	66.85	16.6	24.6 ± 15.8	66.3	107.9 ± 82.6	3.2	11.6 ± 16.3	86.1
	3	100	316.40	71.07	25.98	42.8		203.0		30.4		276.1

$$^a \text{Degree of quaternization} = \%DQ = \left[\frac{\int N^+(CH_3)_3 / g}{\left(\int H - I \right) \times DD} \right] \times 100$$

Table A-2 Degree of quaternization ($\%DQ^a$) was determined from the relative ratio between $-N^+(CH_3)_3$ and H-2.

Sample	n	Integration			$\%DQ^a$	average ± SD (1)	$\%DS_{N(CH_3)_2}$ (2)	average ± SD (1)	$\%DS_{NHCCH_3}$ (3)	average ± SD (1)+(2)+(3)	$\%DS (total)$
		$N^+(CH_3)_3$	$N(CH_3)_2$	$NHCH_3$							
TMC _{CH₃14eq}	1	100	220.94	44.82	21.59	51.5	170.6		69.2		291.2
	2	100	215.98	42.04	26.33	42.2	46.3 ± 4.7	136.7	148.8 ± 18.9	53.2	253.1 ± 33.1
	3	100	205.65	38.3	24.64	45.1	139.1		51.8		232.1
TMC _{CH₃15eq}	1	100	228.75	50.12	29.02	38.3	131.4		57.6		227.2
	2	100	245.20	51.72	29.43	37.8	40.9 ± 5.1	138.9	144.8 ± 17.2	58.6	235.2
	3	100	233.79	70.90	23.74	46.8	164.1		99.6		257.6 ± 45.9
TMC _{CH₃16eq}	1	100	253.62	7.38	32.91	33.8	128.4		7.5		169.7
	2	100	265.82	19.50	30.71	36.2	34.3 ± 1.7	144.3	142.8 ± 13.7	21.2	209.9 ± 45.0
	3	100	316.40	71.07	33.86	32.8	155.7		70.0		258.5

$$^a \text{Degree of quaternization} = \%DQ = \left[\frac{\int N^+(CH_3)_3 / g}{\int (H-2) \times DD} \right] \times 100$$

Table A-3 Degree of quaternization ($\%DQ^a$) was determined from the relative ratio between $-N^+(CH_3)_3$ and $H-2',3,4,5,6,6'$.

Sample	n	Integration			$\%DQ^a$ (1)	$\%DS_{N(CH_3)_2}$ (2)	$\%DS_{N(CH_3)_2}$ average ± SD (3)	$\%DS_{NHCH_3}$ (3)	$\%DS_{(total)}$ (1)+(2)+(3)	average ± SD
		$N^+(CH_3)_3$	$N(CH_3)_2$	$NHCH_3$						
TMC _{CH₃16eq}	1	100	240.15	21.65	360.98	21.7	78.3	14.1	114.1	
	2	100	212.24	13.25	364.34	21.5	21.7 ± 0.1	68.5	71.6 ± 5.7	8.6
	3	100	208.83	12.00	360.69	21.7	68.1	7.8	97.7	10.2 ± 3.4
TMC _{CH₃15eq}	1	100	229.78	36.43	365.82	21.4	73.9	23.4	118.8	
	2	100	218.17	38.11	371.18	21.1	21.1 ± 0.4	69.1	70.3 ± 3.1	24.2
	3	100	218.53	42.09	378.28	20.7	68.0	26.2	114.9	24.6 ± 1.4
TMC _{CH₃16eq}	1	100	264.66	12.72	442.90	17.7	70.3	6.8	94.8	
	2	100	261.50	21.66	439.56	17.8	17.8 ± 0.1	70.0	70.1 ± 0.2	11.6
	3	100	260.75	43.92	438.09	17.9	70.0	23.6	111.5	14.0 ± 8.7

$$^a \text{Degree of quaternization} = \%DQ = \left[\frac{\int N^+(CH_3)_3 / 9}{\left(\int H-2',3,4,5,6,6' / 6 \right) \times DD} \right] \times 100$$

Table A-4 Degree of quaternization ($\%DQ^a$) was determined from the relative ratio between $-N^+(CH_3)_3$ and $H-2,2',3,4,5,6'$.

Sample	n	Integration				$\%DQ^a$ (1)	$\%DS_{N(CH_3)_2}$ (2)	$\%DS_{N(CH_3)_2}$ average ± SD (3)	$\%DS_{NHCH_3}$ (3)	$\%DS_{(total)}$ (1)+(2)+(3)	average ± SD
		$N^+(CH_3)_3$	$N(CH_3)_2$	$NHCH_3$	H-2,2',3,4,5,6'						
TMC _{CH₃14eq}	1	100	220.94	44.82	360.22	25.4	56.1	11.4	92.9		
	2	100	215.98	42.04	398.4	23.0	24.1 ± 1.2	49.6	51.7 ± 3.9	9.7	10.1 ± 1.2
	3	100	205.65	38.3	381.98	24.0	49.3	9.2		82.2	85.8 ± 6.1
TMC _{CH₃15eq}	1	100	228.75	50.12	385.53	23.7	54.3	11.9	89.9		
	2	100	245.20	51.72	412.69	22.2	23.3 ± 0.9	54.4	54.8 ± 0.9	11.5	13.4 ± 3.0
	3	100	233.79	70.90	383.02	23.9	55.9	16.9		88.0	91.5 ± 4.6
TMC _{CH₃16eq}	1	100	253.62	7.38	464.39	19.7	50.0	1.5	71.1		
	2	100	265.82	19.50	478.10	19.1	19.0 ± 0.8	50.9	52.8 ± 4.1	3.7	6.0 ± 6.1
	3	100	316.40	71.07	503.89	18.2	57.5	12.9		73.7	77.8 ± 9.4

$$^{\text{a}} \text{Degree of quaternization} = \%DQ = \left[\frac{\int N^+(CH_3)_3 / 9}{\left(\int H - 2, 2, 3, 4, 5, 6, 6 / 6 \right) \times DD} \right] \times 100$$

Table A-5 Degree of quaternization (%DQ^a) was determined from the relative ratio between $-N^+(CH_3)_3$ and $-NHCOC\bar{H}_3$.

Sample	n	Integration			%DQ ^a (1)	%DS _{N(CH₃)₂} (2)	%DS _{NHCH₃} (3)	%DS (total) (1)+(2)+(3)
		N ⁺ (CH ₃) ₃	N(CH ₃) ₂	NHCH ₃				
TMC _{CH₃14eq}	1	100	220.94	44.82	29.73	19.8	65.6	26.6
	2	100	215.98	42.04	1.87	314.6	216.9 ± 170.7	1019.1
	3	100	205.65	38.3	1.86	316.3	975.6	363.4
TMC _{CH₃15eq}	1	100	228.75	50.12	18.79	31.3	107.4	47.1
	2	100	245.20	51.72	18.74	31.4	27.9 ± 5.9	115.4
	3	100	233.79	70.90	27.94	21.1	73.8	44.8
TMC _{CH₃16eq}	1	100	253.62	7.38	1.82	323.2	1229.6	71.6
	2	100	265.82	19.50	4.67	126.0	155.5 ± 155.0	502.2
	3	100	316.40	71.07	33.78	17.4	82.6	37.1

$$^a \text{Degree of quaternization} = \%DQ = \left[\frac{\int N^+(CH_3)_3 / 9}{(\int NHCOC\bar{H}_3 / 3) \times DD} \right] \times 100$$

APPENDIX B

Determination of IC₅₀ of *In vitro* cytotoxicity on HaCaT cells line

The IC₅₀ values were calculated from a logarithmic regression ($y = a \cdot \log(x) + b$ in Table B-1) which obtained from the graph of the concentration (x axis) and the cell viability (y axis) as shown in Figure B-1.

Table B-1 The IC₅₀ values (*in vitro* cytotoxicity) of chitosan MW 45000, TMCs, and HTACCs on HaCaT cells line as determined by the MTT assay at pH 6.0.

Sample	$y = a \cdot \log(x) + b$			y = 50%	
	a	b	r ²	log (x)	x = IC ₅₀
Chitosan 85%DD	3.6454	91.712	0.2057	-	>5000 ^a
TMC 22%DQ	-13.370	93.279	0.6161	3.237	1725.9
TMC 21%DQ	-28.129	94.633	0.8147	1.587	38.6
TMC 18%DQ	-25.034	78.806	0.9003	1.151	14.1
HTACC 24%DQ	4.8118	91.227	0.4580	-	>1000 ^a
HTACC 84%DQ	-25.316	107.21	0.8141	2.260	181.9
HTACC 139%DQ	-29.271	115.60	0.6599	2.241	174.2

^aThe IC₅₀ value could not certainly calculated due to the log (x) value is negative.

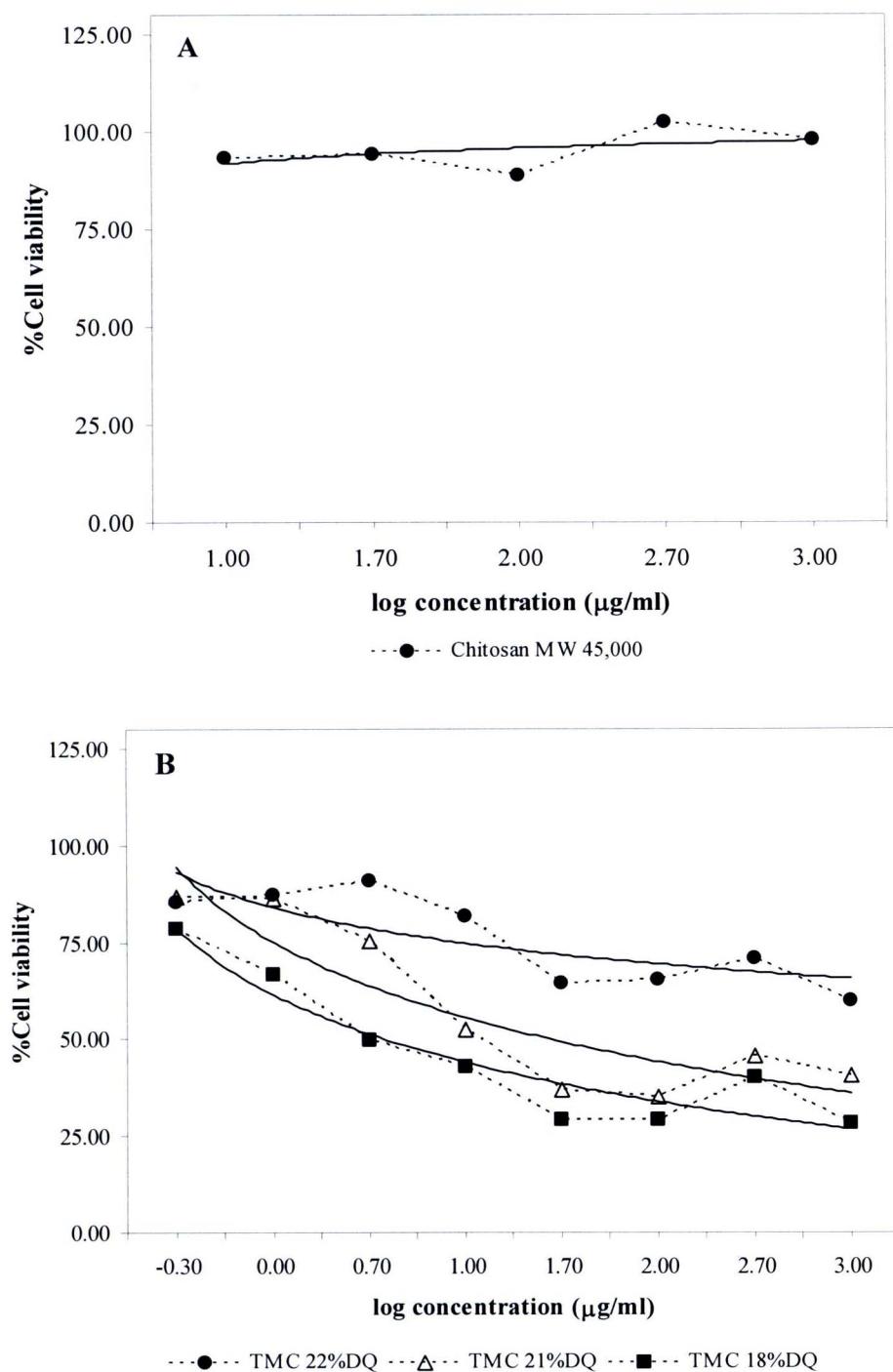


Figure B-1 Relationship between the log concentration ($\mu\text{g/ml}$) and the cell viability (%): (A) chitosan MW 45,000, (B) TMCs, and (C) HTACCs. Logarithmic trendline (solid line) and original trendline (dotted line).

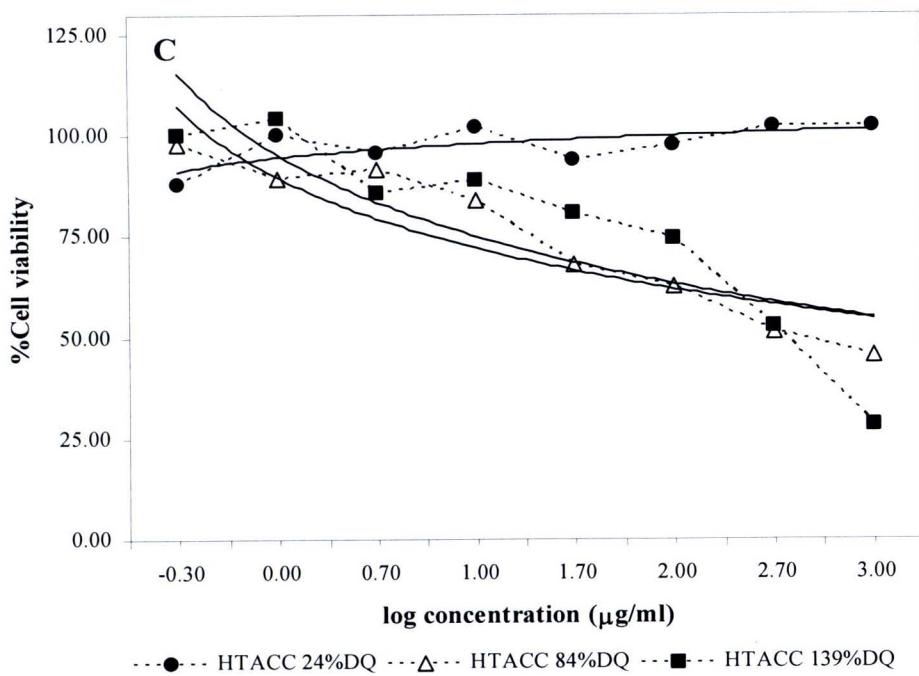


Figure B-1 continued

APPENDIX C

The fracture patterns for five types of hair

As a result, virgin, straightened and UV-damaged hairs are observed with the step pattern as shown in Figure C-1. Waved and dyed hairs are observed with the smooth and splitting pattern, respectively. Additionally, if the hair and its cuticle are near the root end, a smooth break tends to occur. As the fiber becomes dryer, below 90%RH, step fractures are the most commonly observed fracture pattern. Fibrillation and splitting describe a distinct cortical fracturing pattern and tend to occur more with twisted or kinky fibers or when the relative humidity is low.

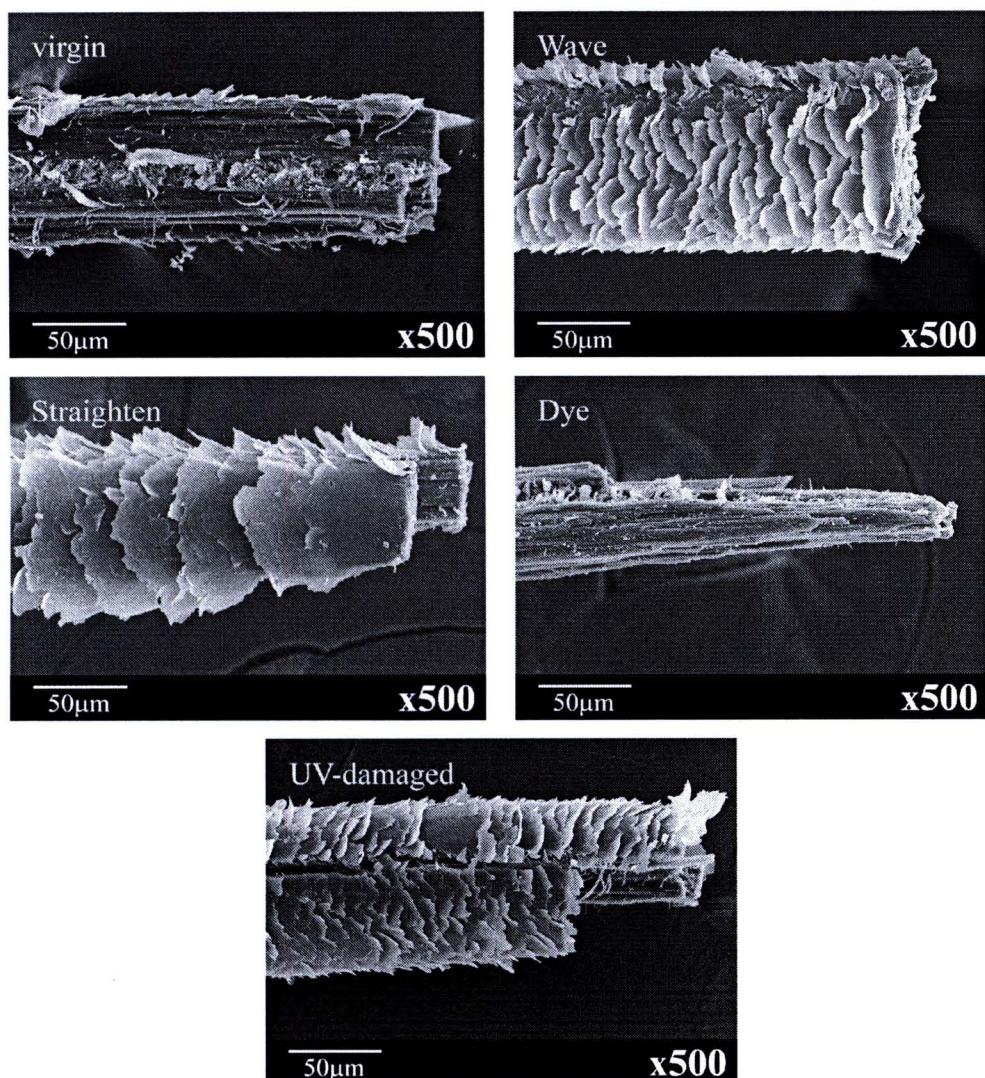


Figure C-1 The fracture patterns for five types of hair.

VITAE



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Presentation in Conference:

- | | |
|----------------|--|
| March 2008 | The 16 th Science Forum, Faculty of Science, Chulalongkorn University, Bangkok, Thailand. |
| July 2008 | The 5 th National Chitin-Chitosan Conference, Center for Chitin-Chitosan Biomaterials (CCB), Metallurgy and Materials Science Research Institute (MMRI), Chulalongkorn University, Bangkok, Thailand. |
| September 2008 | The 5 th Thailand Materials Science and Technology Conference, Miracle Grand Convention Hotel, Bangkok, Thailand. <u>Best Poster Award for Student in Polymer Session</u> |
| December 2008 | The 4 th Mathematics and Physical Science Graduate Conference, Faculty of Science, National University of Singapore, Singapore. |
| January 2009 | Pure and Applied Chemistry International Conference 2009, Faculty of Science, Naresuan University, Phitsanulok, Thailand. <u>The Outstanding Poster Presentation Award</u> |

- Publication:** “Sustained Release of Amoxicillin from Chitosan Tablets” *Archives of Pharmacal Research* 30(4) (2007): 526-531.

