

ABSTRACT

Thesis Title : Ring A Modification of 20-Hydroxyecdysone
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Reaction of 20-hydroxyecdysone 20,22-acetonide (49) with triphenylphosphine-diethyl azodicarboxylate in the presence of formic acid yielded 2-dehydro-3-deoxy-20-hydroxyecdysone 20,22-acetonide (50) as the major product and a mixture of 20-hydroxyecdysone 2-formate 20,22-acetonide (51) and 20-hydroxyecdysone 3-formate 20,22-acetonide (52) as the minor products. The acetonide protecting group in 50 was removed by treatment with 70% acetic acid to give 2-dehydro-3-deoxy-20-hydroxyecdysone (53). Reduction of the 2-dehydro product 50 with sodium borohydride afforded 2-epi-3-deoxy-20-hydroxyecdysone 20,22-acetonide (54) and 3-deoxy-20-hydroxyecdysone 20,22-acetonide (55) in a ratio of 3:1. These two compounds were separately treated with 70% acetic acid to yield 2-epi-3-deoxy-20-hydroxyecdysone (56) and 3-deoxy-20-hydroxyecdysone (57). The moulting hormone activity of the compound 53

in *Musca* bioassay was moderately active, as compared with that of 20-hydroxyecdysone (3). The corresponding alcohol 57 was more active than 53. However its C-2 epimer, the compound 56, was much less active than 57.

Oxidation of 20-hydroxyecdysone 2-acetate 20,22-acetonide (12) with chromium trioxide-pyridine gave 3-dehydro-20-hydroxyecdysone 2-acetate 20,22-acetonide (13) and 20-hydroxyecdysone 2-acetate 3 α ,9 α -epoxide 20,22-acetonide (60), the latter of which was supposed to derive from the former by allylic oxidation at the 9-position followed by intramolecular hemiketal formation. After column chromatography of the crude products consisted of 13 and 60, the compound 2-dehydro-3-epi-20-hydroxyecdysone 3-acetate 20,22-acetonide (53) was additionally obtained, presumably by enolization followed by C-2 to C-3 acetyl migration and C-3 epimerization. Acetonide deprotection of 60 followed by deacetylation yielded 20-hydroxyecdysone 3 α ,9 α -epoxide (62). Reduction of 13 with sodium borohydride gave 3-epi-20-hydroxyecdysone 2-acetate 20,22-acetonide (63), an inseparable mixture of 20-hydroxyecdysone 2-acetate 20,22-acetonide (12) and 20-hydroxyecdysone 3-acetate 20,22-acetonide (64), together with a small quantity of 3-epi-20-hydroxyecdysone 20,22-acetonide (65). Compound 64 derived from C-2 to C-3 acetyl migration of 12 and compound 65 was supposed to be the deacetylation product of 63 occurring in silica column. Sodium borohydride reduction of 59 furnished 2,3-epi-20-hydroxyecdysone 3-acetate 20,22-acetonide (66) and its isomer

2,3-epi-20-hydroxyecdysone 2-acetate 20,22-acetonide (67). Compound 67 was supposed to derive from 66 by C-3 to C-2 acetyl migration. Acetylation of 66 and 67 gave the same diacetate, 2,3-epi-20-hydroxyecdysone 2,3-diacetate 20,22-acetonide (68). When 66 and 67 were separately subjected to deacetylation with guanidine hydrochloride-potassium hydrogen carbonate the same product, 2,3-epi-20-hydroxyecdysone 20,22-acetonide (69), was resulted. With potassium carbonate as the base, both compounds 66 and 67 were converted to 2,3,5-epi-20-hydroxyecdysone 20,22-acetonide (70), which was then epimerized at C-5 to compound 70. Compound 62 was inactive in the *Musca* bioassay. The result was in agreement with of 9,20-dihydroxyecdysone (72) and it was thus concluded that a substituent at C-9 position brought about to complete loss of moulting hormone activity in this bioassay.