CHAPTER IV

RESULTS

The results of this thesis are presented in order as shown in the following flow chart.

4.1 Screening, isolation and identification of organic solvent-tolerant bacteria

- 4.1.1 Primary test of the isolates for organic solvent utilization and tolerance (36 isolates)
- 4.1.2 Secondary test of the isolates for organic solvent utilization and tolerance (12 isolates)
- 4.1.3 Identification of organic solvent-tolerant bacteria
 - 4.1.3.1 The morphological characteristics
 - 4.1.3.2 The biochemical test
 - 4.1.3.3 16S ribosomal DNA gene
- → Four bacterial isolates were chosen for further studies.
- 4.2 Characterization of *Deinococcus geothermalis* T27
- 4.3 Characterization of Bacillus cereus stain 4/1
- 4.4 Characterization of Bacillus subtilis strain 45,
- 4.5 Characterization of Brevibacillus agri strain 13

	Topic n	umber		Topic title
D. geothermalis T27	B. cereus strain 4/1	B. subtilis strain 45	Brevibacillus agri strain 13	•
4.2.1	4.3.1	4.4.1	4.5.1	Characterization of organic-solvent tolerant bacteria
4.2.1.1	4.3.1.1	4.4.1.1	4.5.1.1	Effect of type and concentration of organic solvent on growth and tolerance
4.2.1.2	4.3.1.2	4.4.1.2	4.5.1.2	Effect of organic solvent on cell morphology
4.2.1.3	4.3.1.3	4.4.1.3	4.5.1.3	Effect of organic solvent on fatty acid composition
4.2.1.4	4.3.1.4	4.4.1.4	4.5.1.4	Organic solvent utilization
4.2.1.5	4.3.1.5	4.4.1.5	4.5.1.5	Enzymatic activity involving organic solvent utilization
4.2.2	4.3.2	4.4.2	4.5.2	Factors involving of organic-solvent tolerance
4.2.2.1	4.3.2.1	4.4.2.1	4.5.2.1	Effect of ions on growth and tolerance
4.2.2.2	4.3.2.2	4.4.2.2	4.5.2.2	Effect of nutrient on growth and tolerance

4.1 Screening, isolation and identification of organic solvent-tolerant bacteria

Organic solvent-tolerant bacteria can be isolated from soil. Therefore, soil samples were collected from the oil and hydrocarbon contaminated area, hot spring area, agricultural area and natural area (Table 4.1). Enrichment technique was applied to increase quantity of organic solvent-tolerant bacteria. Bacterial screening was then performed on mineral salt basal (MSB) medium and mineral salt basal medium supplemented with 1% yeast extract (MSBY) agar plate incubated at 45°C with toluene or cyclohexane vapor phase. Thirty-six bacterial isolates that grew on toluene or cyclohexane as a sole carbon and energy source were isolated.

4.1.1 Primary test of the isolates for organic solvent utilization and tolerance

Thirty-six isolates (Table 4.1) were cultured in mineral salt basal medium and mineral salt basal medium supplemented with 1% yeast extract slant directly overlaid with various types of organic solvent at 45°C for 3 day. The growth of bacteria was observed by colony formation. The bacterial isolates could be divided into 5 groups according to mainly their tolerance towards organic solvents with different log $P_{\rm ow}$ values. (Table 4.2): Group I bacteria tolerate to organic solvent with a broad rang of log $P_{\rm ow}$ value (log $P_{\rm ow}$ 0.7-5.6); Group II bacteria tolerate to organic solvent with log $P_{\rm ow}$ 2.0-3.0; Group IV bacteria tolerate to organic solvent with log $P_{\rm ow}$ 2.0-3.0; Group IV bacteria tolerate to organic solvent. Within these five groups of organic solvent-tolerant bacteria, twelve bacteria isolates having comparatively good growth in the presence of organic solvent from each group were chosen for further study (Table 4.3). For group III and group V could not choose for further study because bacteria low tolerate to organic solvent.

4.1.2 Secondary test of the isolates for organic solvent utilization and tolerance

Twelve bacterial isolates chosen from previous study (primary test) were grown at 45°C for 3 day on mineral salt basal liquid medium, mineral salt basal liquid medium supplemented with 1% yeast extract and mineral salt basal liquid medium supplemented with 16mM glucose. Various types of organic solvent were added as vapor phase or directly into cell suspension (20 % v/v) and number of organic solvent in Table 4.2. The growth of bacteria was observed by measuring the optical density at 560 nm using a spectrophotometer. Four bacteria isolates could be selected as representative(s) of each group because growth of these bacterial greater than others (Fig 4.1-4.3).

Table 4.1 Source of soil sample used for screening and bacterial isolates obtained

Enrichment solvent		Source of soil sample	Bacterial isolate number*
		Engine oil, Burirum province	4/1 , 4/2, 6, 6/2, 6/4
	Contaminated	Petroleum oil, Krabi province	-
	area	Machine oil, Krabi province	-
		Machine oil, Bangkok province	8
		Mud, Naeklong district, Krabi province	20, 25/1, 25/2, T27 , 27/1
		Mud, Klongtom district, Krabi province	12, 13 , 22, 36
		Mud, Jaezon, Lumpang province	-
Toluene	Hot spring area	Mud, Pnogdoad, Mahongzon province	-
		Mud, Pai, Mahongzon province	-
		Mud, Sunkumpang, Changmai province	45
		Casava plantation, Rayong province	-
	Agricultural	Rampatan plantation, Krabi province	-
	area	Longan plantation, Chanburi province	-
	Natural area	Soil source, Chulalongkorn University	-
		Engine oil, Burirum province	1, 5/1
	Contaminated	Petroleum oil, Krabi province	-
	area	Machine oil, Krabi province	33
		Machine oil, Bangkok province	-
		Mud, Naeklong district,Krabi province	10,23,24,24/1
		Mud, Klongtom district, Krabi province	21
		Mud, Jaezon, Lumpang province	
Cyclohexane	Hot spring area	Mud, Pnogdoad, Mahongzon province	3C1,3C3,3C4, 3C5,4C1,4C4, 4C6
		Mud, Pai, Mahongzon province	
		Mud, Sunkumpang, Changmai province	44, 46
		Casava plantation, Rayong province	-
	Agricultural	Rampatan plantation, Krabi province	-
	area	Longan plantation, Chanburi province	-
	Natural area	Soil source, Chulalongkorn University	-

^{*} The bolded numbers are the bacterial isolate chosen for further studies

Table 4.2 Primary test for solvent tolerance and solvent utilization on agar overlaid with solvent

See 4.7 4.5 3.5 3.3 3.2 3.1 3.1 3.1 3.1 3.0 2.5	8. <i>o</i> -Xylene 9. <i>m</i> -Xylene 10. <i>p</i> -Xylene	12. Toluene	14. Benzene	15. Chloroform 16. Butyl acetate	I7. n-Butanol	Organic solvent used for screening
Group I Tolerance	3.1 3.1	2.5	3 2.0	2.0 1.8	0.8 0.7	
Croup I Chilization Group I Chilization Count I Chilitation Count I Chilization Chilipation	Group I Toler	ance				
Comp I Offlization	‡	‡	‡	‡	+	Cyclohexane
Coop I Chilization Crop I Chilization Coop I Chilization Character A	‡	‡	,	‡	+	Cyclohexane
Comp I Cilization Group I Cilization Chapter 1	‡	‡	‡	+	+	Cyclohexane
Comp I Clilization Group I Clilization Characteristics	‡	+++++++++++++++++++++++++++++++++++++++	+	++	+	Cyclohexane
Group I Willization Heave the first term of the fi	+	+	++	+ ++	+	Toluene
Group I Utilization Group I Utilization H + + + + + + + + + + + + + + + + + +	+	+	+	+	+	Toluene
Group I Utilization	+		+	+	+	Toluene
	Group I Utiliz	ation				
	-	,	,	+	1	Cyclohexane
+ + + + + + + + + + + + + + + + + + +	+	,		+	+	Cyclohexane
+ + + + + + + + + + + + + + + + + + + +	‡	‡	+	+		Cyclohexane
4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4	+	+	+	+	+	Cyclohexane
	++	,	+	+	+	Toluene
+ + + + + + + + + + + + + + + + + + +	+	+	+	+		Toluene
T27 + + + + + + + + + + + + + + + + + + +	+	‡	+	+	1	Toluene

Table 4.2 (continued)

Organic solvents & Log P _{ow}	l. <i>n</i> -Decane	2. <i>n</i> -Heptane	3. n-Octane	f. n-Hexane	5. Diethylphthalate	5. Cyclohexane	7. Ethylbenzene	9. <i>0-</i> Xylene	onolyX-m.	onolyX-q.01	I. Styrene	f. Toluene	13. I-Heptanol	14. Benzene	ts. Chloroform	66. Butyl acetate	longue-n.7.	18. Ethyl acetate	Organic solvent used for screening
	5.6	4.7	4.5	3.5	3.3	3.2	3.1			3.1	3.0	2.5	2.3	2.0	2.0	1.8	8.0	0.7	
Bacterial isolated									Gro	II dr	Group II Tolerance	ance							
24/1	‡	‡	‡		‡	+	‡	‡	‡	‡	‡	+	+	+	+	+	‡	+	Cyclohexane
23	‡	‡	‡	‡	++	‡	,	,	,	+	+	‡	+	† †	‡			,	Cyclohexane
21	‡	‡	++	-	‡	+	‡	‡	+	‡	ŧ	‡	‡	‡	‡	+	+	+	Cyclohexane
25/1	‡	‡	‡	‡	‡	+	+	‡	‡	+	‡.	‡	+	‡	+	+	+	·	Toluene
9	‡	‡	‡	‡	‡	‡	+	+	+	‡	+	‡	+	‡	+++	+++	1	+	Toluene
45	‡	+	‡	++++	‡	+	+	+	+	+	+	+	+	++++	++++	+	,	+	Toluene
									Grou	II di	Group II Utilization	ation							
24/1			+		+	,	‡	‡	+	+	++++	‡	,		++	+	+	+	Cyclohexane
23	,	,		,	•	,	,	,	,	,		,	,	,		,		,	Cyclohexane
21	‡	+	‡	•	+	+	+	‡	‡	‡	‡	‡	+	+	ı	+	,	+	Cyclohexane
25/1	‡	+	+	+	‡	‡	+	+	+	‡	‡	‡	‡	1	+	+	+	,	Toluene
9	•	,	,	+	+	+	+	+	+	+	+++	+	+	+	+	+	(r	+	Toluene
45	‡	‡	‡	+	‡	‡	‡	+	‡	‡	‡	+	+	‡	+	+		+	Toluene

Table 4.2 (continued)

Solated	Organic solvents & Log Pow	I. <i>n</i> -Decane	2. n-Heptane	3. n-Octane	4. п-Нехапе	5. Diethylphthalate	6. Сусіонехапе	7. Ethylbenzene	9. o-Xylene	9. m-Xylene	10. p-Xylene	II. Styrene	12. Toluene	13. 1-Heptanol	14. Benzene	15. Chloroform	16. Вигуі асегаге	Ionetua-n.71	18. Ейруі асебабе	Organic solvent used for screening
Group III Tolerance		5.6	4.7	4.5	3.5	3.3	3.2	3.1		3.1	3.1	3.0	2.5	2.3	2.0	2.0	1.8	8.0	0.7	
Cross III	Bacterial isolated									roul	IIII	Colera	nce							
The control of the	3C3	++	+	++++	‡	‡	+	‡	+		‡	+	+	‡	+	+	+		,	Cyclohexane
Cross III Control	33	+++		+	,	‡	‡	,			+	+	,	+	++		+	•	,	Cyclohexane
Grant H.	46	+++	+	+	‡	‡	+	+	+	+	+	‡	+	+	+	++++	+	,		Cyclohexane
The control of the	36	‡	+	+	‡	‡	‡	+	+	+	+	+	+	+	+	+	+		+	Toluene
Harry Harr	22	‡	,	1	‡	‡		+	+	+	‡	+		,	‡	+	‡			Toluene
Head of the control	20	‡	‡	‡	+++	‡	,	+	+	+	+	+	‡	+	‡	‡	+	+	+	Toluene
+ ++++ + + + + + + + +	8	+++	‡	‡	+	‡		‡	+	+	+	+	+	+	+	+	+		+	Toluene
Group III Utilization +++	6/2	+	++++	+	1	‡ ‡ ‡	+	+	+	+	+	+	+	‡	+	+	+	,		Toluene
									9	roup	\equiv	Tilizat	ion							
	3C3	+	,	‡	+	+	+	+		+	‡	++	‡	+	+	+	+	,	,	Cyclohexane
	33	+++	ĸ	+	1	‡	‡		,		+	+		+	+++	1	+			Cyclohexane
. .	46	+	+	‡	+	‡	+	‡	‡	+	‡	++++	‡	‡	‡	1	‡		,	Cyclohexane
	36	+	+	+	‡	‡	++	‡	++	+	‡	‡	+	+	‡	+	+		+	Toluene
+ , , , , , , , , , , , , , , , , , , ,	22	‡	,		+++	+++	,	+	+	+	‡	+		,	+++	+	‡	,	,	Toluene
	20	ï				+	1	+	+	+	+	+	+	+	+	+	+	+	+	Toluene
· + + + + + + + + + + + + + + + + + + +	8	•	,	+		+++	•	‡	‡	+	‡	++++	++	‡	,	•	,	,	,	Toluene
	6/2	+	+	+	1	+	+	+	+	+	+	++	+	+	+	+	+			Toluene

+, growth; -, no growth

Table 4.2 (continued)

Solated	Organic solvents & Log P _{ow}	J. n-Decane	2. n-Heptane	3. п-Осбапе	4. n-Hexane	5. Diethylphthalad	6. Cyclohexane	7. Ethylbenzene	9. <i>o-</i> Xylene	9. m-Xylene	919. p-Xylene	II. Styrene	12. Toluene	13. I-Heptanol	14. Benzene	15. Chloroform	16. Butyl acetate	longlu <mark>4-n.</mark> 71	18. Ейруі асебабе	Organic solvent used for screening
Croup IV Tolerance		5.6	4.7	4.5	3.5	3.3	3.2	3.1	3.1	3.1	3.1	3.0	2.5	2.3	2.0	2.0	1.8	8.0	0.7	
Croup IV Utilization	Bacterial isolated									Gro	II dn	Toler	ance							
	34	+				‡	+	+	+	+	‡	‡	+	‡	++	‡	‡	‡	,	Cyclohexane
+ + + + + + + + + + + + + + + + + + +	10	•	‡		‡	•	‡	+	+	+	+	++	‡	‡	-	‡	‡	+	‡	Cyclohexane
+ + + + + + + + + + + + + + + + + + +	4C1	+	+	+	+	++	+	‡	‡	‡	‡	+++	+++	++	++	‡	‡	‡	+	Cyclohexane
+ + + + + + + + + + + + + + + + + + +	5/1	+	+	‡	+	‡	+	‡	‡	‡	‡	+	‡	‡	++++	+	+	+	+	Cyclohexane
+ + + + + + + + + + + + + + + + + + +	4/2	+	+		‡	‡	+	‡	+	+	+	‡	+	+	+	+	++	,		Cyclohexane
+ + + + + + + + + + + + + + + + + + +	4C6/1		+	+	+	+	+	+	+	+	+	++	++	+	+	+	,	+	++	Cyclohexane
Croup IV Utilization Group IV Utilization Group IV Utilization H + + + + + + + + + + + + + + + + + +	13	+	+	+	+	‡	‡	‡	‡	‡	‡	+++	‡	‡	•	‡	‡	‡	‡	Toluene
Group IV Utilization	27/1	‡	+	+	+	‡	+	+	+	+	+	+	+	‡	‡	+	+	+	,	Toluene
Group IV Utilization	6/4		+	‡	,	‡	,	+		,	+	+	,	+	‡	+	+		,	Toluene
Group IV Utilization + + + + + + + + + + + + + + + + + + +	25/2	,				‡	+		‡	+	+	‡	‡	‡		‡	,		,	Toluene
										Gro	up IV		ation							
	34	+	,	,	,	‡	+	+	+	+	‡	+++	‡	‡	+	‡	‡	++		Cyclohexane
	10		‡		‡	,	‡	+	+	+	+	‡	‡	‡	,	‡	‡	‡	‡	Cyclohexane
	4C1	+	+	+	+	‡	+	‡	‡	‡	‡	‡	‡		+	+	+	‡	+	Cyclohexane
<pre>. + , ‡ † † † + + . + † + . + . + + + . + + + . + + + . + + + . + + . + + + . +</pre>	5/1	+	+	+	+	+	+	+		1	+	‡	+	+	+	+	+	+	+	Cyclohexane
+ , ‡ ‡ † + • + † + • + + • + + • † • + ‡ • + ‡ • + ‡ • + ‡ • + ‡ • + ‡	4/2	i.	,	1	+		+		+	,	+	++++			,	+	•	,	1	Cyclohexane
,	4C6/1	ı		1	,	1	+	+	+	+	,	‡	+	,	+	+	+	+	1	Cyclohexane
+ + + + + + + + + + + + + + + + + + + +	13	+		+		+	+	+		1	+	‡	,	+	,	+	1	+	,	Toluene
	27/1	+	‡	‡	+	‡	‡	‡	+	+	‡	+	‡	+	+	+	‡	+	,	Toluene
, , ++++ . , , + . , . + . , . + . , +/9	6/4	Ŀ		+	,		L	+				++++	,			+	+			Toluene
25/2	25/2	,		,	1	‡	+	,	‡	+	+	‡	‡	+		‡	,	1	1	Toluene

Table 4.2 (continued)

Organic solvents & Log Pow	I. <i>n</i> -Dесапе	2. <i>n</i> -Heptane	3. n-Octane	4. n-Hexane	5. Diethylphthalat	6. Cyclohexane	7. Ethylbenzene	8. <i>0-</i> Xylene	эпэГүХ-т. 9	10. <i>p-</i> Xylene	II. Styrene	12. Toluene	13. 1-Heptanol	14. Benzene	15. Chloroform	16. Butyl acetate	longjuð-n.71	18. Еңиуі асетаге	Organic solvent used for screening
	5.6	4.7	4.5	3.5	3.3	3.2	3.1		3.1 3.1	3.1	3.0	2.5	2.3	2.0	2.0	1.8	8.0	0.7	
Bacterial isolated									Gro	V du	Group V Tolerance	ance							
3C1	‡	+	+	+	++	+	+	+	+	+	+	+	+	+	+	+	+	+	Cyclohexane
44	+	+	+++	+	‡	+	+	+	+	+	+	+	+	+	+	+	+	•	Cyclohexane
16/2		,			‡	•		,	+	,		•	,			,	,	1	Toluene
14	‡	+	+	•	+++	+	+	+	+	+	+	+	+	+	+	+	+	1	Toluene
1	‡	,	•	•	‡		,	,	,	,	ï	1	,		,	+		í	Toluene
									Gro	A dn	Group V Utilization	ation	_						
3C1	+	+	+	+	+	+	+	+	+	+	+	++	+	+	+	+	+	+	Cyclohexane
44	+	+	+	+	‡	+	‡	‡	+	+	‡	+	+	+	+	+	+	•	Cyclohexane
16/2	,				1		,	,	,			•	,	,		,	,	,	Toluene
14	•			•	+	+	+	+	+	+	‡	+	+	+	+	+	+	•	Toluene
	,		,	1	,	•		,	1			•	•	,	,				Toluene

 Table 4.3 Organic solvent tolerant bacterial isolates chosen for further study

Group of bacterial isolates		Sacterial nosen for stud	r furth	
Ī	12	3C4	4/1	T27
Organic solvent with a broad rang $\log P_{ow}$ value				
II	25/1	21	6	45
Organic solvent with $\log P_{\rm ow} > 3.0$ value				
Ш	_			
Organic solvent with $\log P_{\rm ow}$ 2.0-3.0 value				
IV Organic solvent with log $P_{\rm ow}$ < 2.0 value	13	4C6/1	4C1	5/1
V Low tolerate to organic solvent	-			

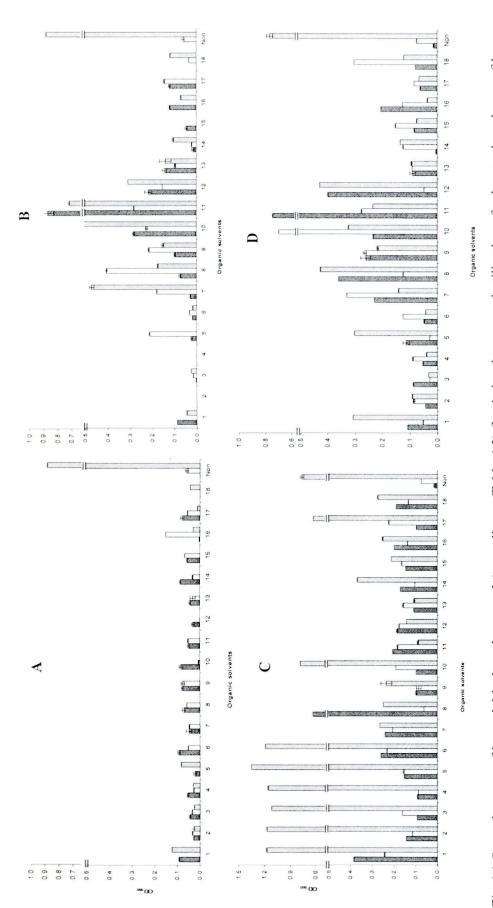


Fig 4.1 Secondary test of bacterial isolates in group I (according to Table 4.3) for their tolerance and utilization of solvent a broad rang of log Pow value. Bacterial isolate 12 (A, B) and isolate 3C4 (C, D) were grown at 45 °C for 3 days in MSB (■), MSBG (□), MSBY (□) medium in presence of solvent vapor (A, C) and directly added (5%, v/v) (B, D). Data are means of the results from at least three individual experiments. The numbers on x-axis represent type of organic solvent shown in Table 4.4.

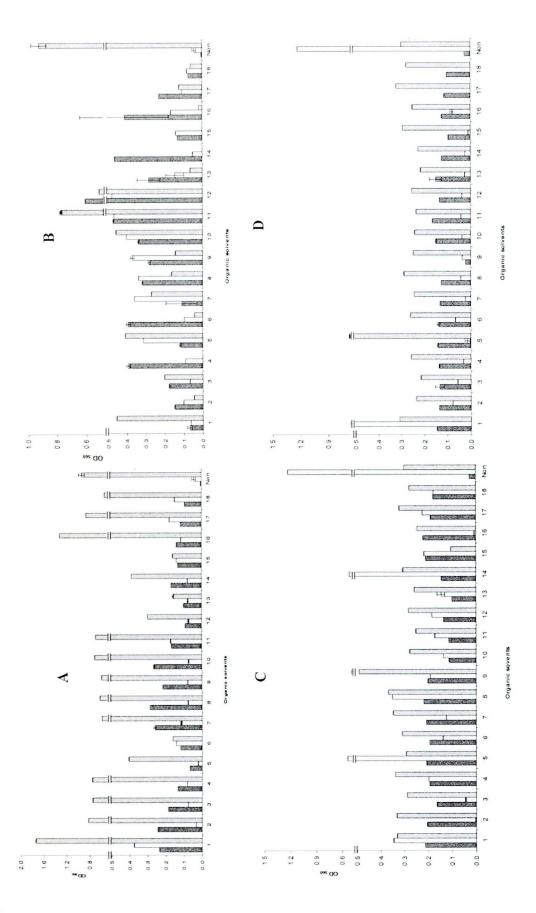


Fig 4.1 (continued) Bacterial isolate 4/1 (A, B) and T27 isolate (C, D) were grown at 45 °C for 3 days in MSB (■), MSBG (□), MSBY (□) medium in presence of solvent vapor (A, C) and directly added (5%, v/v) (B, D). Data are means of the results from at least three individual experiments. The numbers on x-axis represent type of organic solvent shown in Table 4.4.

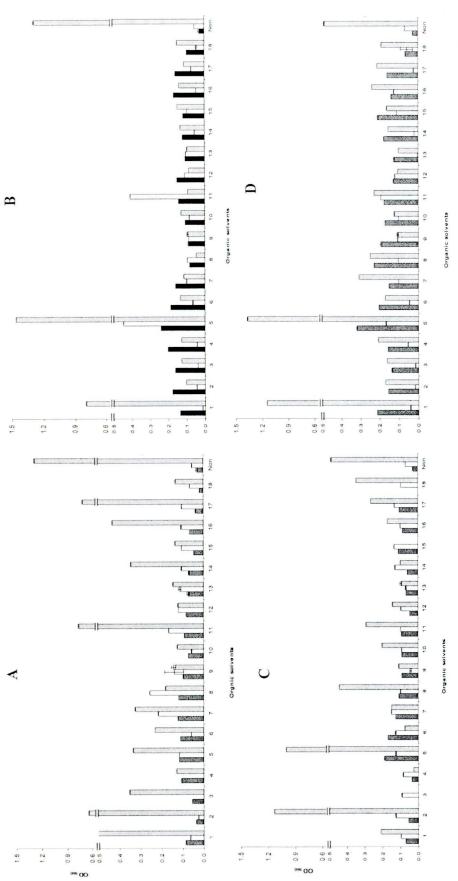
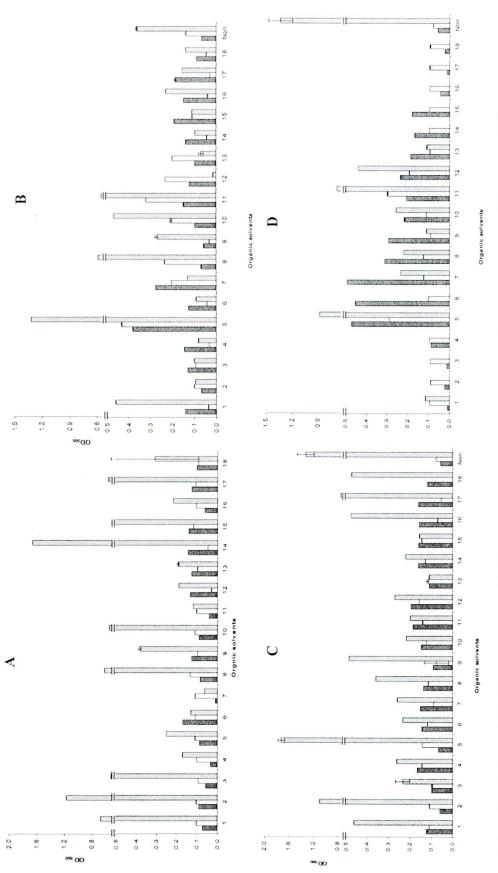


Fig 4.2 Secondary tests of bacterial isolates in group II (according to Table 4.3) for their tolerance and utilization of organic solvent with log medium in presence of solvent vapor (A, C) and directly added (5%, v/v) (B, D). Data are means of the results from at least three individual $P_{\text{ow}} > 3.0$ value. Bacterial isolate 25/1 (A, B) and isolate 21 (C, D) were grown at 45 °C for 3 days in MSB (\blacksquare), MSBG (\square), MSBY (\square) experiments. The numbers on x-axis represent type of organic solvent shown in Table 4.4.



medium in presence of solvent vapor (A, C) and directly added (5%, v/v) (B, D). Data are means of the results from at least three individual Fig 4.2 (continued) Bacterial isolate 6 (A, B) and isolate 45 (C, D) were grown at 45 °C for 3 days in MSB (■), MSBG (□), MSBY (□) experiments. The numbers on x-axis represent type of organic solvent shown in Table 4.4.

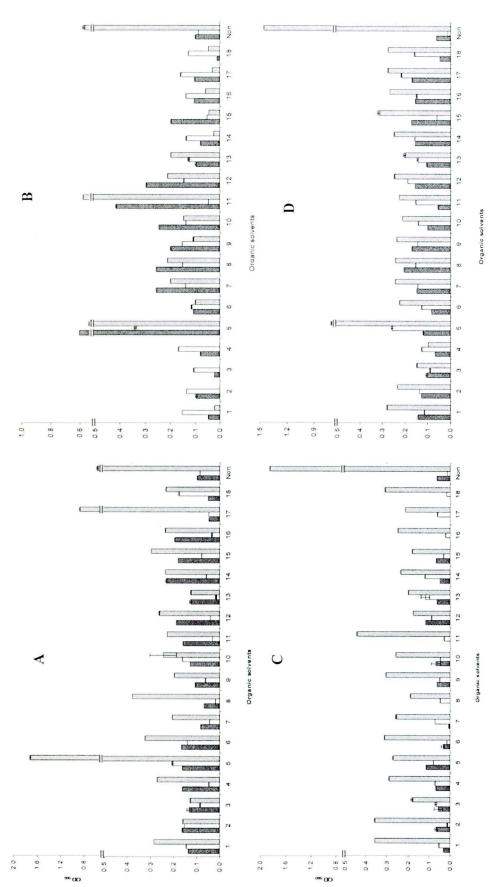
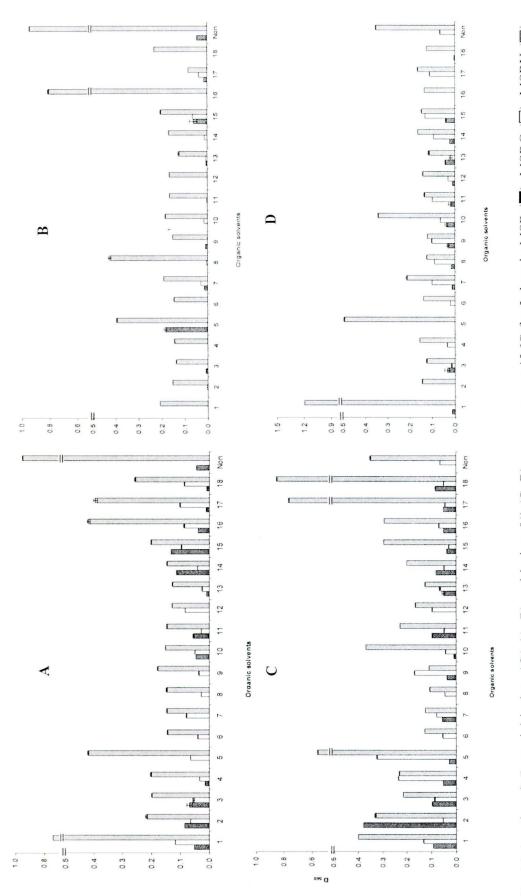


Fig 4.3 Secondary tests of bacterial isolates in group IV (according to Table 4.3) for their tolerance and utilization of organic solvent with log Pow < 2. Bacterial isolate 13 (A, B) and isolate 4C6/1 (C, D) were grown at 45 °C for 3 days in MSB (■), MSBG (□), MSBY (□) medium in presence of solvent vapor (A, C) and directly added (5%, v/v) (B, D). Data are means of the results from at least three individual experiments.

The numbers on x-axis represent type of organic solvent shown in Table 4.4.



medium in presence of solvent vapor (A, C) and directly added (5%, v/v) (B, D). Data are means of the results from at least three individual Fig 4.3 (continued) Bacterial isolate 4C1 (A, B) and isolate 5/1 (C, D) were grown at 45 °C for 3 days in MSB (■), MSBG (□), MSBY (□) experiments. The numbers on x-axis represent type of organic solvent shown in Table 4.4.

4.1.3 Identification of organic solvent-tolerance bacteria

4.1.3.1 The morphological characteristics of organic-solvent tolerance bacteria

Thirty-six bacterial strains were identified for their morphological characteristic by bacterial gram staining (Appendix A). Furthermore, the characteristic of cell colonis on Luria Bertani (as described in Method 3.3.4.1) agar were observed. The characteristics of the four bacterial isolates (isolate T27, 4/1, 45, and 13) are shown in Table 4.4.

4.1.3.2 The biochemical test of organic-solvent tolerance bacteria

The biochemical tests were used to identify the species of bacteria (Appendix A). The biochemical characterization results (Table 4.5) were compared to the Manual of Clinical Microbiology supporting by the laboratory of Institution for Scientific Research, Department of Medical Sciences, and Ministry of Public Health in Thailand, and used to identify type of the bacteria.

4.1.3.3 16S ribosomal DNA gene for organic-solvent tolerance bacterial identification

The determination of 16S ribosomal DNA gene sequence was used to identify organic solvent tolerant bacteria. The specific primers (63f and 1387r) (Marchesi *et al.*,1998) were used to amplify the target sequence as described in Materials and Methods 3.3.3.3. These primers were found to be useful for 16S rDNA gene identification of bacterial species from environmental samples than other PCR primers. After PCR amplification, PCR products were analyzed by using 0.8% agarose gel electrophoresis. The length of the amplified 16S rDNA fragment product was

approximately 1.3 kbps. Subsequently, the blastN program was used to compare and analyze the 16S rDNA sequence against NCBI database (www.ncbi.nlm.gov). The result of alignment of 16S rDNA sequences of 4 bacterial isolates in this study are shown in Table 4.6.

Table 4.4 Morphological characteristic of bacteria

																	1		1	100	W.	1
																			ESE !	ARC	H 1.14	RAT
	Surface	Smooth	Smooth	Rough	Smooth	Smooth	Smooth	Smooth	Rough	Smooth	Smooth	Smooth	Rough	Smooth	Smooth	Smooth	Rough	Rough	Smooth	Smooth	Smooth	Smooth
eristics	Margin	Entire	Entire	Entire	Entire	Undulate	Undulate	Entire	Undulate	Entire	Entire	Entire	Entire	Entire	Entire	Entire	Undulate	Entire	Entire	Entire	Entire	Entire
Colony characteristics	Elevation	Convex	Convex	Flat	Convex	Convex	Convex	Convex	Flat	Convex	Convex	Flat	Flat	Convex	Convex	Convex	Flat	Flat	Convex	Convex	Flat	Convex
Col	From	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circular	Circula	Circular
	Color	Cream	Cream	Yellow-Cream	Pink	Transparent	Cream	Cream	Yellow	Transparent	Yellow	Yellow-Cream	Yellow-Cream	Yellow-Cream	White be turbid	Yellow-Cream	Yellow-Cream	Cream	Transparent	Transparent	Cream	Transparent
Morphology	Cell shape	Bacilli	Bacilli	Rod	Cocci	Bacilli	Bacilli	Bacilli	Rod	Coccobacilli	Coccobacilli	Rod	Bacilli	Bacilli	Bacilli	Rod	Coccobacilli	Rod	Cocci	Coccobacilli	Bacilli	Bacilli
Mor	Gram stain	Positive	Positive	Positive	negative	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive
Ractorial	Isolated	4/1	12	24	T27	3C5	3C4	4C4	9	23	25/1	21	24/1	45	6/2	8	20	22	33	36	46	3C3
	Groups				-						Ш	='			I			H	•			

PG; poor growth, -; non detect, NR; not reported

Table 4.4 (continue) Morphological characteristic of bacteria

	Dootoriol	Moi	Morphology		Colony c	Colony characteristics	cs	
Groups		Gram stain	Cell shape	Color	From	Elevation	Margin	Surface
	4/2	Positive	Bacilli	Yellow	Circular	Flat	Entire	Rough
	5/1	Positive	Bacilli	White be turbid	Circular	Convex	Entire	Rough
	6/4	Positive	Bacilli	White	Circular	Convex	Entire	Smooth
	10	Positive	Rod	Cream	Circular	Convex	Entire	Smooth
IV	13	Positive	Bacilli	Cream	Circular	Flat	Entire	Smooth
	25/2	Positive	Rod	Transparent	spindle	Convex	Entire	Smooth
•	27/1	Positive	Rod	White	Circular	Convex	Entire	Smooth
•	34	Positive	Bacilli	Cream	Circular	Convex	Entire	Smooth
	4C1	Positive	Coccobacilli	White be turbid	Circular	Convex	Entire	Smooth
	4C6	Positive	Coccobacilli	Cream	Circular	Convex	Entire	Smooth
	-	Positive	Bacilli	Pink	Circular	Convex	Entire	Smooth
	14	Positive	Bacilli	Yellow-Cream	Circular	Flat	Undulate	Rough
>	16/2	Positive	Cocci	Cream be turbid	Circular	Convex	Undulate	Smooth
	44	Positive	Bacilli	Cream	Circular	Convex	Entire	Smooth
	3C1	Positive	Rod	White	Circular	Convex	Undulate	Smooth

PG; poor growth, -; non detect, NR; not reported

Table 4.5 The biochemical test result of four bacterial isolates were chosen

Dischamical Tests			ial isolates	
Biochemical Tests	No.45	No.4/1	No.13	No.27
Gram stain	+ bacilli	+ bacilli	+ bacilli	+ cocci
Hemolysis	β	β	β	
Anaerobic growth		+		
TSI	K/A	K/A	K/K	N/N
H ₂ S				-
Catalase	+	+	+	+
Oxidase	+	+	+	+
Motility	+	+	+	_
Indole				
Citrate	+	+	-	_
Urease	+	+	-	-
Nitrate	+	+	_	_
N ₂ Gas				
Esculin Escul				-
42°C(Growth)				-
Acetate Acetate				
VP	+	+		+
Gelatinase	+	+	+	
Egg Yolk	-	+		+w
Glucose/Gas	+	+	+	
Lactose	1	L.	T	
Maltose				-
Mannitol	+	-	+	
D-Xylose	+	-	-	-
Sucrose		_		-
L-Arabinose	+	_	-	
Fructose	· · · · · · · · · · · · · · · · · · ·			_
Trehalose	+	+	+	
Starch hydrolysis	+	+	+	+
LDA		_		<u> </u>
Lysine				
Arginine	-		-	-
Ornithine		-		
Alkaline phos.				
Camp test				
Flagella				
Bacterial	Bacillus	Bacillus	Bacillus	Dienococcus
identification	subtilis	cereus	circulans	sp.

Table 4.6 Comparison of 16S ribosomal DNA gene for bacterial identification

Bacterial			Blast	D 1
Isolated	% Similarity	Accession No.	Organism	Results
		AB244465.1	Bacillus cereus strain: C10-1	
4/1	98%	CP000001.1	Bacillus cereus E33L	Bacillus cereus
		AY138279.1	Bacillus cereus strain 2000031513	
		AJ864721.1	Deinococcus geothermalis strain E50051	
T27	99%	CP000359.1	Deinococcus geothermalis DSM 11300	Deinococcus
		AJ000002.1	Deinococcus geothermalis strain E50053	geothermalis
		EU221342.1	Bacillus subtilis strain PAB1C4	
45	100%	EU221340.1	Bacillus subtilis strain YM1C11	Bacillus subtilis
		AB305020.1	Bacillus subtilis strain: BS1	
		AJ586388.1	Brevibacillus agri strain R-20121	D
13	99%	AB039334.1	Brevibacillus agri strain:M1-5	Brevibacillus
		AB112716.1	Brevibacillus agri strain:DSM 6348T	agri

4.2 Deinococcus geothermalis T27

4.2.1 Characterization of organic-solvent toleran

4.2.1.1 Effect of types and concentrations of organic solvent on growth and tolerance

Growths of *D. geothemalis* T27 were monitored after solvent shock with various types and quantities of organic solvents. Cells were grown to late log phase (about 6 h) before the solvent was added (Method 3.3.5.1). Cell growth and cell survival was assessed by viable cell number (CFU.ml $^{-1}$). Cell survival after solvent shock are displayed in Fig 4.4, showing a relationship between cell number and the amount of solvent added (5% (v/v) and 20% (v/v)) to the media compared to that in the absenc of organic solvent. Organic solvents were re-classified into three groups:

- 1) organic solvent with $\log P_{\text{ow}} \ge 3.0$; (*n*-decane (5.6), diethylphthalate (3.3), cyclohexane (3.2), *o*-xylene (3.1), *m*-xylene (3.1), *p*-xylene (3.1)
- 2) organic solvent with log P_{ow} 2-3; (styrene (3.0), toluene (2.5), 1-hepanol (2.3), benzene (2.0)
- 3) organic solvent with $\log P_{\text{ow}} \le 2$ (butyl acetate (1.8), *n*-butanol (0.8), ethyl acetate (0.7).

The results demonstrated that D. geothemalis T27 exhibited tolerance to a broad rage of organic solvent. When exposed to solvent with $\log P_{\rm ow}$ greater than 3, i.e. n-decane (5.6), and diethyl phthalate (3.3), the number of viable cells only slightly decreased at both solvent concentrations tested suggesting a relative high tolerance of cells. Although the $\log P_{\rm ow}$ value of cyclohexane (3.2) is close to that of diethylphathalate, cells were adversely affected by the solvent exposure in that cell viability was decreased by ten times 3-h of solvent addition. The exposure of isomers of

xylene (3.1) showed comparatively similar results in that they exhibited toxicity to cells causing cell viability to significantly decrease (Fig 4.4 A, B). The investigation was also carried out with solvents with a log $P_{\rm ow}$ value between 2-3. Styrene (3.0) and n-heptanol (2.3) were clearly toxic to cells at both solvent concentrations. The viable cell numbers were rapidly decreased when the solvent was added. While toluene and benzene showed a modest toxicity effect to cells. A gradual increase of viable cell numbers within 1 h of benzene addition suggested that cells could adapt to the presence of high concentration of benzene (Fig 4.4 C, D).

Further investigation on solvent tolerance of D. geothemalis T27 was extended using solvent with a log $P_{\rm ow}$ value lower 2.0. D. geothemalis T27 demonstrated a sensitivity to n-butanol, which it showed and high tolerance towards butyl acetate and ethyl acetate when exposed to butyl acetate and ethyl acetate; Cells were able to tolerate the solvent toxicity, although slight decrease of the numbers of viable cells was observed immediately after solvent addition (Fig 4.4 E, F).

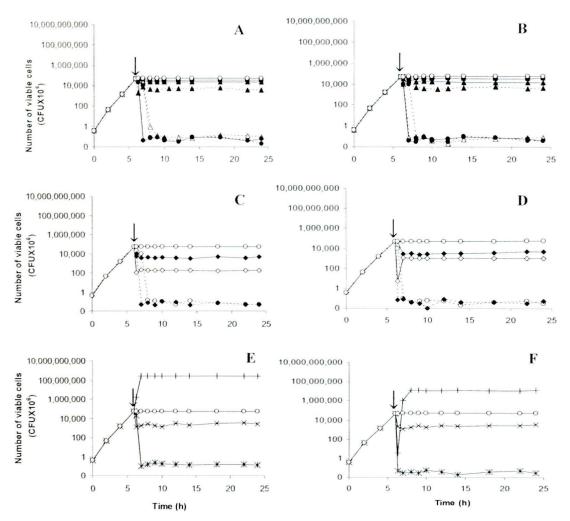
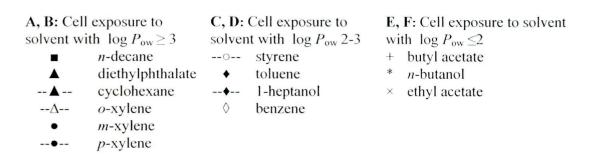


Fig 4.4 Growth inhibitions of D. geothemalis T27 when exposed to organic solvent with $\log P_{\rm ow} \ge 3$ (A, B), $\log P_{\rm ow}$ 2-3 (C, D) and $\log P_{\rm ow} \le 2$ (E, F). Organic solvent was added (\downarrow) to the final volume of 5% (A, C, E) and 20% (B, D, F). Cell growth in absence to organic solvent (\circ) was carried out as a positive control.



4.2.1.2 Effect of organic solvent on cell morphology

D. geothermalis T27 demonstrated unique ability to tolerate ethyl acetate. Effect of organic solvent stress on morphology of D. geothermalis T27 was determined. Cells were grown in HLB medium at 45°C for 6 h, than ethyl acetate was added to medium (20% v/v) and further incubated for 6 h. The control was cells grown in the absence of organic solvent. The morphology of the bacterium exposed to solvent analyzed by the gram strain, SEM and TEM (Fig 4.5). As compared to the morphology of cells grown with and without solvent exposure, D. geothermalis T27 cells in a direct contact to ethyl acetate appears to decreased in size with an approximately 40% size reduction (Table 4.7). Cells dimensions of cylindrical bodies were directly measured from the TEM photographs to calculate cell volume and cell surface area (APPENDIX C).

Table 4.7 Cell size of *D. geothermalis* T27 exposed and not exposed to ethyl acetate

Condition	Surface area (μm²)	Volume (μm³)	
non exposed	0.115 ± 0.010	1.227 ± 0.089	
exposed	0.071 ± 0.038	0.947 ± 0.027	

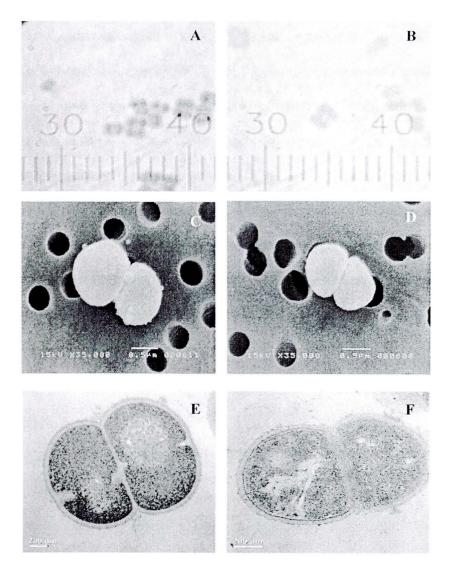


Fig 4.5 Cell morphology of *D. geothermalis* T27. Gram strain (A, B), scanning electron micrographs (C, D) and transmission electron micrographs (E, F) of cells grown in HLB medium for 12 h without organic solvent at 45°C (A, C, E). Gram strain, scanning electron micrographs and transmission electron micrographs of cell were grown in HLB medium for 6 h followed by 6 h ethyl acetate exposure (B, D, F).

4.2.1.3 Effect of organic solvent on fatty acid composition

Fatty acid composition of cells exposed and non-exposed to ethyl acetate was determined based on a method previously described (Unagul *et al.*,2007). The effect of ethyl acetate exposure on the fatty acid composition of cells exposed to ethyl acetate for 6 h was determined (Table 4.8). As shown in Table 4.8, no increase in the concentration of unsaturated fatty acids was observed in ethyl acetate-adapted cells. When *D. geothermalis* T27 grown in HLB medium, it contains approximately 24% and 30% of C15:0 and C17:0, respectively, in relative to all compositions. Fatty acid composition analysis of cells exposed to ethyl acetate revealed that there was no significant change in the level of fatty acid composition or proportion.

Table 4.8 Fatty acid composition of *D. geothermalis* T27 non-exposed and exposed to ethyl acetate

Fatty acid	Bacterial cell fatty acid (% wt)		
	Non-exposure	Ethyl acetate exposure	
C12:0	1.115	-	
C13:0	1.143	-	
C14:0	2.113	0.453	
C14:1	1.565	-	
C15:0	24.517	11.613	
C15:1	1.713	-	
C16:0	11.773	8.986	
C16:1	5.792	-	
C17:0	29.464	14.691	
C17:1	2.637	1.338	
C20:0	1.483	-	

4.2.1.4 Organic solvent utilization

The ability of *D. geothermalis* T27 to utilize of organic solvent as a carbon source was investigated using resting cell technique in minimal medium containing 2.838 mM and 0.709 mM of ethyl acetate (Dashti *et al.*,2008). *D. geothermalis* T27 could degrade 88% and 70% of 2.838 mM and 0.709 mM ethyl acetate respectively in MSB liquid medium within 24 h (Fig 4.6).

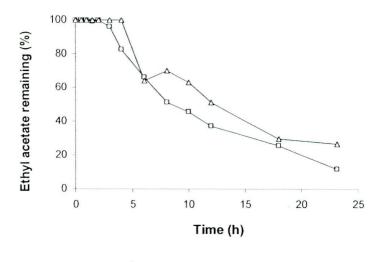


Fig 4.6 Utilization of ethyl acetate by *D. geothermalis* T27. Cells were grown in minimal medium containing 2.838 mM (\square) and 0.709 mM (Δ) of ethyl acetate.

4.2.1.5 Enzymatic activity involving organic solvent utilization

D. geothermalis T27 were grown in 2 conditions of liquid medium 1) mineral medium containing 1% yeast extract, 2) mineral medium containing 1% yeast extract and 5% (v/v) of ethyl acetate. Cells were grown at 45 °C for 24 h. The specific activities of these enzymes were determined by measuring the apparent product using a spectrophotometer. Esterase was apparently detected in intracellular cell exposure to ethyl acetate (Table 4.9), while non-exposure cells showed lower specific activities of esterase. Then, esterase was apparent in extra cellular cell both with and with out ethyl acetate but different specific activities. Suggestion, esterase was inducible enzyme.

Table 4.9 Esterase activity of *D. geothermalis* T27

Growth conditions	Location of enzyme	Specific activities Unit/mg protein	
MSBY	Extra cellular enzyme	0.01 ± 0.001	
MSBY+ ethyl acetate		0.012 ± 0.002	
MSBY	Intracellular enzyme	0.035 ± 0.003	
MSBY+ ethyl acetate		0.309 ± 0.05	

4.2.2 Factors involving of organic-solvent tolerance

4.2.2.1 Effect of ions on growth and organic-solvent tolerance

The influence of various metal ions on cell growth and organic-solvent tolerance was investigated. Bacterial cells were grown at 45°C for 24 h in HLB medium supplemented with 10 mM divalent ions and monovalent ions (Cu²⁺, Zn²⁺, Mn²⁺, Fe²⁺, Co²⁺, Ni²⁺, Mg²⁺, Ca²⁺ and Na⁺). Supplementation of Cu²⁺, Zn²⁺, Mn²⁺, Fe²⁺, Co²⁺ and Ni²⁺ inhibited cell growth; therefore, their influences on organic solvent tolerance were not further examined. Divalent ions Ca²⁺, Mg²⁺, Na⁺ enhanced cell growth; therefore, the influence of divalent ions on stabilization of organic solvent tolerance was investigated. To investigate the influence of metal ions towards solvent tolerance and cell growth, cells were grown on HLB containing Ca²⁺, Mg²⁺, Na⁺ (at 2, 10 and 20 mM) for 6 h then, either toluene (Fig 4.7) and ethyl acetate (Fig 4.8) was added to the final volume of 5% (v/v), 20% (v/v). The result showed that addition of Ca²⁺ at 10 and 20 mM respectively. Ca²⁺ supplementation at all three concentrations tested improved cell tolerance to ethyl acetate. The presence of Mg²⁺ promoted cell growth but did not significantly enhance cell solvent tolerance. The addition of Na⁺ slightly decreased cell yield and did not facilitate the solvent tolerance.

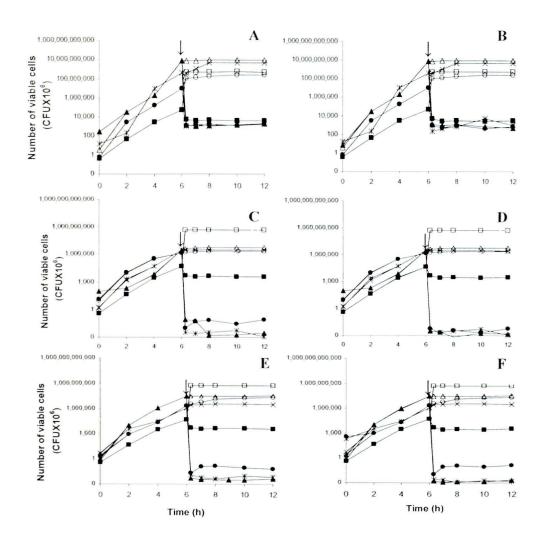


Fig 4.7 Effect of divalent ion on toluene tolerance in *D. geothermalis* T27. Cells were grown on HLB containing Ca^{2+} (A, B), Mg^{2+} (C, D), Na^{+} (E, F) at 2, 10 and 20 mM for 6 h. Then, solvent was added (\downarrow) to 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F) compared to that in the absence of organic solvent (\circ , \times , Δ). Symbols of organic solvents are:

- A,C, E

 2mM ion, 0% solvent

 2mM ion, 5% solvent

 10mM ion, 0% solvent

 10mM ion, 5% solvent

 20mM ion, 0% solvent

 20mM ion, 5% solvent

 0mM ion, 0% solvent

 0mM ion, 5% solvent

 0mM ion, 5% solvent
- B, D, F

 □ 2mM ion, 0% solvent
 □ 2mM ion, 20% solvent
 □ 10mM ion, 0% solvent
 □ 10mM ion, 20% solvent
 □ 20mM ion, 20% solvent
 □ 20mM ion, 20% solvent
 □ 0mM ion, 0% solvent
 □ 0mM ion, 20% solvent
 □ 0mM ion, 20% solvent

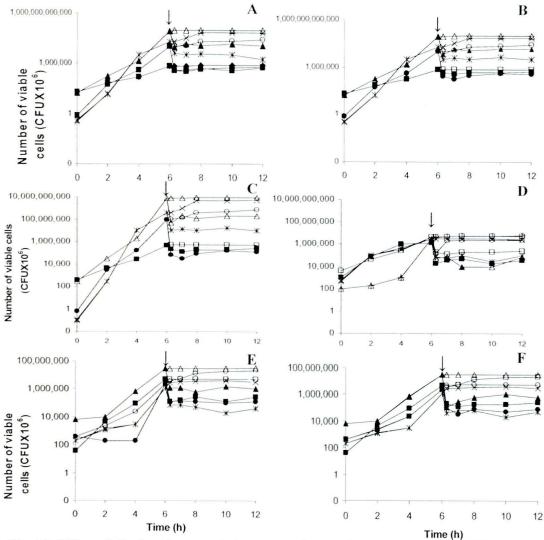


Fig 4.8 Effect of divalent ion on ethyl acetate tolerance in *D. geothermalis* T27. Cells were grown on HLB containing Ca^{2+} (A, B), Mg^{2+} (C, D), Na^{+} (E, F) at 2, 10 and 20 mM for 6 h. Then, solvent was added (\downarrow) to 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F) compared to that in the absence of organic solvent (\circ , \times , Δ). Symbols of organic solvents

A,C, E

○ 2mM ion, 0% solvent

○ 2mM ion, 5% solvent

× 10mM ion, 0% solvent

* 10mM ion, 5% solvent

△ 20mM ion, 0% solvent

△ 20mM ion, 5% solvent

□ 0mM ion, 0% solvent

□ 0mM ion, 5% solvent

□ 0mM ion, 5% solvent

are:

B, D, F

2mM ion, 0% solvent

2mM ion, 20% solvent

10mM ion, 0% solvent

10mM ion, 20% solvent

20mM ion, 0% solvent

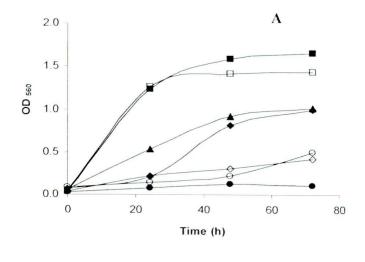
20mM ion, 20% solvent

0mM ion, 0% solvent

0mM ion, 20% solvent

4.2.2.2 Effect of nutrient on growth and organic solvent tolerance of *D. geothermalis* T27

The following carbon sources were tested for cell growth and solvent-tolerance enhancement: MSB medium containing 16 mM of glucose (MSBG), fructose (MSBF), galactose (MSBGa), sucrose (MSBS), xylose (MSBX), rhamnose (MSBR), mannitol (MSBM), citrate (MSBC), succinate (MSBSc), 1% of yeast extract (SBY), tryptone (MSBT), LB, HLB. The addition of MSBGa, MSBS, MSBX, MSBR, MSBM, MSBC and MSBSc slightly affected cell growth. *D. geothermalis* T27 showed good growth rate in MSBG, MSBF, HLB, LB, MSBY and MSBT medium (Fig 4.9). To investigate the influence of nutrient towards solvent tolerance, cells were grown on MSBG, MSBY and HLB for 6 h then, either cyclohexane, toluene, benzene and ethyl acetate was added to 5% (v/v), 20% (v/v). Supplementation of glucose apparently improved cell solvent tolerance resulting in the increase of number viable cells, while the improvement of solvent tolerance was not significantly observed in cells grown in the MSBY medium (Fig 4.10).



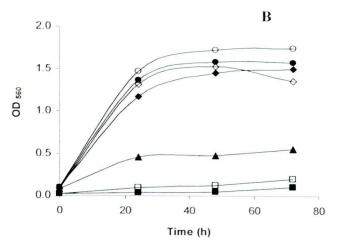


Fig 4.9 Growth of *D. geothermalis* T27 in various types of nutrients. Symbols of organic solvents are:

	Glucose	В		Citrate
	Fructose			Succinate
•	Rhamnose		•	HLB
0	Xylose		0	LB
*	Mannitol		*	MSBY
\Diamond	Galactose		\Diamond	MSBT
A	Sucrose		A	MSBP
	• • •	 □ Fructose • Rhamnose ○ Xylose • Mannitol ◊ Galactose 	 □ Fructose • Rhamnose ○ Xylose • Mannitol ◊ Galactose 	 □ Fructose ■ Rhamnose □ Xylose ■ Mannitol ♦ Galactose

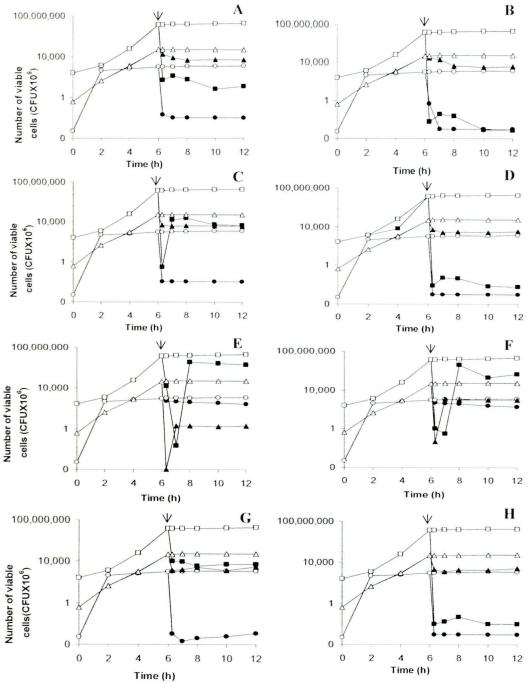


Fig 4.10 Effect of nutrient on cyclohexane (A, B); toluene (C, D); benzene (E, F); ethyl acetate (G, H) tolerance in *D. geothermalis* T27. Cells were grown on MSBG (■), MSBY (●) and HLB (▲) medium at 45°C for 6 h. Then, organic solvent was added (↓) to 5% (v/v) (A, C, E, G, I) and 20% (v/v) (B, D, F, H, J); compared to absent organic solvent □, ○, Δ, respectively.

4.3 Bacillus cereus strain 4/1

4.3.1 Characterization of organic solvent tolerant bacteria

4.3.1.1 Effect of types and concentration of organic solvent on growth and tolerance

Growths of were monitored after organic solvent shock with various types and quantities of organic solvent. Cells were grown to late log phase (about 6 h) before the organic solvent added (Method 3.3.5.1). Cells growth and cell survival was assessed by viable cell number (CFU.mf⁻¹). Cell survival after organic solvent shock are displayed in Fig 4.11, showing a relationship between cell number and the amount of solvent added (5% and 20%, v/v) to the media compared to that in the absence of organic solvent. When exposed to toluene, the number of viable cells slightly decreased at both solvent concentrations tested suggesting a relative high tolerance of cells. When exposed to toluene cells have a good growth but styrene and toluene high toxicity to cells. Cells increase when exposure to low concentration of chloroform. Besides exposed to low concentration of 1-heptanol and to both concentration of toluene, the decrease of cell was observed after 30 min. However increase of within 1h of 1-heptanol and toluene addition suggested that cells could adapt to the 1-heptanol and toluene.

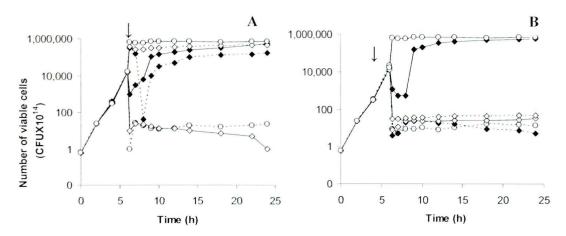


Fig 4.11 Growth inhibition of B. cereus strain 4/1 when exposed to organic solvent. Organic solvent was added (\downarrow) to the final volume 5% (A) and 20% (B). Cell growth in the absence of organic solvent (\circ) was carried out as a positive control. Symbols of organic solvents are:



4.3.1.2 Effect of organic solvent on cell morphology of B. cereus strain 4/1

B. cereus strain 4/1 demonstrated unique ability to tolerate chloroform. Effect of organic solvent stress on morphology of B. cereus strain 4/1 was determined. Cells were grown in HLB medium at 45°C for 6 h, then chloroform was added to medium (20%, v/v) and further incubated for 6 h. The control was cells grown in the absence of organic solvent. The morphology of the bacterium exposed to organic solvent was analysis by gram strain and TEM (Fig 4.12). As compared to the morphology of cells grown with and without organic solvent expose, B. cereus strain 4/1 cells in a direct contract to chloroform appears to not different significance of statistics at confidence 95% with compared control (Table 4.10). B. cereus strain 4/1 exposed to chloroform did not exhibit distorted cell cells structure. The outer and cytoplasmic membrances were

apparently well defined even after the solvent shock. Cells dimensions of cylindrical bodies were directly measured from the TEM photographs to calculate cell volume and cell surface area (APPENDIX C).

Table 4.10 Cell size of B. cereus strain 4/1 exposed and not exposed to chloroform

Condition	Surface area (μm²)	Volume (μm³)
Non expose	0.18 ± 0.126	2.01 ± 0.059
Chlorofrom expose	0.20 ± 0.170	2.21 ± 0.056

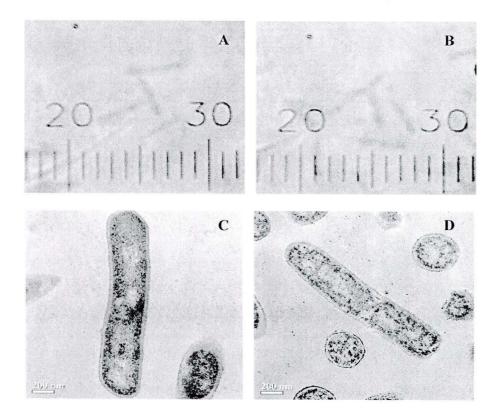


Fig 4.12 Cell morphology of *B. cereus* strain 4/1. Gram, transmission electron micrographs of cell were grown in HLB medium for 12 h at 45°C (A, C). Gram, transmission electron micrographs of cell were grown in HLB medium for 6 h followed by 6 h chloroform (B, D).

4.3.1.3 Effect of organic solvent on fatty acid composition

The effect of chloroform on the fatty acid composition of cells exposed to chloroform for 6 h was determined (Table 4.11). When, cells exposure with chloroform the fatty acid composition (C16:1) 1.67 time increase.

Table 4.11 Fatty acid composition of *B. cereus* strain 4/1 non-exposed and exposed to chloroform

Fatty said	Bacterial cell fatty acid (% wt)		
Fatty acid	Non-exposure	Chloroform-exposure	
C14:0	0.515	-	
C15:0	32.962	29.255	
C15:1	15.056	15.138	
C16:0	5.555	5.613	
C16:1	11.839	19.719	
C17:0	22.662	19.239	
C17:1	0.234	-	



4.3.1.4 Organic solvent utilization

The ability of *B. cereus* strain 4/1 to utilize of organic solvent as a carbon source was investigated using resting cell technique in minimal medium containing 0.678 mM of toluene. *B. subtilis* strain 4/1 can degrade 46.73% of toluene at 0.678 mM (Fig 4.13) in MSB liquid medium within 6 h.

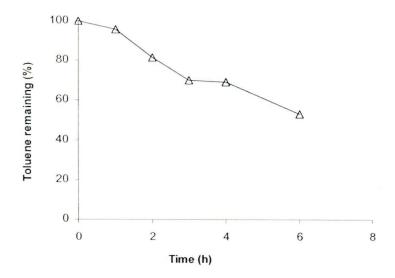


Fig 4.13 Utilization of toluene by *B. cereus* strain 4/1. Cells were grown in minimal medium containing 0.678 mM (Δ) of toluene.

4.3.2 Factors involving of organic-solvent tolerance

4.3.2.1 Effect of ions on growth and organic-solvent tolerance

The influence of various metal ions on cell growth and organic solvent tolerance was investigated. Bacterial cells were grown at 45°C for 24 h in HLB medium supplemented with 10 mM divalent ions and monovalent ions (Cu²⁺, Zn²⁺, Mn²⁺, Fe²⁺, Co²⁺, Ni²⁺, Mg²⁺, Ca²⁺ and Na⁺). Supplementation of Cu²⁺, Zn²⁺, Mn²⁺, Fe²⁺, Co²⁺ and Ni²⁺ inhibited cell growth; therefore, their influences on organic solvent tolerance were not further examined. Divalent ions Ca²⁺, Mg²⁺, Na⁺ enhanced cell growth; therefore, the influence of divalent ions on stabilization of organic solvent tolerance was investigated. To investigate the influence of metal ions towards solvent tolerance and cell growth, cells were grown on HLB containing Ca²⁺, Mg²⁺, Na⁺ (at 2, 10 and 20 mM) for 6 h then, either styrene (Fig 4.14) and toluene (Fig 4.15) was added to the final volume of 5% (v/v), 20% (v/v). The result show that addition of Ca²⁺ at all three concentrations tested improved cell toluene tolerance. While, the presence of Mg²⁺ and Na⁺ at all three concentrations were not significantly affected to styrene and toluene tolerance.

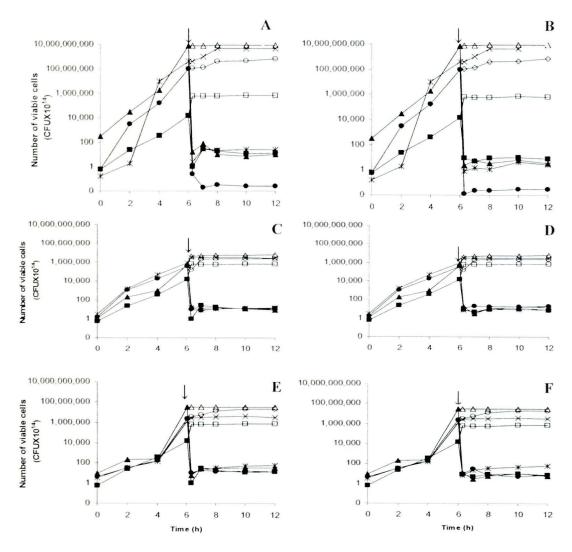


Fig 4.14 Effect of divalent ion on styrene tolerance in *B. cereus* strain 4/1. Cells were grown on HLB containing Ca^{2+} (A, B), Mg^{2+} (C, D), Na^{+} (E, F) at 2, 10 and 20 mM for 6 h. Then, organic solvent was added (\downarrow) to 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F) compared to that in the absence of organic solvent (\circ , \times , Δ). Symbols of organic solvents are:

- A,C, E 2mM ion, 0% solvent
 2mM ion, 5% solvent
 × 10mM ion, 0% solvent
 * 10mM ion, 5% solvent
 △ 20mM ion, 0% solvent
 △ 20mM ion, 5% solvent
 □ 0mM ion, 0% solvent
 □ 0mM ion, 5% solvent
- B, D, F

 □ 2mM ion, 0% solvent

 □ 2mM ion, 20% solvent

 □ 10mM ion, 0% solvent

 □ 20mM ion, 20% solvent

 □ 20mM ion, 20% solvent

 □ 0mM ion, 20% solvent

 □ 0mM ion, 0% solvent

 □ 0mM ion, 20% solvent

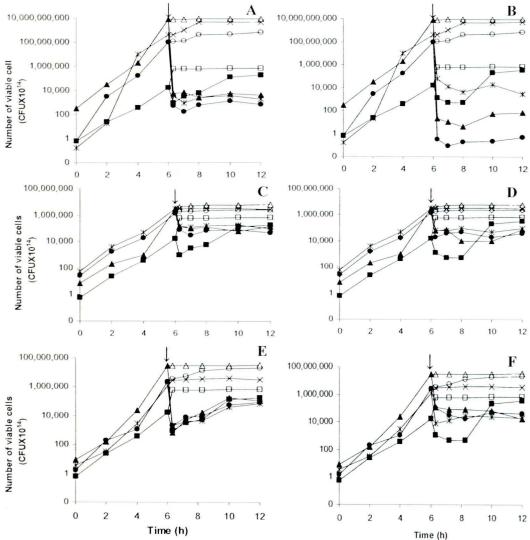


Fig 4.15 Effect of divalent ion on toluene tolerance in *B. cereus* strain 4/1. Cells were grown on HLB containing Ca²⁺ (A, B), Mg²⁺ (C, D), Na⁺ (E, F) at 2, 10 and 20 mM for 6 h. Then, organic solvent was added (\downarrow) to 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F) compared to that in the absence of organic solvent (\circ , \times , Δ). Symbols of organic solvents are:

A,C, E ○ 2mM ion, 0% solvent • 2mM ion, 5% solvent × 10mM ion, 0% solvent * 10mM ion, 5% solvent Δ 20mM ion, 0% solvent Δ 20mM ion, 5% solvent □ 0mM ion, 0% solvent □ 0mM ion, 5% solvent

B, D, F

□ 2mM ion, 0% solvent

□ 2mM ion, 20% solvent

× 10mM ion, 0% solvent

* 10mM ion, 20% solvent

□ 20mM ion, 0% solvent

□ 20mM ion, 20% solvent

□ 0mM ion, 0% solvent

□ 0mM ion, 20% solvent

□ 0mM ion, 20% solvent

4.3.2.2 Effect of nutrient on growth and organic-solvent tolerance

The following carbon sources were tested for cell growth and solvent-tolerance enhancement: MSB medium containing 16 mM of glucose (MSBG), fructose (MSBF), galactose (MSBGa), sucrose (MSBS), xylose (MSBX), rhamnose (MSBR), mannitol (MSBM), citrate (MSBC), succinate (MSBSc), 1% of yeast extract, tryptone, LB, HLB. *B. cereus* strain 4/1 chowed good growth rat in HLB, LB, MSBY, MSBT and MSBP medium (Fig 4.16). To investigate the influence of nutrient towards organic solvent tolerance, cells were grown on MSBY, MSBT and HLB for 6 h at 45°C then, either styrene, toluene and chloroform added to 5% (v/v) and 20% (v/v) (Fig 4.17). Supplementation of yeast extract and tryptone was not significantly solvent tolerance.

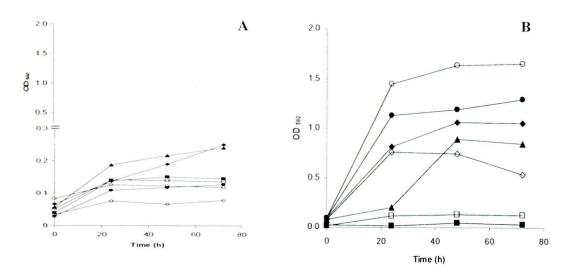


Fig 4.16 Growth of *B. cereus* strain 4/1 in various types of nutrients. Symbols of organic solvents are:

\mathbf{A}		Glucose B		Citrate
		Fructose		Succinate
	•	Rhamnose	•	HLB
	0	Xylose	0	LB
	•	Mannitol	•	MSBY
	\Diamond	Galactose	\Diamond	MSBT
	\blacktriangle	Sucrose	A	MSBP

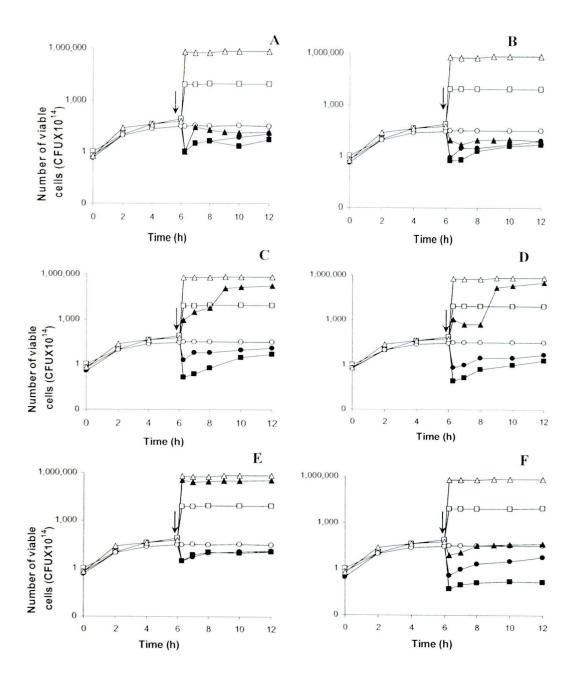


Fig 4.17 Effect of nutrient on styrene (A, B); toluene (C, D); chloroform (E, F) tolerance in *B. cereus* strain 4/1. Cells were grown on MSBT (\blacksquare), MSBY (\bullet) and HLB (\blacktriangle) medium at 45°C for 6 h. Then, organic solvent was added (\downarrow) 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F); compared to absent organic solvent \Box , \circ , Δ , respectively.

4.4 Bacillus subtilis strain 45

4.4.1 Characterization of organic-solvent tolerant bacteria

4.4.1.1 Effect of types and concentrations of organic solvent on growth and tolerance

Growths of *B. subtilis* strain 45 were monitored after solvent shock. Cells were grown to late log phase (about 6 h) before the solvent was added (Method 3.3.5.1). Cell growth and cell survival was assessed by viable cell number (CFU-ml⁻¹). Cells survival after solvent shock are displayed in Fig 4.18, showing a relationship between cell number and the amount of solvent added (5% and 20% v/v) to the media compared to that in the absence of organic solvent. Organic solvents were re-classified into three groups of log P_{ow} value,

- 1) $\log P_{\text{ow}} > 3.5$ (*n*-decane (5.6), *n*-heptane (4.7), *n*-octane (4.5));
- 2) $\log P_{\text{ow}} > 3.2-3.5$ (*n*-hexane (3.5), diethylphthalate (3.3), cyclohexane (3.2);
- 3) $\log P_{\text{ow}} 3.1$ (ethylbenzene (3.1), o-xylene (3.1), m-xylene (3.1), p-xylene (3.1)).

The results demonstrated that *B. subtilis* strain 45 exhibited tolerance to log P_{ow} value > 3 of organic solvent. When exposed to solvent with log P_{ow} greater than 3.5 (Fig 4.18 A, B) and log P_{ow} >3.2-3.5 (Fig 4.18 C, D) the number of viable cells only slightly decreased at both solvent concentrations tested suggesting a relative high tolerance of cells. The exposure of log P_{ow} 3.1 (Fig 4.18 E, F), although the log P_{ow} value of ethylbenzene is equal to isomers of xylene, cells were adversely affected. The exposure of isomers of xylene showed difference results in that they exhibited toxicity to cells.

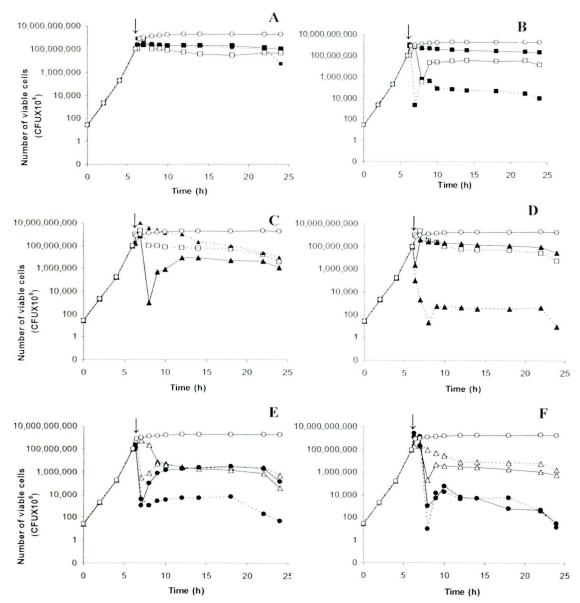


Fig 4.18 Growth inhibition of *B. subtilis* strain 45 when exposed to organic solvent with $\log P_{\rm ow} > 3.5$, $\log P_{\rm ow} 3.2$ -3.5 and $\log P_{\rm ow} 3.1$. Organic solvent was added (\downarrow) to the final volume of 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F). Cell growth in the absent of organic solvent (\circ) was carried out as a positive control. Symbols of organic solvents are:

A, B:	$\log P_{\rm ow} > 3.5$	C , D : log	$g P_{ow} > 3.2-3.5$	E , F :	$\log P_{\rm ow} 3.1$
■ ■	<i>n</i>-decane<i>n</i>-heptane<i>n</i>-octane	□ ▲ ▲	<i>n</i> -hexane diethylphthalate cyclohexane	Δ Δ •	ethylbenzene o-xylene m-xylene p-xylene

4.4.1.2 Effect of organic solvent on cell morphology of B. subtilis strain 45

B. subtilis strain 45 demonstrated unique ability to tolerate n-decane. Effect of organic solvent stress on morphology of B. subtilis strain 45 was determined. Cells were grown in HLB medium at 45°C for 6 h, then n-decane was added to medium (20%, v/v) and further incubated for 6 h. The control was cells grown in the absence of organic solvent. The morphology of the bacterium exposed to organic solvent was analyzed by gram stain and TEM (Fig 4.19). As compared to the morphology of cells grown with and without solvent exposure, B. subtilis strain 45 cells in a direct contact to n-decane not different significance of statistics at confidence 95% with compared control (Table 4.12). B. subtilis strain 45 exposed to chloroform did not exhibit distorted cell cells structure. The outer and cytoplasmic membrances were apparently well defined even after the solvent shock (Fig 4.19 D) Cells dimensions of cylindrical bodies were directly measured from the TEM photographs to calculate cell volume and cell surface area (APPENDIX C).

Table 4.12 Cell size of *B. subtilis* strain 45 grown with and without *n*-decane

Condition	Surface area (μm²)	Volume (μm³)	
non exposed	0.315 ± 0.047	2.850 ± 0.261	
exposed	0.354 ± 0.711	3.124 ± 0.320	

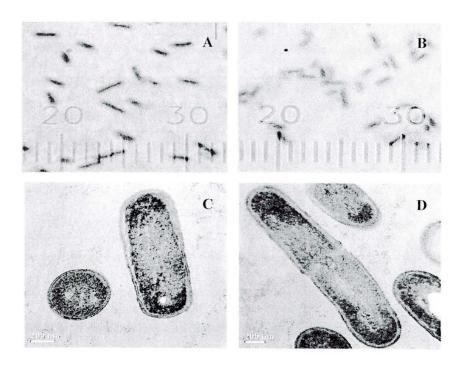


Fig 4.19 Cell morphology of *B. subtilis* **strain 45.** Gram staining and transmission electron micrographs of cell were grown in HLB medium for 12 h at 45°C (A, C). Gram strain and transmission ram strain micrographs of cell were grown in HLB medium for 6 h followed by 6 h *n*-decane (B, D).

4.4.1.3 Effect of organic solvent on fatty acid composition

The effect of *n*-decane on the fatty acid composition of cells exposed to *n*-decane for 6 h was determined (Table 4.13). Fatty acid composition of cells exposed and non-exposed to *n*-decane was determined based on a method previously described (Unagul *et al.*,2007). When *B. subtilis* strain 45 was grown in HLB medium, it contains approximately 22%, 21% and 31% of C15:0, C15:1 and C16:0, respectively, in relative to all compositions. Fatty acid composition analysis of cells exposed to *n*-decane revealed that there was no significant change in the level of fatty acid composition or proportion.

Table 4.13 Fatty acid composition of *B. subtilis* strain 45 non-exposed and exposed to *n*-decane

Fatty acid	Bacterial cell fatty acid (% wt)		
	Non-exposure	<i>n</i> -decane-exposure	
C8:0	1.538	-	
C14:0	1.385	-	
C15:0	21.769	27.314	
C15:1	20.815	24.135	
C16:0	3.446	3.717	
C16:1	30.603	24.027	
C17:0	14.144	17.219.	
C22:2	4.99	2.153	

4.4.1.4 Organic solvent utilization

The ability of *B. subtilis* strain 45 to utilize of organic solvent as a carbon source was investigated using resting cell technique in minimal medium containing 0.878 mM and 1.485 mM of *n*-decane and cyclohexane respectively. *B. subtilis* strain 45 can degrade 54.37% and 32.65% of *n*-decane (Fig 4.20) and cyclohexane respectively (Fig 4.21) in MSB liquid medium within 6 h.

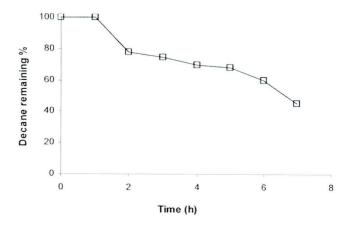


Fig 4.20 Utilization of *n*-decane by *B. subtilis* strain 45. Cells were grown in minimal medium containing 0.878 mM (\square) of *n*-decane.

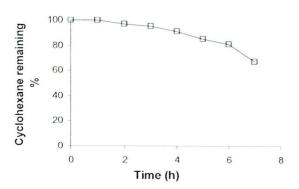


Fig 4.21 Utilization of cyclohexane by B. subtilis strain 45. Cells were grown in minimal medium containing 1.485 mM (\square) of cyclohexane.

4.4.2 Factors involving of organic-solvent tolerance of B. subtilis strain 45

4.4.2.1 Effect of ions on growth and organic-solvent tolerance

The influence of various metal ions on cell growth and organic solvent tolerance was investigated. Bacterial cells were grown at 45°C for 24 h in HLB medium supplemented with 10 mM divalent ions and monovalent ions (Cu²⁺, Zn²⁺, Mn²⁺, Fe²⁺, Co²⁺, Ni²⁺, Mg²⁺, Ca²⁺ and Na⁺). Supplementation of Cu²⁺, Zn²⁺, Mn²⁺, Fe²⁺, Co²⁺ and Ni²⁺ inhibited cell growth; therefore, their influences on organic solvent tolerance were not further examined. Divalent ions Ca²⁺, Mg²⁺, Na⁺ enhanced cell growth; therefore, the influence of divalent ions on stabilization of organic solvent tolerance was investigated. To investigate the influence of metal ions towards solvent tolerance and cell growth, cells were grown on HLB containing Ca²⁺, Mg²⁺, Na⁺ (at 2, 10 and 20 mM) for 6 h then, either 1-heptanol, cyclohexane and ethylbenzene was added 5% (v/v), 20% (v/v). The result show that addition of Ca²⁺ at 2 mM concentrations tested improved cell *n*-heptane tolerance. The presence of Mg²⁺ and Na⁺ at all three concentrations tested improved cell *n*-heptane tolerance (Fig 4.22-4.24).

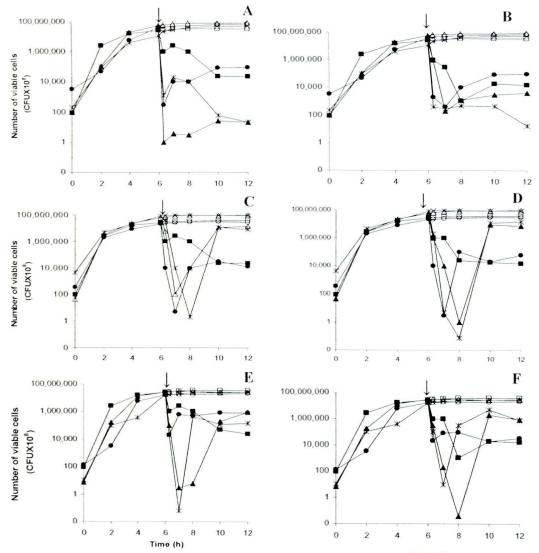


Fig 4.22 Effect of divalent ion on *n*-heptane tolerance in *B. subtilis* strain 45. Cells were grown on HLB containing Ca^{2+} (A, B), Mg^{2+} (C, D), Na^{+} (E, F) at 2, 10 and 20 mM for 6 h. Then organic solvent was added (\downarrow) to 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F) compared to that in the absence of organic solvent (\circ , \times , Δ). Symbols of organic solvents are:

A,C, E

○ 2mM ion, 0% solvent

≥ 2mM ion, 5% solvent

× 10mM ion, 0% solvent

* 10mM ion, 5% solvent

△ 20mM ion, 0% solvent

△ 20mM ion, 5% solvent

□ 0mM ion, 0% solvent

0mM ion, 5% solvent

B, D, F

2mM ion, 0% solvent
2mM ion, 20% solvent
10mM ion, 0% solvent
10mM ion, 20% solvent
20mM ion, 20% solvent
20mM ion, 20% solvent
0mM ion, 0% solvent
0mM ion, 20% solvent
0mM ion, 20% solvent

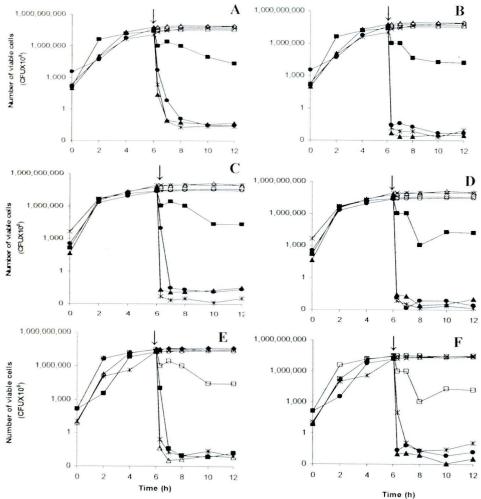


Fig 4.23 Effect of divalent ion on cyclohexane tolerance in B. subtilis strain 45. Cells were grown on HLB containing Ca²⁺ (A, B), Mg²⁺ (C, D), Na⁺ (E, F) at 2, 10 and 20 mM for 6 h. Then, organic solvent was added (\psi) to 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F) compared to that in the absence of organic solvent (\circ , \times , Δ). Symbols of organic solvents are:

A,C, E 2mM ion, 0% solvent

- 2mM ion, 5% solvent
- 10mM ion, 0% solvent
- 10mM ion, 5% solvent
- 20mM ion, 0% solvent Δ
- 20mM ion, 5% solvent 0mM ion, 0% solvent
- 0mM ion, 5% solvent

B, D, F

- 2mM ion, 0% solvent
- 2mM ion, 20% solvent
- 10mM ion, 0% solvent
- 10mM ion, 20% solvent
- Δ 20mM ion, 0% solvent
- 20mM ion, 20% solvent
- 0mM ion, 0% solvent
- 0mM ion, 20% solvent

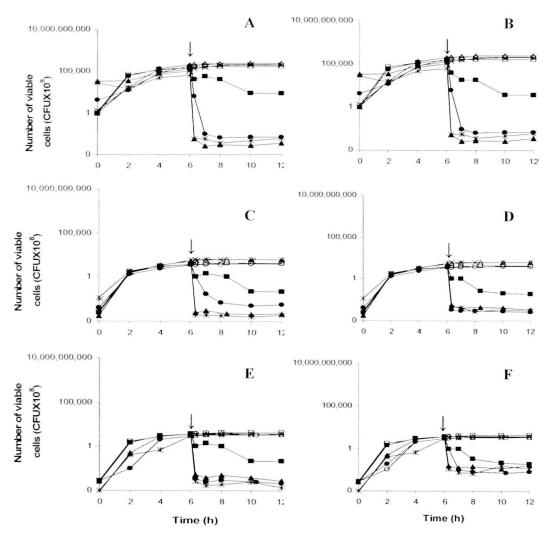


Fig 4.24 Effect of divalent ion on ethylbenzene tolerance in *B. subtilis* strain 45. Cells were grown on HLB containing Ca^{2+} (A, B), Mg^{2+} (C, D), Na^{+} (E, F) at 2, 10 and 20 mM for 6 h. Then, organic solvent was added (\downarrow) to 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F) compared to that in the absence of organic solvent (\circ , \times , Δ). Symbols of organic solvents are:

A,C, E

○ 2mM ion, 0% solvent

○ 2mM ion, 5% solvent

× 10mM ion, 0% solvent

* 10mM ion, 5% solvent

△ 20mM ion, 0% solvent

△ 20mM ion, 5% solvent

□ 0mM ion, 0% solvent

□ 0mM ion, 5% solvent

B, D, F

2mM ion, 0% solvent

2mM ion, 20% solvent

10mM ion, 0% solvent

10mM ion, 20% solvent

20mM ion, 0% solvent

20mM ion, 20% solvent

0mM ion, 0% solvent

0mM ion, 20% solvent

0mM ion, 20% solvent

4.3.2.2 Effect of nutrient on growth and organic solvent tolerance of *B. subtilis* strain 45

The following carbon sources were tested for cell growth and solvent-tolerance enhancement: MSB medium containing 16 mM of glucose (MSBG), fructose (MSBF), galactose (MSBGa), sucrose (MSBS), xylose (MSBX), rhamnose (MSBR), mannitol (MSBM), citrate (MSBC), succinate (MSBSc), 1% of yeast extract, tryptone, LB, HLB. The *B. subtilis* strain 45 showed good growth rates in HLB, LB, MSBY, MSBT and MSBP medium (Fig 4.25). To investigate the influence of nutrient towards organic solvent tolerance, cells were grown on MSBY, MSBT and HLB for 6 h at 45°C then, either *n*-decane, *n*-heptane, *n*-hexane, cyclohexane and ethylbenzene was added to 5% (v/v) and 20% (v/v) (Fig 4.26). Supplementation of yeast extract and tryptone were not significantly to organic solvent tolerance.

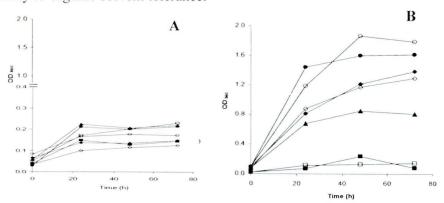


Fig 4.25 Growth of *B. subtilis* strain 45 in varies type of nutrients. Symbols of organic solvents are:

	Glucose]	3		Citrate
	Fructose			Succinate
•	Rhamnose		•	HLB
0	Xylose		0	LB
•	Mannitol		*	MSBY
\Diamond	Galactose		\Diamond	MSBT
A	Sucrose		A	MSBP
	•	 □ Fructose • Rhamnose ○ Xylose • Mannitol ◊ Galactose 	 □ Fructose • Rhamnose ○ Xylose • Mannitol ◊ Galactose 	 □ Fructose □ Rhamnose ○ Xylose ◆ Mannitol ♦ Galactose

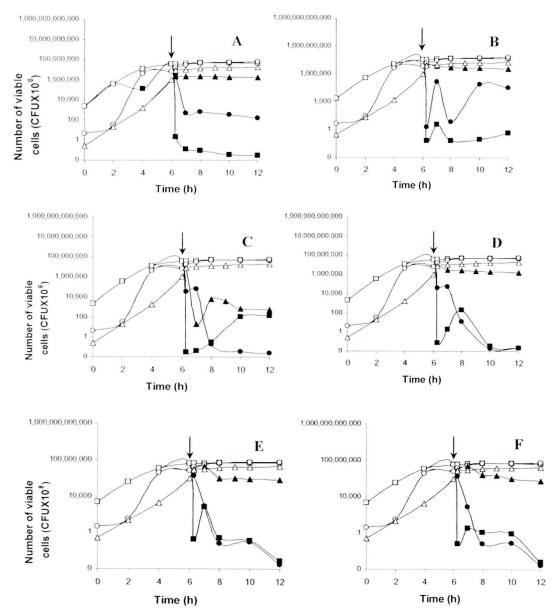


Fig 4.26 Effect of nutrient on *n*-decane (A, B); *n*-heptane (C, D); *n*-hexane (E, F); cyclohexane (G, H); ethylbenzene (I, J) tolerance in *B. subtilis* strain 45. Cells were grown on MSBT (\blacksquare), MSBY (\bullet) and HLB (\blacktriangle) medium at 45°C for 6 h. Then, organic solvent was added (\downarrow) to 5% (v/v) (A, C, E, G, I) and 20% (v/v) (B, D, F, H, J); compared to absent organic solvent \Box , \bigcirc , \triangle , respectively.

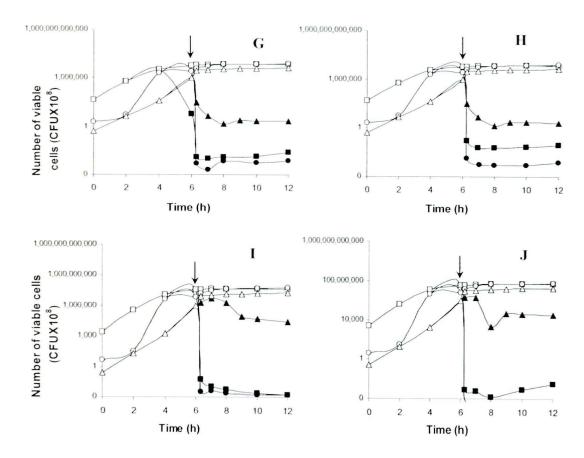


Fig 4.26 (continue) Effect of nutrient on n-decane (A, B); n-heptane (C, D); n-hexane (E, F); cyclohexane (G, H); ethylbenzene (I, J) tolerance in B. subtilis strain 45. Cells were grown on MSBT (\blacksquare), MSBY (\bullet) and HLB (\blacktriangle) medium at 45°C for 6 h. Then, organic solvent was added (\downarrow) to 5% (v/v) (A, C, E, G, I) and 20% (v/v) (B, D, F, H, J); compared to absent organic solvent \Box , \bigcirc , \triangle , respectively.

4.5 Brevibacillus agri strain 13

4.5.1 Characterization of organic solvent tolerant

4.5.1.1 Effect of types and concentration of organic solvent on growth and tolerance

Growths of *Brevibacillus agri strain 13* were monitored after solvent shock with various types and quantities of organic solvents. Cells were grown to late log phase (about 6 h) before the organic solvent was added (Method 3.3.5.1). Cells growth and cell survival was assessed by viable cell number (CFU.mf). Cell survival after organic solvent shock are displayed in Fig 4.27, showing a relationship between cell number and the amount of solvent added (5% and 20% (v/v)) to the media compared to that in the absence of organic solvent. The ability of *Brevibacillus agri strain 13* to withstand solvent toxicity with $\log P_{\rm ow} < 2$ i.e. butyl acetate (1.8), *n*-butanol (0.8) and ethyl acetate (0.7). When exposed to ethyl acetate, the number of viable cells be fast increased at both solvent concentrations tested suggesting a relative high tolerance of cells. Although exposed to *n*-butanol, the number of viable cells slightly increased at 5% (v/v) concentration tested.

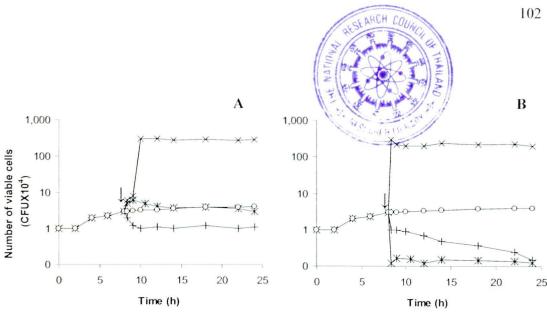


Fig 4.27 Growth inhibition of *Brevibacillus agri strain 13* when exposed to butyl acetate; +, n-butanol; *, ethyl acetate; \times . Organic solvent was added (\downarrow) to the final volume 5% (A) and 20% (B). Cell growth in the absence of organic solvent (\circ) was carried out as a positive control.

4.5.1.2 Effect of organic solvent on cell morphology

Brevibacillus agri strain 13 demonstrated unique ability to tolerant ethyl acetate. Effect of organic solvent stress on morphology of Brevibacillus agri strain 13 was determined. Cells were grown in HLB medium at 45°C for 6 h, then ethyl acetate was added to medium (20%, v/v) and further incubated for 6 h. The control was cells grown in the absence of organic solvent. The morphology of the bacterium exposed to organic solvent was analyzed by gram strain and TEM (Fig 4.28). As compared to the morphology of cells grown with and without organic solvent exposure. Brevibacillus agri strain 13 cells in a direct contact to ethyl acetate not different significance of statistics at confidence 95% with compared control (Table 4.14).

Table 4.14 Cell size of Brevibacillus agri strain 13 exposed with ethyl acetate

Condition	Surface area (µm²)	Volume (μm³)	
non exposed	0.697 ± 0.073	5.167 ± 0.596	
exposed	0.847 ± 0.078	5.984 ± 0.443	

Cell length and radius were estimated by measuring the TEM photographs.

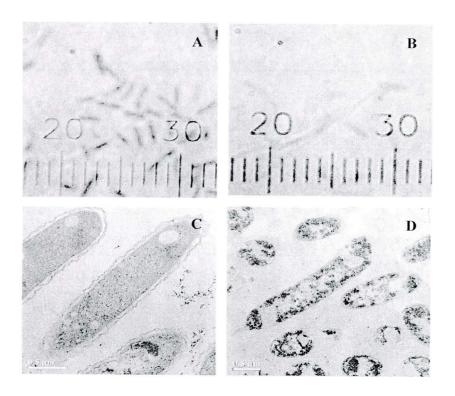


Fig 4.28 Cell morphologies of *Brevibacillus agri* strain 13. Gram strain and transmission electron micrographs of cell were grown in HLB medium for 12 h at 45C (A, C). Gram strain and transmission electron micrographs of cell were grown in HLB medium for 6 h followed by 6 h ethyl acetate (B, D).

4.5.1.3 Effect of organic solvent on fatty acid composition

The effect of ethyl acetate on the fatty acid composition of cells exposed to ethyl acetate for 6 h was determined (Table 4.15). When *Brevibacillus agri* strain 13 was grown in HLB medium non-expose ethyl acetate, it contains approximately 31% of C15:0 and C15:1; 15 % of C16:1, in relative to all compositions. As shown in Table 4.15, increase in the concentration of unsaturated fatty acids was observed in ethyl acetate-adapted cells. Furthermore, under no growth condition could trans-unsaturated fatty acids be detected.

Table 4.15 Fatty acid composition of *Brevibacillus agri* strain 13 non-exposed and exposed to ethyl acetate

Fatty acid	Bacterial cell fatty acid (% wt)		
ratty actu	Non-expose	Ethyl acetate-exposure	
C15:0	31.1	19.2	
C15:1	31.1	22.2	
C16:0	2.8	2.2	
C16:1	14.9	39.5	
C17:0	9.2	7.7	
C17:1	10.8	9.3	

4.5.1.4 Organic solvent utilization

The ability of *Brevibacillus agri* strain 13 to utilize of organic solvent as a carbon source was investigated using resting cell technique in minimal medium containing 1.686 mM and 1.419 mM of *n*-butanol and ethyl acetate respectively. *Brevibacillus agri* strain 13 can degrade 85.89% and 8.53% of (1.686 mM) *n*-butnaol and (Fig 4.29) (1.419 mM) ethyl acetate (Fig 4.30) respectively in MSB liquid medium within 3 h.

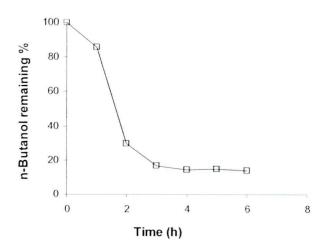


Fig 4.29 Utilization of *n*-butanol by *Brevibacillus agri* strain 13. Cells were grown in minimal medium containing 1.686 mM (\square) of *n*-butanol.

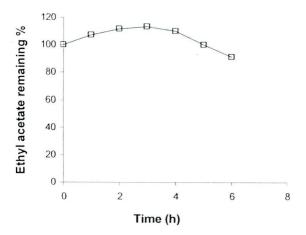


Fig 4.30 Utilization of ethyl acetate by *Brevibacillus agri* strain 13. Cells were grown in minimal medium containing 1.419 mM (□) of ethyl acetate.

4.5.1.5 Enzymatic activity involving organic solvent utilization

Brevibacillus agri strain 13 grown in 2 conditions of liquid medium 1) mineral medium containing 1% yeast extract, 2) mineral medium containing 1% yeast extract and 5% (v/v) of *n*-butanol or ethyl acetate. Cells were grown at 45 °C for 24 h. The specific activities of these enzymes were determined by measuring the apparent product using a spectrophotometer (Vangnai *et al.*,2004). The results shown in Table 4.16 showed that esterase was apparently detected in both extracellular and intracellular cell at similar level whether or not cells were exposed to ethyl acetate.

Similar experiment was carried out using n-butanol as a substrate. Although butanol dehydrogenase activity assay did not yield significant activity level when assayed spectrophotometically, the activity of enzyme towards n-butanol was positively detected using native-polyacrylamilar gel electrophoresis with activity staining. This positive result required lenger incubation period (5 h) in order to detect a band of butanol dehydrogenase. (Fig 4.31)

Table 4.16 Esterase activity of *Brevibacillus agri* strain 13

Growth conditions	Location of enzyme	Specific activities Unit/mg protein	
MSBY	Extra cellular enzyme	0.012 ± 0.0005	
MSBY+ ethyl acetate	J	0.018 ± 0.0010	
MSBY	Intracellular enzyme	0.034 ± 0.0025	
MSBY+ ethyl acetate	J	0.031 ± 0.0008	



Fig 4.31 Native- polyacrylamind gel electrophoresis with activity staining of butanol dehydrogenase by *Brevibacillus agri* strain 13 grown in *n*-butanol. The cell-free extracts (100 μ g protein) from the cells grown on HLB (lane 1) and MSB with *n*-butanol (5% v/v) (lane 2) loaded on a native PAGE and activity stained with the alcohols.

4.5.2 Factors involving of organic-solvent tolerance

4.5.2.1 Effect of ions on growth and organic solvent tolerance

The influence of various metal ions on cell growth and organic solvent tolerance was investigated. Bacterial cells were grown at 45°C for 24 h in HLB medium supplemented with 10 mM divalent ions and monovalent ions (Cu²⁺, Zn²⁺, Mn²⁺, Fe²⁺, Co²⁺, Ni²⁺, Mg²⁺, Ca²⁺ and Na⁺). Supplementation of Cu²⁺, Zn²⁺, Mn²⁺, Fe²⁺, Co²⁺ and Ni²⁺ inhibited cell growth; therefore, their influences on organic solvent tolerance were not further examined. Divalent ions Ca²⁺, Mg²⁺, Na⁺ enhanced cell growth; therefore, the influence of divalent ions on stabilization of organic solvent tolerance was investigated. To investigate the influence of metal ions towards solvent tolerance and cell growth, cells were grown on HLB containing Ca²⁺, Mg²⁺, Na⁺ (at 2, 10 and 20 mM) for 6 h then, either butyl acetate (Fig 4.32), *n*-butanol (Fig 4.33) and ethyl acetate (Fig 4.34) was added to the final volume 5% (v/v), 20% (v/v). The result show that addition of Ca²⁺, Mg² and Na⁺ at all three concentrations tested improved cell growth of *Brevibacillus agri* strain 13. While, the presence of Ca²⁺, Mg²⁺ and Na⁺ at all three concentrations were not significantly to butyl acetate, *n*-butanol and ethyl acetate tolerant.

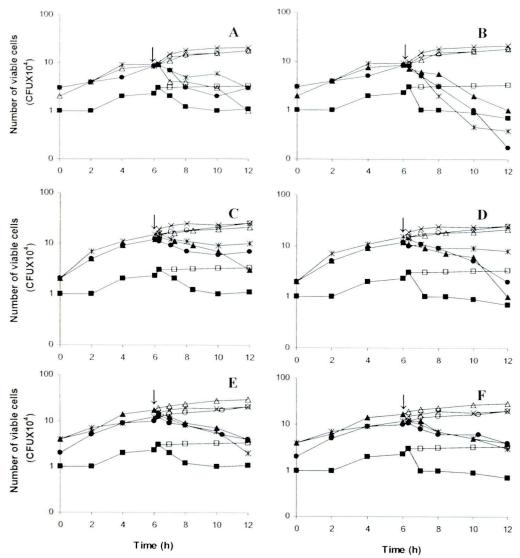


Fig 4.32 Effect of divalent ion on butyl acetate tolerance in *Brevibacillus agri* strain 13. Cells were grown on HLB containing Ca^{2+} (A, B), Mg^{2+} (C, D), Na^{+} (E, F) at 2, 10 and 20 mM for 6 h. Then, solvent was added (\downarrow) to 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F) compared to that in the absence of organic solvent (\circ , \times , Δ). Symbols of organic solvent are:

A,C, E o 2mM ion, 0% solvent
• 2mM ion, 5% solvent

× 10mM ion, 0% solvent

* 10mM ion, 5% solvent

Δ 20mM ion, 0% solventΔ 20mM ion, 5% solvent

DmM ion, 0% solvent

■ 0mM ion, 5% solvent

B, D, F o 2mM ion, 0% solvent

• 2mM ion, 20% solvent

× 10mM ion, 0% solvent

* 10mM ion, 20% solvent

 Δ 20mM ion, 0% solvent

▲ 20mM ion, 20% solvent

□ 0mM ion, 0% solvent

■ 0mM ion, 20% solvent

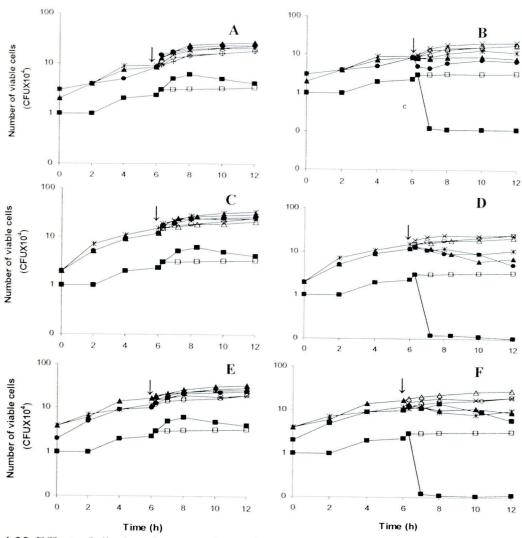


Fig 4.33 Effect of divalent ion on *n*-butanol tolerance in *Brevibacillus agri* strain 13. Cells were grown on HLB containing Ca^{2+} (A, B), Mg^{2+} (C, D), Na^{+} (E, F) at 2, 10 and 20 mM for 6 h. Then, solvent was added (\downarrow) to 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F) compared to that in the absence of organic solvent (\circ , \times , Δ). Symbols of organic solvent are:

A,C,E

○ 2mM ion, 0% solvent

○ 2mM ion, 5% solvent

× 10mM ion, 0% solvent

10mM ion, 5% solvent

△ 20mM ion, 0% solvent

△ 20mM ion, 5% solvent

□ 0mM ion, 0% solvent

□ 0mM ion, 5% solvent

B, D, F ○ 2mM ion, 0% solvent • 2mM ion, 20% solvent × 10mM ion, 0% solvent * 10mM ion, 20% solvent Δ 20mM ion, 0% solvent Δ 20mM ion, 20% solvent □ 0mM ion, 0% solvent □ 0mM ion, 20% solvent

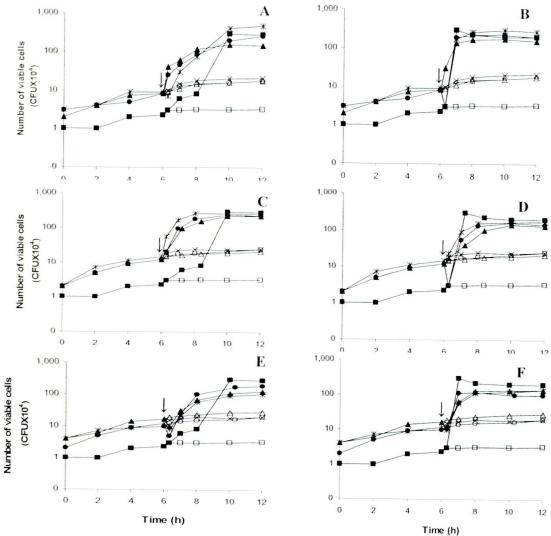


Fig 4.34 Effect of divalent ion on ethyl acetate tolerance in Brevibacillus agri strain 13. Cells were grown on HLB containing Ca2+ (A, B), Mg2+ (C, D), Na+ (E, F) at 2, 10 and 20 mM for 6 h. Then, solvent was added (1) to 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F) compared to that in the absence of organic solvent (\circ , \times , Δ). Symbols of organic solvent are:

B, D, F

A,C, E 2mM ion, 0% solvent 0 2mM ion, 5% solvent 10mM ion, 0% solvent 10mM ion, 5% solvent 20mM ion, 0% solvent Δ 20mM ion, 5% solvent 0mM ion, 0% solvent 0mM ion, 5% solvent

2mM ion, 0% solvent 2mM ion, 20% solvent 10mM ion, 0% solvent 10mM ion, 20% solvent 20mM ion, 0% solvent Δ 20mM ion, 20% solvent 0mM ion, 0% solvent 0mM ion, 20% solvent

4.5.2.2 Effect of nutrient on growth and organic-solvent tolerance

The following carbon sources were tested for cell growth and solvent-tolerance enhancement: MSB medium containing 16 mM of glucose (MSBG), fructose (MSBF), galactose (MSBGa), sucrose (MSBS), xylose (MSBX), rhamnose (MSBR), mannitol (MSBM), citrate (MSBC), succinate (MSBSc), 1% of yeast extract, tryptone, LB, HLB. *Brevibacillus agri* strain 13 showed good growths in HLB, LB medium (Fig 4.35). To investigate the influence of nutrient towards organic solvent tolerance, cells were grown on LB and HLB medium at 45°C for 6 h then, either butyl acetate, *n*-butanol and ethyl acetate was added to 5% (v/v), 20% (v/v) (Fig 4.36).The result show that LB medium tested improved cell growth of *B. circulans* strain 13 but not significantly to tolerant.

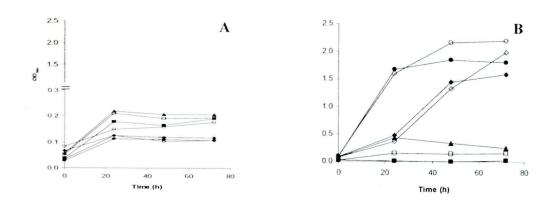


Fig 4.35 Growth of *Brevibacillus agri* strain 13 in various types of nutrients. Symbols of organic solvents are:

\mathbf{A}	•	Glucose	В	-	Citrate
		Fructose			Succinate
	•	Rhamnose		•	HLB
	0	Xylose		0	LB
	*	Mannitol		•	MSBY
	\Diamond	Galactose		\Diamond	MSBT
	A	Sucrose		A	MSBP

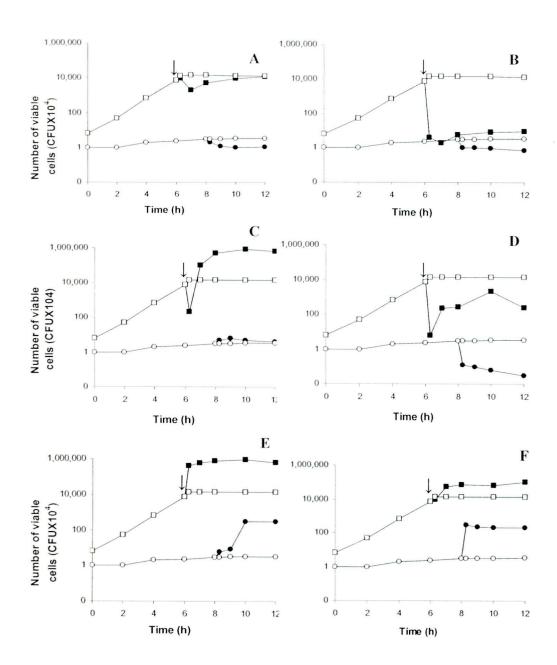


Fig 4.36 Effect of nutrient on butyl acetate (A, B); n-butanol (C, D); ethyl acetate (E, F) tolerance in *Brevibacillus agri* strain 13. Cells were grown on LB (\blacksquare) and HLB (\bullet) medium at 45°C for 6 h. Then, organic solvent was added (\downarrow) 5% (v/v) (A, C, E) and 20% (v/v) (B, D, F).