

RFERENCES

1. Calame K, Lin KI, and Tunyaplin C. Regulatory mechanism that determine the development and function of plasma cells. *Annu Rev Immunol.* 2003; 21: 205-30.
2. Kallies A and Nutt SL. Terminal differentiation of lymphocytes depends on Blimp 1. *Cur Opin Immunol* 2007; 19: 156-62.
3. Lin KI, Duclos CA, Kuo TC, Calame K. Blimp-1-dependent repression of Pax-5 is required for differentiation of B cell to immunoglobulin M-secreting plasma cells. *Mol Cell Biol.* 2002; 22: 4771-80.
4. Holmes ML, Pridans C, Nutt SL. The regulation of the B-cell gene expression programme by Pax5. *Immunol cell Biol.* 2008; 86: 47-53.
5. Shelef SM, Lin KI, Williams Mc, Liao J, Williams MGM, and Calame K. Blimp-1 is required for the formation of immunoglobulin secreting plasma cells and pre-plasma memory B cells. *Immunity.* 2003; 19: 607-20.
6. Lin KI, Tunyaplin C, Calame K. Transcriptional regulatory cascades controlling plasma cell differentiation. *Immunol Rev.* 2003; 194: 19-28.

7. Martins G, Calame K. Regulation and Functions of Blimp-1 in T and B lymphocytes. *Annu Rev Immunol.* 2008; 26: 133-69
8. Tunyaplin C, Shapiro MA, Calame K. Characterization of the B lymphocyte-induced maturation protein-1 (Blimp-1) gene, mRNA isoforms and basal promoter. *Nucleic Acids Res.* 2000; 28: 4846-55.
9. Nagy M, Chapuis B, Matthes T. Expression of transcription factors Pu.1, Spi-B, Blimp-1, BSAP and oct-2 in normal human plasma cells and in multiple myeloma cells. *Br J Haematol.* 2002; 116: 429–35.
10. Nilsson J, Söderberg O, Nilsson K, Rosén A. Differentiation-associated redox-regulation in human B cell lines from stem cell/pro-B to plasma cell. *Immunol Lett.* 2004; 15: 83-9.
11. Shaffer AL, Shapiro-Shelef M, Iwakoshi NN, Lee AH, Qian SB, Zhao H, et al. XBP1, downstream of Blimp-1, expands the secretory apparatus and other organelles, and increases protein synthesis in plasma cell differentiation. *Immunity.* 2004; 21: 81-93.
12. Nakase K, Ishimaru F, Fujii K, Tabayashi T, Kozuka T, Sezaki N, et al. Overexpression of novel short isoforms of Helios in a patient with T-cell acute lymphoblastic leukemia. *Exp Hematol.* 2002; 30: 313-7.

13. Tabayashi T, Ishimaru F, Takata M, Kataoka I, Nakase K, Kozuka T, et al. Characterization of the short isoform of Helios overexpressed in patients with T-cell malignancies. *Cancer Sci.* 2007 Feb; 98(2):182-8.
14. Györy I, Fejér G, Ghosh N, Seto E and Wright KL. Identification of a functionally impaired positive regulatory domain I binding factor 1 transcription repressor in myeloma cell lines. *J Immunol.* 2003, 170: 3125-3133.
15. Keller AD and Maniatis T. Identification and characterization of a novel repression of beta-interferon gene expression. *Gene.* 1991; 5: 868-79.
16. Turner CA, Jr Mack DH, Davis MM. Blimp-1, a novel zinc finger-containing protein that can drive the maturation of B lymphocytes into immunoglobulin-secreting cells. *Cell.* 1994; 77: 297-306.
17. Kallies A, Hasbold J, Tarlinton DM, Dietrich W, Corcoran LM, Hodgkin PD, et al. Plasma cell ontogeny defined by quantitative changes in Blimp-1 expression. *J Exp Med.* 2004; 200: 967
18. Calame K. Transcriptional factors that regulate memory in humoral responses. *Immunol Rev.* 2006; 211: 269-79.

19. Kallies A, Hasbold J, Fairfax K, Pridans C, Emslie D, McKenzie BS, et al.

Initiation of plasma-cell differentiation is independent of the transcription factor Blimp-1. *Immunity*, 2007; 26: 555-566.

20. Lin Y, Wong K, Calame K. Repression of c-myc transcription by Blimp-1, an

inducer of terminal B cell differentiation. *Science*. 1997; 276: 596-9.

21. Sciammas R, Davis MM. Modular nature of Blimp-1 in the regulation of gene

expression during B cell maturation. *J Immunol*. 2004; 172: 5427-40.

22. Tooze RM, Stephenson S and Doody GM. Repression of IFN- γ Induction of Class

II Transactivator: A Role for PRDM1/Blimp-1 in regulation of cytokine signaling. *J Immunol*, 2006, 177: 4584-4593.

23. Piskurich JF, Lin KI, Lin Y, Wang, Ting JPY, Calame K. BLIMP-1 mediates

extinction of major histocompatibility class II transactivator expression in plasma cells. *Nat Immunol*. 2000; 1: 526-32.

24. Buckland J. B-cell Development: BLIMP-1, BCL-6 and B-cell fate. *Nat Rev Immunol*. 2002;2; 629

25. Johnson K, Shelef MS, Tunyaplin C, Calame K. Regulatory events in early and late

B cell differentiation. *Mol Immunol*. 2005; 42: 749-61.

26. Shaffer AL, Lin KI, Kuo TC, Yu X, Hurt EM, Rosenwald, A, et al. Blimp-1 orchestrates plasma cell differentiation by extinguishing the mature B cell gene expression program. *Immunity*. 2002; 17: 51-62.
27. Marker KW and Wilson CB. Sound of a silent Blimp-1. *Nat Immunol*. 2004; 5: 241-2.
28. Northrup DL and Allman D. Transcriptional regulation of early B cell development. *Immunol Res*. 2008; 42: 106-17.
29. Yu J, Angelin-Duclos C, Greenwood J, Liao J, Calame K. Transcriptional repression by blimp-1 (PRDI-BF1) involves recruitment of histone deacetylase. *Mol Cell Biol*. 2000; 20: 2592-603.
30. Gyory I, Wu J, Fejer G, Seto E, Wright KL. PRDI-BF1 recruits the histone H3 methyltransferase G9a in transcriptional silencing. *Nat Immunol*. 2004; 5: 299-308.
31. Carrasco DR, Sukhdeo K, Protopopova M, Sinha R, Enos M, Carrasco DE, et al. The differentiation and stress response factor XBP-1 drives multiple myeloma pathogenesis. *Cancer Cell*. 2007; 11: 349-60.

31. Giorgio C, Cristina AD, Rita S, Zhou H, Wang D, Bachir A. PRDM1/Blimp-1 is expressed in human B-lymphocytes committed to the plasma cell lineage. *Pathology*. 2005; 206: 76-86.
32. Schmidt D, Nayak A, Schumann JE, Schimpl A, Berberich I, Berberich-Siebelt F. Blimp-1 Δ 7: A naturally occurring Blimp-1 deletion mutant with auto-regulatory potential. *Exp Cell Res*. 2008; 314: 3614-3627.
33. Petsophonsakul W, Kaewrakmuk, J, Punyot, M, Charoenkwan P and Nawarawong. Expression of Blimp-1 transcription factors mRNA isoforms in mononuclear cells from bone marrow of leukemia and lymphoma patients. The 13th International congress of immunology, Rio de Janeiro-Brazil, August 21-25, 2007.
34. Shapiro-Shelef M, Lin-I K, Savitsky D, Calame K. Blimp-1 is required for maintenance of long-lived plasma cells in the bone marrow. *J Exp Med* 2005; 202(11): 1471-76. http://www.nature.com/ni/journal/v1/n6/full/ni1200_526.html - a2#a2

APPENDIX

Reagents and buffers

Carbonate buffer (coating buffer)

| | | |
|---------------------------------|-----|---|
| Na ₂ CO ₃ | 1.6 | g |
| NaHCO ₃ | 2.9 | g |
| NaN ₃ | 0.2 | g |

Dissolve reagents in 500 ml distilled water and adjust pH to 9.5 with 1N HCl or 1N NaOH, and then fill distilled water up to 1,000 ml. Store at 4°C.

Phosphate buffer saline

| | | |
|--|------|---|
| NaH ₂ PO ₄ (anhydrous) | 0.23 | g |
| Na ₂ HPO ₄ (anhydrous) | 1.15 | g |
| NaCl | 9.00 | g |

Dissolve reagents in 500 ml distilled water and adjust pH to 7.2-7.4 with 1N HCl or 1N NaOH, and then fill distilled water up to 1,000 ml. Store at 4°C.

PBS-Tween

| | | |
|-------------------------|-------|----|
| Phosphate buffer saline | 1,000 | ml |
| Tween 20 | 500 | μl |

Mix well and store at 4°C.

5% BSA

Phosphate buffer saline 100 ml

BSA 5 g

Mix well and store at 4°C.

10x TBE buffer

Tris Base 108 g

Boric acid 55 g

0.5 M EDTA pH 8.0 40 ml

Dissolved reagents in 500 ml distilled water and then fill distilled water up to 1,000 ml

Store at room temperature.

6x loading buffer

Formamide 2700 µl

10x TBE 150 µl

Xylene cyanol 0.6 mg

Bromophenol blue 0.6 mg

Distilled water 150 µl

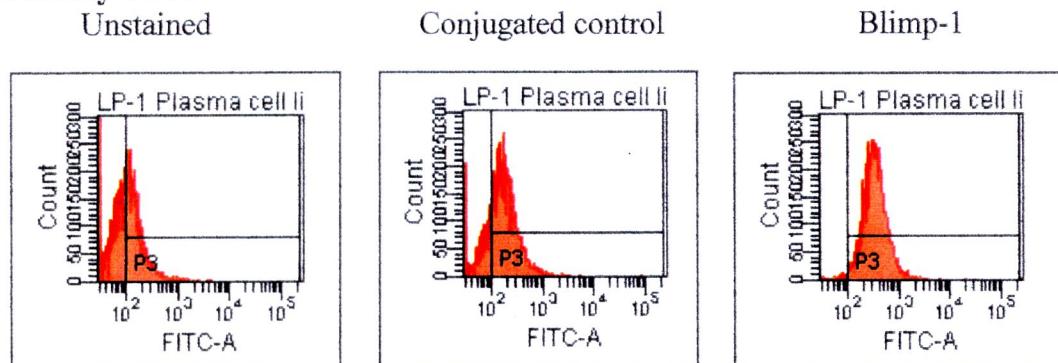
Store at 4°C.

RPMI-10

| | | |
|-----------------------|-----|----|
| RPMI medium | 500 | ml |
| FBS | 50 | ml |
| Sodium pyruvate 100X | 5 | ml |
| Gentamycin (40 mg/ml) | 625 | µl |

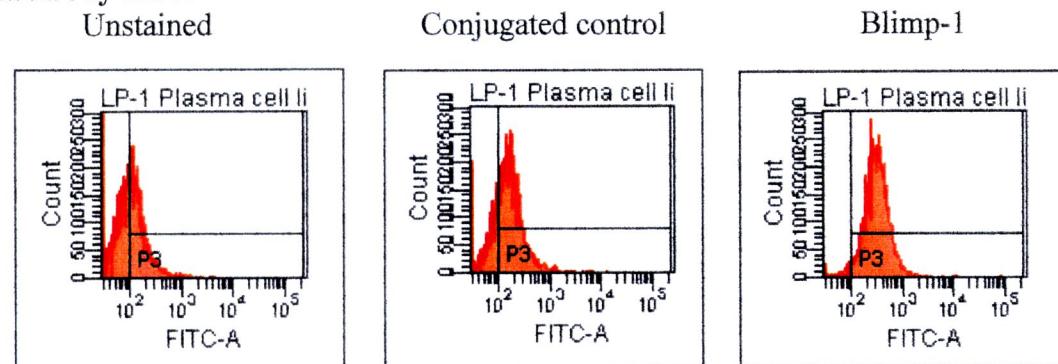
Sterile filtration using 0.45 mM Millipore filter and check sterility by incubation of 2 ml medium in 5% CO₂ at 37°C for a week. Medium was stored at 4°C.

**Titration of FITC-conjugated Antibody
Antibody 1:100**



| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 21.5 | 50.1 | 92.7 |
| FITC Mean | 251 | 301 | 376 |
| FITC Median | 131 | 229 | 322 |

Antibody 1:200



| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 21.5 | 47.5 | 90.1 |
| FITC Mean | 251 | 301 | 376 |
| FITC Median | 131 | 229 | 322 |

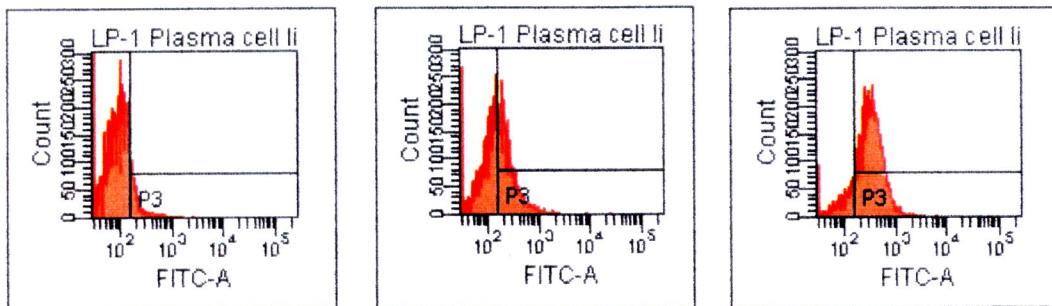
Comparison of permeabilizing agent

Cold Methanol

Unstained

Conjugated control

Blimp-1



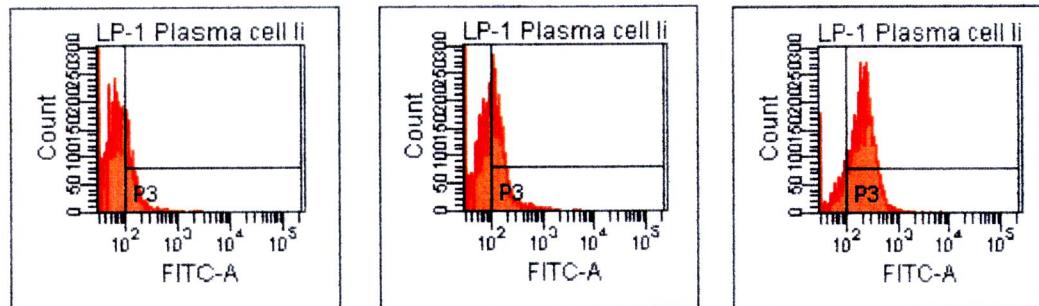
| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 9.2 | 41.1 | 82.4 |
| FITC Mean | 251 | 301 | 376 |
| FITC Median | 131 | 229 | 322 |

Triton x 100

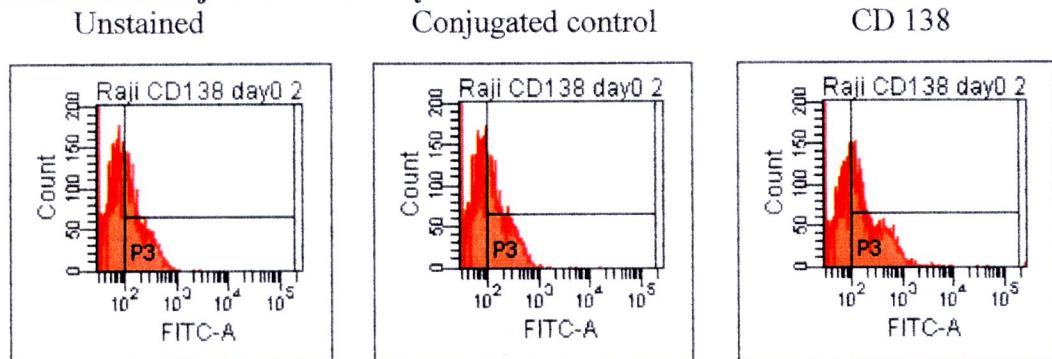
Unstained

Conjugated control

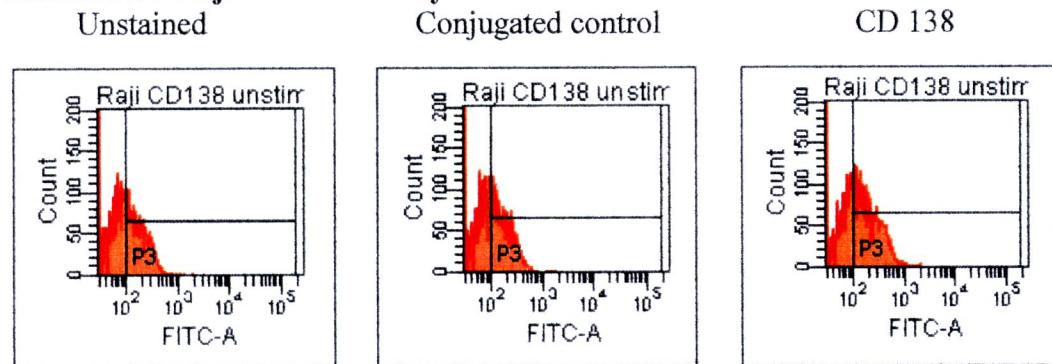
Blimp-1



| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 18.9 | 39.0 | 80.8 |
| FITC Mean | 178 | 188 | 246 |
| FITC Median | 131 | 140 | 215 |

Untstimulated Raji cell CD138 day 0

| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 39.7 | 41.0 | 51.7 |
| FITC Mean | 210 | 225 | 347 |
| FITC Median | 158 | 166 | 190 |

Unstimulated Raji cell CD138 day 3

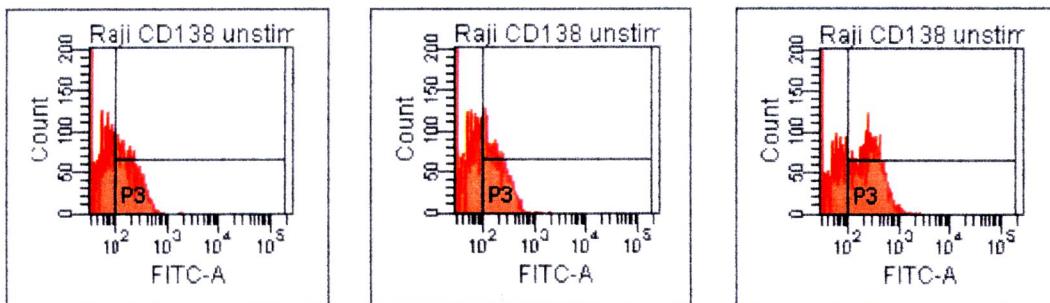
| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 42.2 | 43.4 | 57.6 |
| FITC Mean | 193 | 193 | 245 |
| FITC Median | 166 | 164 | 188 |

Unstimulated Raji cell CD138 day 6

Unstained

Conjugated control

CD 138



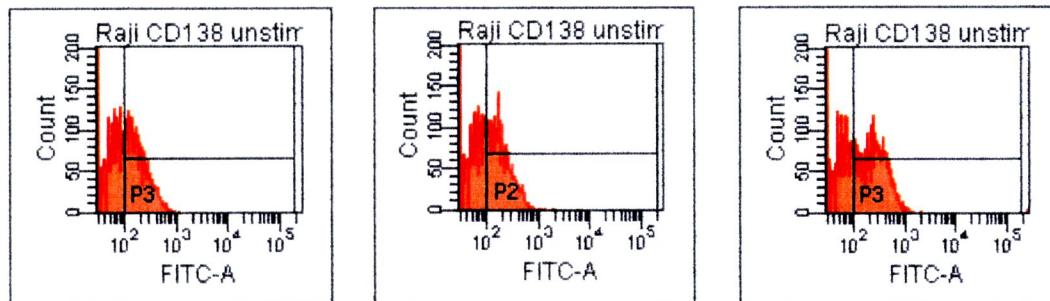
| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 38.2 | 41.5 | 56.4 |
| FITC Mean | 203 | 202 | 289 |
| FITC Median | 170 | 169 | 255 |

Unstimulated Raji cell CD138 day 9

Unstained

Conjugated control

CD 138



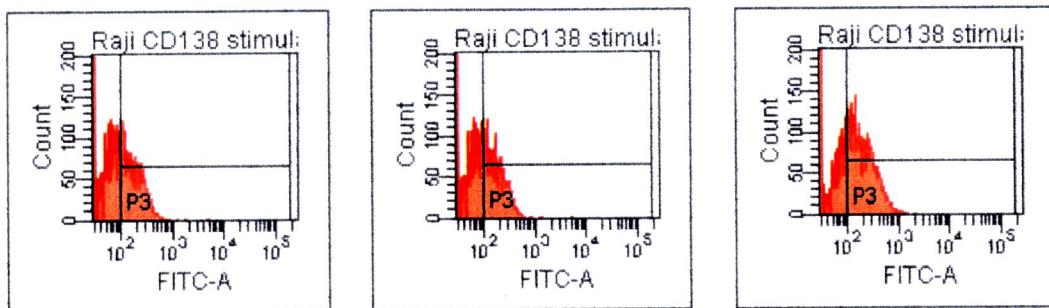
| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 44.9 | 46.3 | 52.5 |
| FITC Mean | 204 | 215 | 294 |
| FITC Median | 164 | 176 | 251 |

Stimulated Raji cell CD138 day 3

Unstained

Conjugated control

CD 138



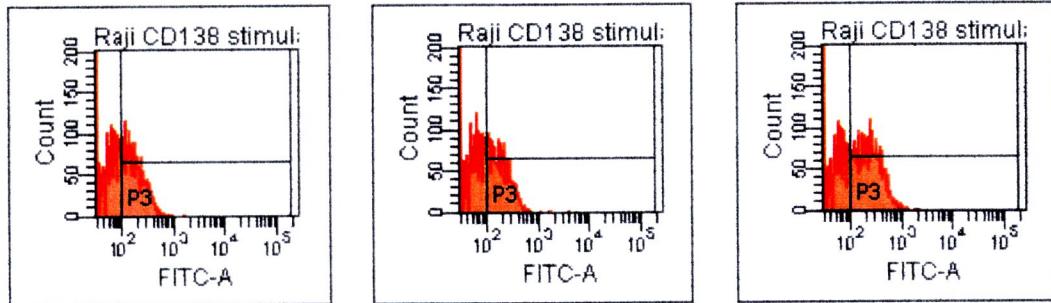
| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 41.8 | 42.4 | 52.4 |
| FITC Mean | 192 | 189 | 132 |
| FITC Median | 164 | 160 | 190 |

Stimulated Raji cell CD138 day 6

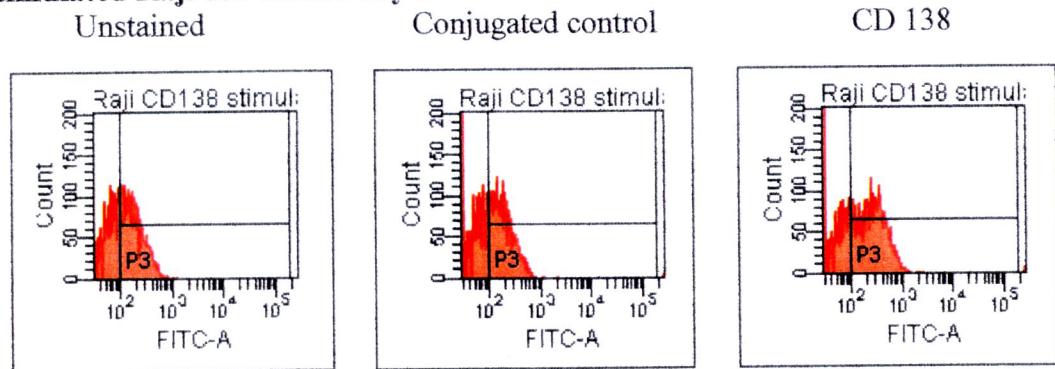
Unstained

Conjugated control

CD 138



| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 40.3 | 40.4 | 54.7 |
| FITC Mean | 195 | 200 | 263 |
| FITC Median | 166 | 172 | 219 |

Stimulated Raji cell CD138 day 9

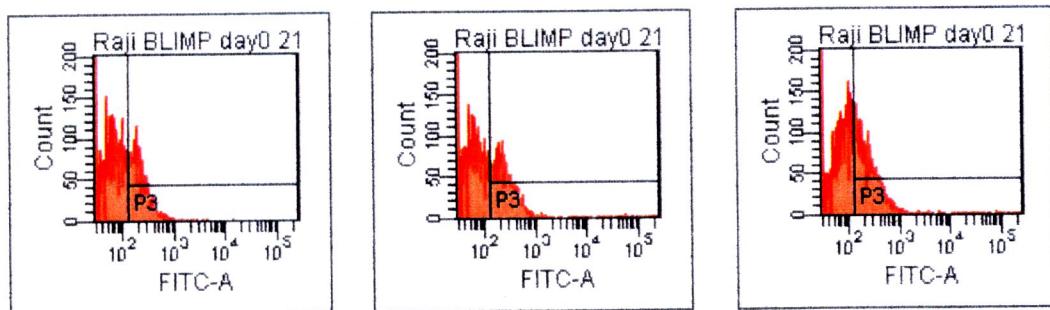
| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 46.1 | 50.0 | 60.4 |
| FITC Mean | 202 | 204 | 305 |
| FITC Median | 169 | 172 | 260 |

Unstimulated Raji cell Blimp-1 day 0

Unstained

Conjugated control

Blimp-1



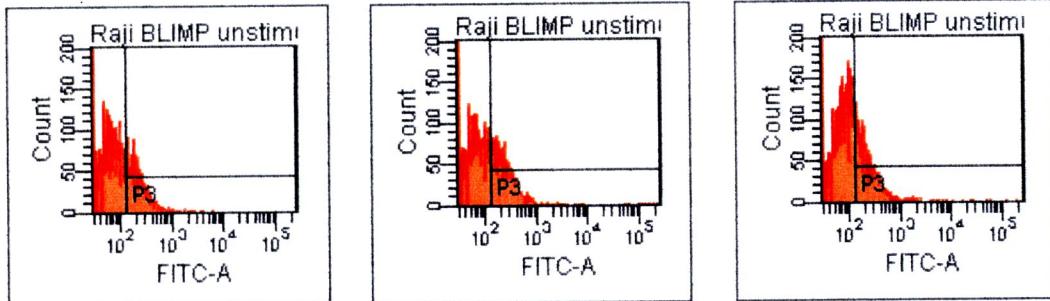
| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 28.2 | 30.4 | 40.6 |
| FITC Mean | 253 | 2015 | 1437 |
| FITC Median | 198 | 233 | 223 |

Unstimulated Raji cell Blimp-1 day 3

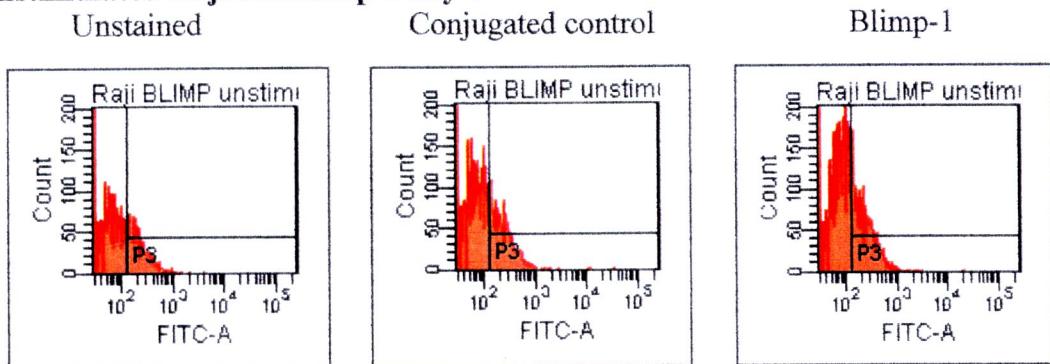
Unstained

Conjugated control

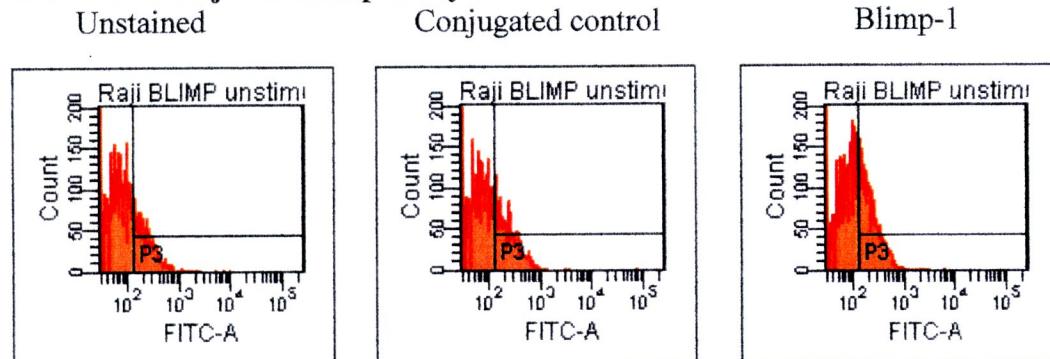
Blimp-1



| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 26.3 | 31.5 | 32.2 |
| FITC Mean | 269 | 915 | 631 |
| FITC Median | 202 | 221 | 198 |

Unstimulated Raji cell Blimp-1 day 6

| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 24.4 | 29.1 | 32.7 |
| FITC Mean | 254 | 285 | 271 |
| FITC Median | 198 | 211 | 213 |

Unstimulated Raji cell Blimp-1 day 9

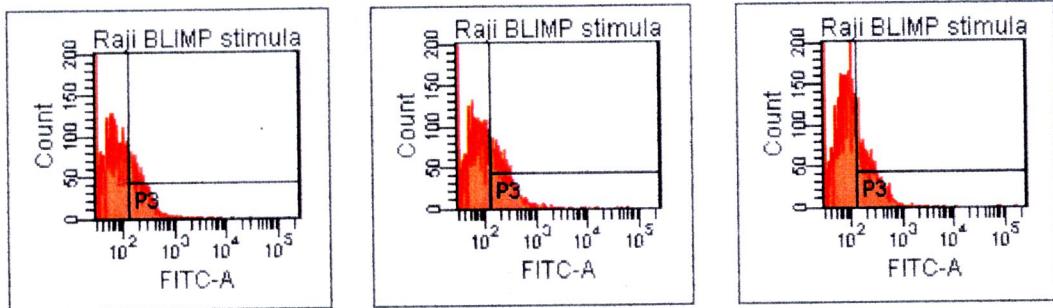
| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 22.6 | 27.2 | 37.8 |
| FITC Mean | 272 | 302 | 245 |
| FITC Median | 206 | 216 | 197 |

Stimulated Raji cell Blimp-1 day 3

Unstained

Conjugated control

Blimp-1



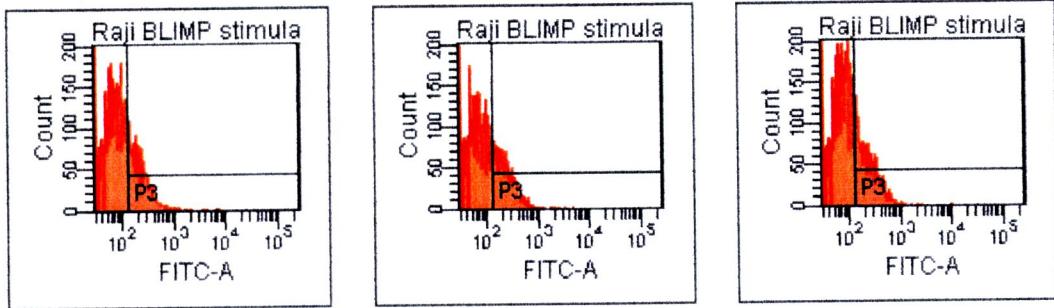
| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 24.2 | 32.5 | 28.5 |
| FITC Mean | 313 | 400 | 522 |
| FITC Median | 200 | 213 | 209 |

Stimulated Raji cell Blimp-1 day 6

Unstained

Conjugated control

Blimp-1



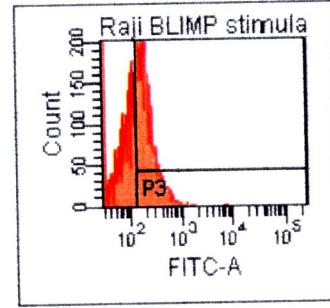
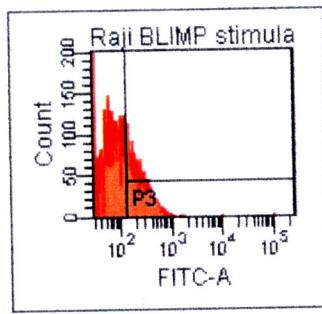
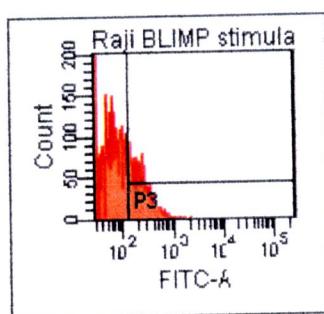
| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 25.2 | 25.1 | 27.8 |
| FITC Mean | 255 | 254 | 256 |
| FITC Median | 198 | 209 | 206 |

Stimulated Raji cell Blimp-1 day 9

Unstained

Conjugated control

Blimp-1



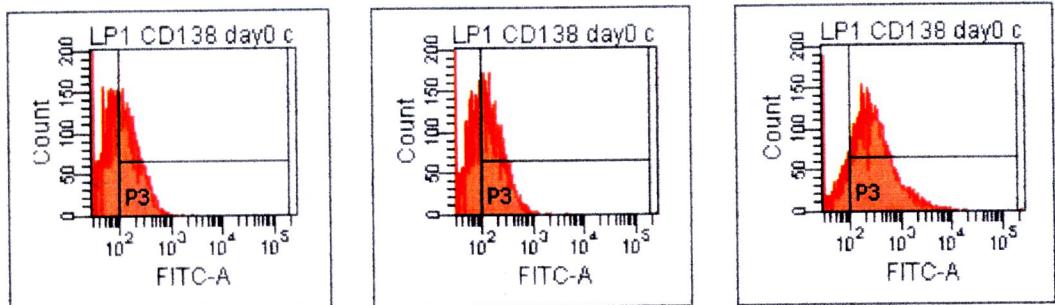
| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 24.9 | 27.8 | 54.3 |
| FITC Mean | 246 | 316 | 228 |
| FITC Median | 207 | 209 | 189 |

LP-1 cell CD138 day 0

Unstained

Conjugated control

CD 138



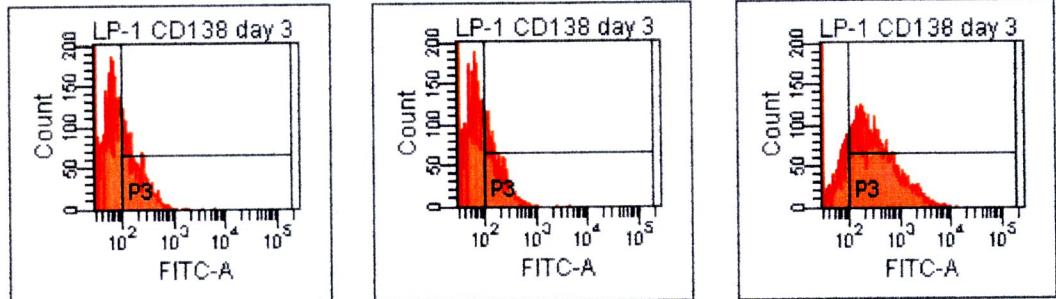
| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 45.9 | 50.6 | 83.6 |
| FITC Mean | 202 | 207 | 511 |
| FITC Median | 163 | 166 | 284 |

LP-1 cell CD138 day 3

Unstained

Conjugated control

CD 138



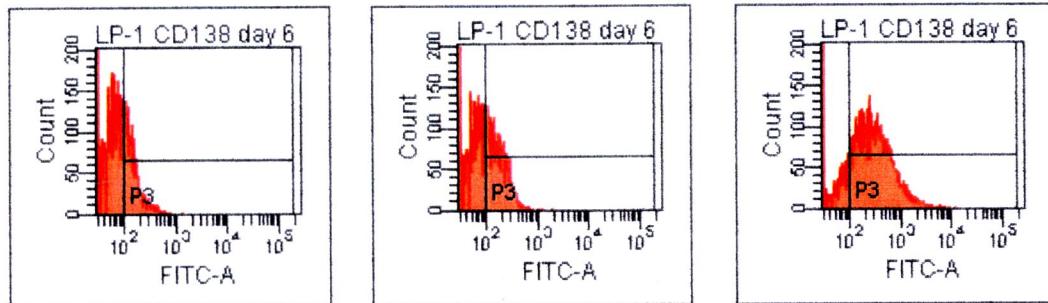
| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 31.4 | 25.3 | 74.4 |
| FITC Mean | 215 | 185 | 588 |
| FITC Median | 161 | 143 | 303 |

LP-1 cell CD138 day 6

Unstained

Conjugated control

CD 138



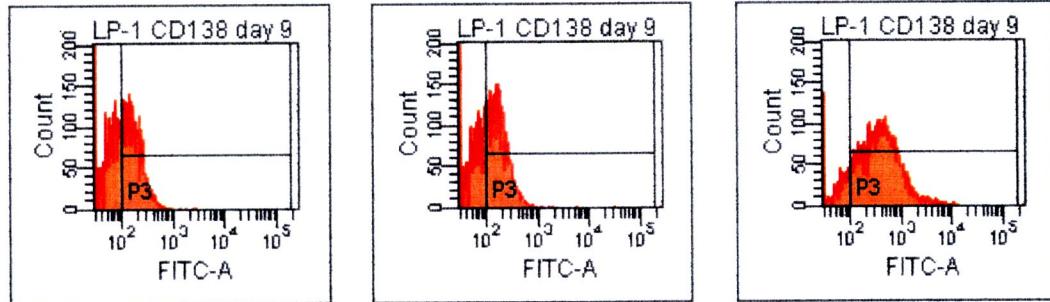
| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 25.7 | 36.8 | 81.4 |
| FITC Mean | 155 | 184 | 449 |
| FITC Median | 128 | 158 | 289 |

LP-1 cell CD138 day 9

Unstained

Conjugated control

CD 138



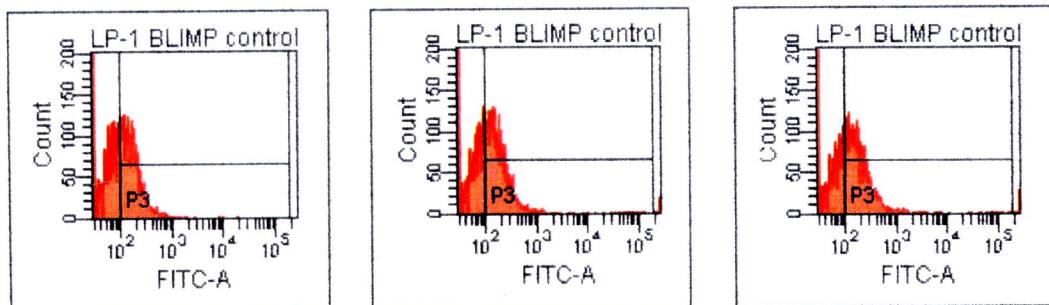
| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 46.4 | 51.2 | 86.1 |
| FITC Mean | 186 | 200 | 595 |
| FITC Median | 159 | 162 | 382 |

LP-1 cell BLIMP-1 day 0

Unstained

Conjugated control

Blimp-1



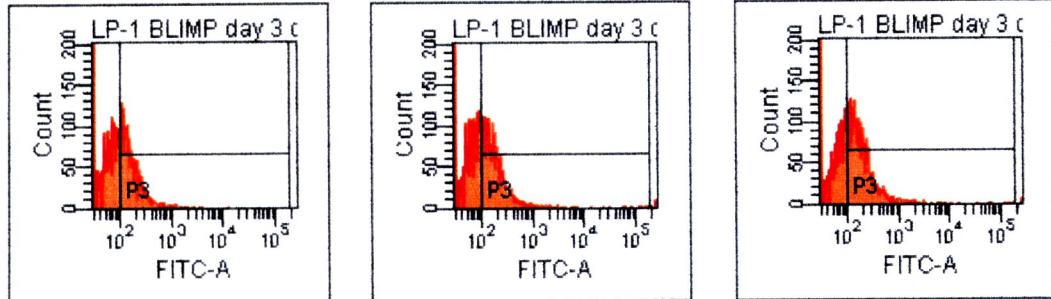
| | Unstained | Conjugated control | CD 138 |
|-------------|-----------|--------------------|--------|
| % Cell | 48.5 | 53.8 | 57.8 |
| FITC Mean | 199 | 658 | 666 |
| FITC Median | 153 | 161 | 166 |

LP-1 cell BLIMP-1 day 3

Unstained

Conjugated control

Blimp-1



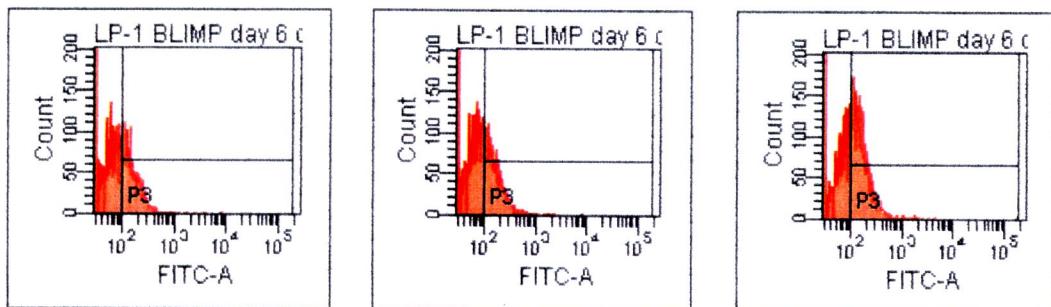
| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 44.1 | 48.6 | 56.5 |
| FITC Mean | 205 | 659 | 458 |
| FITC Median | 145 | 158 | 161 |

LP-1 cell BLIMP-1 day 6

Unstained

Conjugated control

Blimp-1



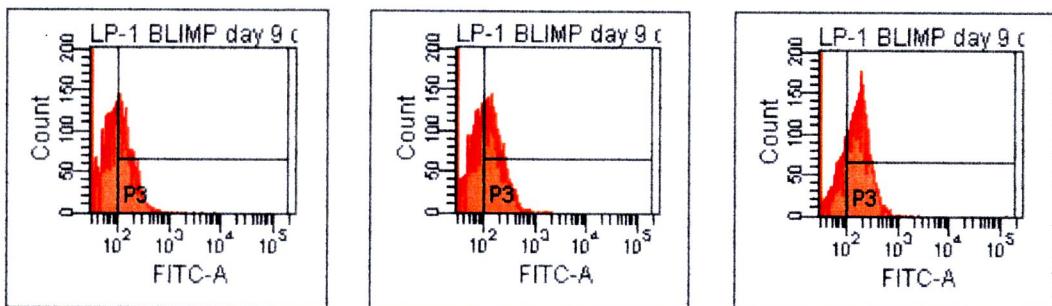
| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 32.5 | 33.8 | 50.4 |
| FITC Mean | 177 | 178 | 243 |
| FITC Median | 140 | 142 | 143 |

LP-1 cell BLIMP-1 day 9

Unstained

Conjugated control

Blimp-1



| | Unstained | Conjugated control | Blimp-1 |
|-------------|-----------|--------------------|---------|
| % Cell | 41.1 | 51.7 | 70.5 |
| FITC Mean | 179 | 187 | 241 |
| FITC Median | 142 | 153 | 178 |



CURRICULUM VITAE

Name Mr. Jedsada Kaewrakmuk

Date of Birth March 5, 1982

Education 2006: Bachelor of Science (Medical Technology),
Faculty of Associated Medical Science, Chiang Mai University,
Chiang Mai, Thailand

