

## เอกสารอ้างอิง

กาญจนา ชาญส่ง่าเวช. 2552. วิธีคัดเลือกไโรโซบีนถั่วเหลืองเพื่อผลิตเป็นปุ๋ยชีวภาพสำหรับถั่วเหลืองที่เก็บรักษาได้ที่อุณหภูมิห้อง (สถาบันทรัพย์สินทางปัจญญาแห่งจุฬาลงกรณ์มหาวิทยาลัยขอคสิทธิบัตรเมื่อวันที่ 25 มิถุนายน 2552 เลขที่คำขอคสิทธิบัตร 0901002866)

นิชานันท์ กานเกรย์ และ กาญจนา ชาญส่ง่าเวช. 2552. การใช้มัลติเพล็กซ์พีซีอาร์ทำนายศักยภาพการเข้าสร้างปมของไโรโซบีนถั่วเหลือง บทความวิจัยดีพิมพ์ในหนังสือรวมบทความการประชุมวิชาการระบบเกษตรแห่งชาติ ครั้งที่ 5 “พลังงานขาดแคลนและความมั่นคงทางอาหารเพื่อมนุษยชาติ” วันที่ 2-4 กรกฎาคม 2552 ณ โรงแรมอุบลอนเตอร์เรนชั่นแนล จ. อุบลราชธานี. หน้า 481-458.

ธันปภา จันทพีชร และ กาญจนา ชาญส่ง่าเวช. 2552. การใช้ถ่ายพิมพ์ดีอีนเอยตรวจสอบความสามารถของไโรโซบีนถั่วเหลืองในการแบ่งขั้นเข้าสร้างปมภาคสนาม บทความวิจัยดีพิมพ์ในหนังสือรวมบทความการประชุมวิชาการระบบเกษตรแห่งชาติ ครั้งที่ 5 “พลังงานขาดแคลนและความมั่นคงทางอาหารเพื่อมนุษยชาติ” วันที่ 2-4 กรกฎาคม 2552 ณ โรงแรมอุบลอนเตอร์เรนชั่นแนล จ. อุบลราชธานี. หน้า 234-243.

アナヌ ตันโซ. 2550. ระบบเกษตรในประเทศไทย : การเกษตรแบบปัจจุบันหรือเกษตรคนเม. ศูนย์ชื่อมูลเกษตรกรรมแม่โขฯ ภาควิชาทรัพยากรดินและสิ่งแวดล้อม คณะผลิตกรรมเกษตร มหาวิทยาลัยแม่โขฯ ([www.maejonaturalfarming.org](http://www.maejonaturalfarming.org)).

Abaidoo, R.C., Keyser, H.H., Singleton, P.W., Dashiell, K.E., Sanginga, N., 2007. Population size, distribution, and symbiotic characteristics of indigenous *Bradyrhizobium* spp. that nodulate TGx soybean genotypes in Africa. *Appl. Soil Ecol.* 35 : 57-67.

Aguilar, D.M., Lopez, M.V., and Riccillo, P.M. 2001. The diversity of rhizobia nodulating beans in Northwest Argentina as a source of more efficient inoculant strains. *J. Biotechnol.* 91: 181-188.

Albrecht, S.C., Maier, R.J., Hanus, F.J., Russell, S.A., Emerich, D.W. and Evans, H.J. 1979. Hydrogenase in *Rhizobium japonicum* increases nitrogen fixation by nodulated soybeans. *Science* 203 : 1255-1257.

Ando, S. and Yokoyama, T. 1999. Phylogenetic analyses of *Bradyrhizobium* strains nodulating soybean (*Glycine max*) in Thailand with reference to the USDA strains of *Bradyrhizobium*. *Can. J. Microbiol.* 45(8): 639-645.

Appunu, C., N' Zoue, A., and Laguerre, G. 2008. Genetic diversity of native bradyrhizobia isolated from soybeans (*Glycine max* L.) in different agricultural-ecological-climatic regions of India. *Appl. Environ. Microbiol.* 74(19): 5991-5996.

- Bala, A., Karanja, N., Murwira, M., Lwimbi, L., Abaidoo, R., and Giller, K. 2011. Production and use of rhizobial inoculants in Africa. [WWW.N2Africa.org](http://WWW.N2Africa.org), 21 pp.
- Brutti, L., Rivero, E., Basurco, J. C. P., Nicolas, M., Iriarte, L., Abbiati, N., Ljunggren, H. and Martensson. A. 1998. Persistence of *Bradyrhizobium japonicum* in arable soils of Argentina. *Appl. Soil Ecology*. 10: 87-94.
- Camacho, M., Santamaría, C., Temprano, F., Rodríguez-Navarro, D.N., Daza, A., Espuny, R., Bellogín, R., Ollero, F. J., Lyra de, M.C.C.P., Buendía-Clavería, A., Zhou, J., Li, F.D., Mateos, C., Velázquez, E., Vinardell, J M., and Ruiz-Sainz, J. E. 2002. Soils of the Chinese Hubei province show a very high diversity of *Sinorhizobium fredii* strains. *Syst. Appl. Microbiol.* 25(4): 592-602.
- Carter, K.R., Jennings, N.T., Hanus, J., and Evans, H.J. 1977. Hydrogen evolution and uptake by nodules of soybeans inoculated with different strains of *Rhizobium japonicum*. *Can. J. Microbiol.* 23 : 307-311.
- Chanaseni, C., and Kongngoen, S. 1992. Extension programs to promote rhizobial inoculants for soybean and groundnut in Thailand. *Can. J. Microbiol.* 38: 594-597.
- Chansa-ngavej, K. 2005. Molecular mechanisms for heat tolerance in soybean rhizobia. In. Mamoru Yamada (ed.) Death and Survival in Bacteria. Kerala : Research Signpost. India. p. 247-254.
- Chansa-ngavej, K., Chongfuengprinya, W., and Udomchotipruet, S. 2010. Soybean rhizobium technology for sustainable agriculture. Proceedings of the Technology and Innovation for Sustainable Development International Conference. March 4-6, 2010. Nongkhai, Thailand, p. 285-289.
- Chansa-ngavej. K., Ly, K.P., and Chongfuengprinya, W. 2009. Research and development for commercial production of soybean rhizobium biofertilizers with DNA fingerprints and can be kept at room temperature. Proceedings of Thailand Research Symposium 2009. p. 12-22.
- Chang, S. K. C. 2003. Protein analysis. In: Nielsen, S.S. (ed.) *Food Analysis*. 3<sup>rd</sup> Edition. Springer-Verlag. p.134-135.
- Chen, L.S., Figueiredo, A., Pedrosa, F.O., and Hungria, M. 2000. Genetic characterization of soybean rhizobia in Paraguay. *Appl. Environ. Microbiol.* 66: 5099-5103.
- Chen, W., Huang, Q., and Xiong, X. 2004. Distribution and biodiversity of soybean rhizobia in the soils of Shennongjia forest reserve, China. *Biol. Fertil. Soils*. 40: 306-312.



- Chen, W.X., Yan, G.H., and Li, I. L. 1988. Numerical taxonomic study of fast-growing soybean rhizobia and a proposal that *Rhizobium fredii* be assigned to *Sinorhizobium* gen. nov.
- Int. J. Syst. Bacteriol.* 38 :392-397
- de Jensen, C.E., Kurle, J.E., and Percich, J.A. 2004. Integrated management of edaphic and biotic factors limiting yield of irrigated soybean and dry bean in Minnesota. *Field Crops Res.* 86: 211-224.
- Dowdle, S., and Bohlool, B.B. 1985. Predominance of fast-growing *Rhizobium japonicum* in a soybean field in the People's Republic of China. *Appl. Environ. Microbiol.* 56: 1171-1176.
- Duteau, N.M., Palmer, R.G., and Atherly, A.G. 1986. Fast-growing *Rhizobium fredii* are poor nitrogen-fixing symbionts of soybean. *Crop Sci.* 26: 884-889.
- Edie, S.A. and Phillips, D.A. 1983. Effect of the host legume on acetylene reduction and hydrogen evolution by *Rhizobium* nitrogenase. *Plant Physiol.* 72(1): 156-160.
- Elkan, G.H., and Bunn, C.R. 1992. The Rhizobia. In Balows, A., Truper, H.G., Dworkin, M., Harder, W., Schleifer, K-H (eds). *The Prokaryotes. 2<sup>nd</sup> Edition. Chapter 107.* New York : Springer Verlag.
- Emampaiwong, D. 2006. Development of primers for identification of fast-or slow-growing bacteria isolated from soybean root nodules. *M.Sc. thesis, Industrial Microbiology program.* Chulalongkorn University. 90 pp.
- Fisher, R.F. and Long, S.R. 1993. Interactions of NodD at the *nod* box: Nod D binds to two distinct sites on the same face of the helix and induces a bend in the DNA. *J. Mol. Biol.* 233: 336-346.
- Fuhrmann, J. 1990. Symbiotic effectiveness of indigenous soybean bradyrhizobia as related to serological, morphological, rhizobitoxine, and hydrogenase phenotypes. *Appl. Environ. Microbiol.* 56(1) : 224-229.
- Gil-Serrano, A. M., Franco-Rodríguez, G., Tejero-Mateo, P., Thomas- Oates, J., Spaink, H. P., Ruiz-Sainz, J. E., Megías, M., and Lamrabet, Y. 1997. Structural determination of the lipo-chitin oligosaccharide nodulation signals produced by *Rhizobium fredii* HH103. *Carbohydr. Res.* 303 : 435-443.
- Goodwin, T.W. and Mercer, E.I. 2005. Introduction to Plant Biochemistry. 2<sup>nd</sup> Edition. Delhi: CBS Publishers and Distributors. 359 pp.
- Haeze, W. D., and Holsters, M. 2002. Nod factor structures, responses, and perception during initiation of nodule development. *Glycobiology.* 12 (6) : 79-105.

- Han, L.L., Wang, E.T., Han, T.X., Liu, J., Sie, X.H., Chen, W.F., and Chen, W.X. 2008. Unique community structure and biogeography of soybean rhizobia in the saline-alkaline soils of Xinjiang, China. *Plant and Soil.* 324(1-2): 291-305.
- Hungria, M., Campo, R.J., Chueire, L.M.O., Grange, L., and Megias, M. 2001. Symbiotic effectiveness of fast-growing rhizobial strains isolated from soybean nodules in Brazil. *Biol. Fertil. Soils.* 33: 387-394.
- Jitacksorn, S. 2006. Bradyoxetin and *nodD2* control nodulation efficiency of *Bradyrhizobium japonicum* in a density dependent manner. Ph.D. Dissertation. University of Minnesota, USA.
- Jordan, D.C. 1982. Transfer of *Rhizobium japonicum* Buchanan 1980 to *Bradyrhizobium* gen. nov., a genus of slow growing, root nodule bacteria from leguminous plants. *Int. J. Syst. Bacteriol.* 32 : 136-139.
- Kosslak, R. M., Bookland, R., Barkai, J., Paaren, H. E., and Applebaum, E. R., 1987. Induction of *Bradyrhizobium japonicum* common *nod* genes by isoflavones isolated from *Glycine max*. *Proc. Natl. Sci USA.* 84 : 7428-7432.
- Kuykendall, L.D., Saxena, B., Devine, T.E., and Udell, S.E. 1992. Genetic diversity in *Bradyrhizobium japonicum* Jordan 1982 and a proposal for *Bradyrhizobium elkanii* sp. nov. *Can. J. Microbiol.* 38 : 501-505.
- Lambert, G.R., Harker, A.R., Cantrell, M.A. Hanus, F.S., Russell, S.A., Haugland, R.A., and Evans, H.J. 1987. Symbiotic expression of cosmid-borne *Bradyrhizobium japonicum* hydrogenase genes. *Appl. Environ. Microbiol.* 53(2) : 422-428.
- Loh, J., Yuen-Tsai, J.P.Y., Stacey, M.G., Lohar, D., Welborn, A., and Stacey, G. 2001. Population density-dependent regulation of the *Bradyrhizobium japonicum* nodulation genes. *Mol. Microbiol.* 42(1) : 37-66.
- Loh, J., Carlson, R.W., York, W.S. and Stacey, G. 2002a. Bradyoxetin, a unique chemical signal involved in symbiotic gene regulation. *Proc. Natl. Acad. Sci. USA* 99(22) : 14446-14451.
- Loh, J., Pierson, E.A., Pierson, L.S., Stacey, G. and Chatterjee, A. 2002b. Quorum sensing in plant-associated bacteria. *Curr. Opin. Plant Biol.* 5 : 285-290.
- Loh, J. and Stacey, G. 2003. Nodulation gene regulation in *Bradyrhizobium japonicum* : a unique integration of global regulatory circuits. *Appl. Environ. Microbiol.* 69(1) : 10-17.
- Machado, D., Pueppke, S.G., Vinordel, J. M., Ruiz-Saing-S.E., and Krishnan, H.B. 1998. Expression of *nodD1* and *nodD2* in *Sinorhizobium fredii*, a nitrogen-fixing symbiont of soybean and other legumes. *Mol. Plant Microbe Interactions.* 11(5) : 375-384.

- Maruekarajtinplaeng, S. 2010. Diversity of soybean rhizobia in 16 subdistricts of Phitsanulok province. Ph.D. thesis. Microbiology program. Chulalongkorn University. 176 pp.
- Minamisawa, K., Nakatsuka, Y., and Isawa, T. 1999. Diversity and field site variation of indigenous populations of soybean bradyrhizobia in Japan by fingerprints with repeated sequences RS $\alpha$  and RS $\beta$ . *FEMS Mirobiol. Ecol.* 29: 171-178.
- Nuntagij, A., Abe, M., Uchiumi, T., Seki, Y., Boonkerd, N., and Higashi, S. 1997. Characterization of 187 *Bradyrhizobium* strains isolated from soybean cultivation in Thailand. *J. Gen. Appl. Microbiol.* 43: 183-187.
- Peng, G.X., Tan, Z.Y., Wang, E.T., Reinhold-Hurek, B., Chan, W.F., and Chen, W.X. 2002. Identification of isolates from soybean nodules in Xinjiang region as *Sinorhizobium xinjiangense* and genetic differentiation of *S. xinjiangense* from *Sinorhizobium fredii*. *Int. J. Syst. Evol. Mirobiol.* 52 : 457-462.
- Pomeranz, Y. and Meloan, C.E. 1971. Food Analysis : Theory and Practice. Westport : AVI Pub. Co. p. 632-636.
- Pueppke, S.G., Bolanos-Vasquez, M.C., Werner, D., Bec-ferte, M., Prome, J-C., and Krishnan, H. B. 1998. Release of flavonoids by the soybean cultivars McCall and Peking and their perception as signals by nitrogen-fixing symbiont *Sinorhizobium fredii*. *Plant Physiol.* 117 : 599-608.
- Rodriguez-Navarro, D.N., Oliver, I.M., Contreras, M.A., and Ruiz-Sainz, J.E. 2010. Soybean interactions with soil microbes, agronomical and molecular aspects. *Agron. Sustain. Dev.* 001 10 1051/agro/2010023. Published online September 2. 2010.
- Sambrook, J., Fritsch, E.F., and Maniatis, T. 1989. Molecular cloning : A Laboratory Manual, 2<sup>nd</sup> Edition. New York : Cold Spring Harbor Laboratory Press. Book 1. p. A3.
- Schlaman, H.R.M., Okkar, R.J.H., and Lugtenberg, B.J.J. 1992. Minireview: Regulation of nodulation gene expression by NodD in rhizobia. *J. Bacteriol.* 174(6) : 5177-5182.
- Sharma, A., Sahgal, M., and Johri, B.N. 2003. Microbial communication in the rhizosphere : operation of quorum sensing. *Curr. Sci.* 85(8) : 1164-1172.
- Somasegaran, P. and Hoben, H. 1994. Handbook for Rhizobia ; Methods in legume-*Rhizobium* Technology. p. 340, 370-1, New York : Springer Verlag.
- Stacey, G., 1995. *Bradyrhizobium japonicum* nodulation genetics. *FEMS Mirobiol. Letters.* 127 : 1-9.
- Suzuki, K., Oguro, H., Yamakawa, T., Yamamoto, A., Akao, S., and Saeki, Y. 2008. Diversity and distribution of indigenous soybean-nodulating rhizobia in the Okinawa islands, Japan. *Soil Sci. & Pl. Nutri.* 54(2): 237-246.

- Teaumroong, N. and Boonkerd, N. 1998. Detection of *Bradyrhizobium* spp. and *B. japonicum* in Thailand by primer-based technology and direct DNA extraction. *Plant and Soil.* 204: 127-134.
- Thomas-Oates, J. et al. (36 co-authors). 2003. A catalogue of molecular, physiological and symbiotic properties of soybean-nodulating rhizobial strains from different soybean cropping areas of China. *Syst. Appl. Microbiol.* 26: 453-465.
- Thompson, J.A., Bhromsiri, A., Shutsrirung, A., and Lillakan, S. 1991. Native root-nodule bacteria of traditional soybean growing areas of northern Thailand. *Plant and Soil.* 135: 53-65.
- van Berkum, P., and Saloger, C. 1991. Hydrogen oxidation by the host-controlled uptake hydrogenase phenotype of *Bradyrhizobium japonicum* in symbiosis with soybean host plants. *Appl. Environ. Microbiol.* 57(6) : 1863-1865.
- Wang, S-P., and Stacey, G. 1991. Studies of the *Bradyrhizobium japonicum nodD1* promoter : A repeated structure for the *nod* box. *J. Bacteriol.* 173 (11) : 3356-3365.
- Xu, L.M., Ge, C., Cui, Z. Li, J., and Fan, H. 1995. *Bradyrhizobium liaoningense* sp. nov. isolated from the root nodules of soybeans. *Int. J. Syst. Bacteriol.* 45 (4) : 706-711.
- Yokoyama, T., Ando, S., Murakami, T., and Imai, H. 1996. Genetic variability of the common *nod* gene in soybean bradyrhizobia isolated in Thailand and Japan. *Can. J. Microbiol.* 42: 1209-1218.
- Yokoyama, T., Ando, S., and Tsuchiya, K. 1999. Serological properties and intrinsic antibiotic resistance of soybean bradyrhizobia isolated in Thailand. *Soil Sci. Plant Nutri.* 45: 505-515.

## ภาคผนวก ก

Multiple alignments of *nodD1* of fast-growing (*S. fredii*) and slow-growing soybean rhizobia. (*B. elkanii* USDA 194 and *B. japonicum* USDA110).

	10	20	30	40	50	60	
<i>B. elkanii</i> USDA194-nodD1	1	.....	.....	.....	.....	.....	1
USDA110-nodD1	1	.....	.....	.....	.....	.....	1
<i>S. fredii</i> -nodD1	1	CGC GAGGAAC	GGC GAAAAGG	CGC GCGCGAA	GCC CTCGAT	TGTTAGTGCG	AAGGCTGTG
Chstral Consensus							60
	70	80	90	100	110	120	
<i>B. elkanii</i> USDA194-nodD1	1	.....	.....	.....	.....	.....	1
USDA110-nodD1	1	.....	.....	.....	.....	.....	1
<i>S. fredii</i> -nodD1	61	CGGTGGCGT	TGACGGTTTC	GCTCCGGCTC	GACCCCTACGG	TCGAAAAGAC	GAAATGGCAG
Chstral Consensus							120
	130	140	150	160	170	180	
<i>B. elkanii</i> USDA194-nodD1	1	.....	.....	.....	.....	.....	1
USDA110-nodD1	1	.....	.....	.....	.....	.....	1
<i>S. fredii</i> -nodD1	121	GAGCGGTGG	CCGTCTTGCC	ACAAAGCCTG	ATCTTCAGGA	GCGCAGATGTA	AGAGGGCGGG
Chstral Consensus							180
	190	200	210	220	230	240	
<i>B. elkanii</i> USDA194-nodD1	1	.....	.....	.....	.....	.....	1
USDA110-nodD1	1	.....	.....	.....	.....	.....	1
<i>S. fredii</i> -nodD1	181	CGACGGCATG	AGCCGTGAAT	GCACCTTGACT	TCAGATTAA	TAAGCGCTTT	CTAACGATT
Chstral Consensus							240
	250	260	270	280	290	300	
<i>B. elkanii</i> USDA194-nodD1	1	.....	.....	.....	.....	.....	1
USDA110-nodD1	1	.....	.....	.....	.....	.....	1
<i>S. fredii</i> -nodD1	241	GCATAATTGA	TCGTTCGGAT	GACAACCATC	CGCACTGTGG	ATTGCCAGA	ACATGCGTT
Chstral Consensus							300
	310	320	330	340	350	360	
<i>B. elkanii</i> USDA194-nodD1	1	.....	.....	.....	.....	.....	1
USDA110-nodD1	1	.....	.....	.....	.....	.....	1
<i>S. fredii</i> -nodD1	301	TAAGGGCCTT	GATCTCAATC	TCCTCGTTGC	GCTCGACGCA	CTGATGACCG	AACGCAAAC
Chstral Consensus							360
	370	380	390	400	410	420	
<i>B. elkanii</i> USDA194-nodD1	1	.....	CTA	GCAGCGCC	CGCTTG	GTTTCC	TGCAGATCCC
USDA110-nodD1	1	.....	CTA	ACAGCGCC	CTG	GCTCTTG	CCCAGATGCC
<i>S. fredii</i> -nodD1	361	CACGGCGCGT	GCACGCCAGC	TCAACCTGAG	CGACCCGCCG	ATGAGCAG	CCATCACCG
Chstral Consensus							420
	430	440	450	460	470	480	
<i>B. elkanii</i> USDA194-nodD1	44	ATGTTGGA	CGCCCTCTCG	AACCAA	TATC	CGCGGCATCC	AAATGCTCGC
USDA110-nodD1	44	ATGTTGGA	TGCTCTCG	AGCRAA	TATC	CGADGCATCC	AGATGCTCGC
<i>S. fredii</i> -nodD1	421	GCTTCGAGC	TATTGCGG	ACGAGCTATT	TACCATGAAT	GGTCGGGAAC	TTGTACAAAC
Chstral Consensus							480
	490	500	510	520	530	540	
<i>B. elkanii</i> USDA194-nodD1	100	ATTGTTGGAAC	GCAGCGCCAC	TGCACTGGCCT	CGGTAAGAAGT	GCGAATGGGG	AGTGGTGGCT
USDA110-nodD1	100	ATTGTTGGAAT	GAGCGCCAC	TGCAAGGGCCT	CTGTTGAATGT	GGGGAAAGGGGG	AGGGGGCGGTT
<i>S. fredii</i> -nodD1	481	TCCGCAGAGCA	GAAGCGCTCG	CAACCGCAGT	CGCGGAGGCC	CTGGTTCACAA	TCCATCTCTC
Chstral Consensus							540
	550	560	570	580	590	600	
<i>B. elkanii</i> USDA194-nodD1	159	CGACGATCG	CAGGGGAA	TCCACTTTTC	GAAGTGTCTG	GCTAAGACGTA	AGGGCATTTGT
USDA110-nodD1	159	GGATGATCG	CAACGGGA	TCCGCTTTTC	GAAGTGTCTG	GCCGCTCGT	AGGGCATCTGT
<i>S. fredii</i> -nodD1	541	CATCATTTAA	TGGGATCGT	TCAACCCAGC	GCAGTCAGAT	CGCGATTTCA	GGATCATTC
Chstral Consensus							600
	610	620	630	640	650	660	
<i>B. elkanii</i> USDA194-nodD1	217	TCTCT	ATGCGGCCG	TGTCGAAACAA	TATGGGCGGGA	ATCAGGCTAA	AGCCCTGCA
USDA110-nodD1	217	GCCG	ATACGGCTCG	TGTCCTAGCAA	CAGGGGCGGGA	ATCAGGCTAA	AGCCCTGCA
<i>S. fredii</i> -nodD1	601	TTCGACTTC	ATGACGCTAA	TGTTTTTGA	AAGGGTCTGTG	GTGAGAGCTGG	CGGGGAAGC
Chstral Consensus							660
	670	680	690	700	710	720	
<i>B. elkanii</i> USDA194-nodD1	271	AACGACCTCG	ATGCGCTTT	TCAAGCGCTG	CTGGAGCAA	--GAACCACT	CCTCGAGGTT
USDA110-nodD1	271	GACGACCTCA	ATTCGCTCTCC	TCAAGCGCTG	CTCAAGCAA	--AAACCAATT	CTTCGAGGTT
<i>S. fredii</i> -nodD1	661	GCGCGCGGTC	AGTTTCGAGT	TGCTCGCGTT	TTCGATGAG	CCAGATGAGC	TTCTCCGGCG
Chstral Consensus							720
	730	740	750	760	770	780	
<i>B. elkanii</i> USDA194-nodD1	328	GGGCTCTAGC	GGCCGTCGA	ACTTGGCGGT	GACGTGCCC	ATGGAATTGT	ATTTCTCGAA
USDA110-nodD1	328	GGGTCTCAGT	GGCGGTCGA	ACTTGGCAAGT	AACGTGCCC	ATCGAGATGT	ATTGTTCGAA
<i>S. fredii</i> -nodD1	721	TGGTGTAGTC	GGTTCTGCTGA	TCTTACCAAGA	AATGTTATG	TCGGACACAC	ATC-CCAGAG
Chstral Consensus							780

Bellanii-USDA194-nodD1	388	T G T A A G T G G T	790	C G C - A A T A G C	388	T G C T T G T T G	790	T G C G G C A T C C	388	C A C G C A T A C G	790	A G E G T C T C A T	388	C G T - A G C T G C	790	C G C G G C A T C C	388	G A C G C A T A C G	790	A G E G T C T C A T	446
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					
Bellanii-USDA194-nodD1	447	C G A A T A G C G T	850	C G C C T T A G G A	447	T G C G C G C T C G	850	A C A T G A A C A A	447	T T C C G G C A A A	850	A T G A G A A A A A T	447	C G A A C A G C G T	850	T G C C G C G C A G A	447	A T G A G A A A A A G T	850	A T G A G A A A A A G T	506
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					174
Bellanii-USDA194-nodD1	507	C C A C T T C G C C	910	G G G G C G A A G C	507	A - G C T C A C C G	910	G G T T C A T - C G	507	G A A A A T E G C A	910	G C A A T T C G A A	507	C G A C C T C G C C	910	C C G C C G C A G C	507	G A A A A T E G C A	910	G C A A T T C G A A	564
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					195
Bellanii-USDA194-nodD1	565	G C G C A U U G C A	970	G G C G C T C T C T	565	G T G C G C A T A D G	970	A T C A A C A A T T	565	C T C C G A R A A A A	970	A . . . . . C A C	565	G C G C A C C G C G	970	G G G G C T T C T T	565	G T G C G C A T A D G	970	A . . . . . C A C	618
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					211
Bellanii-USDA194-nodD1	619	G A T T G T C A T G	1010	A A A T C G G G A G A	619	G A A T C G G G A G A	1010	A C C C T C A C C G	619	T G A A G C G T C G	1010	G C T C G A C A B G	619	G T C G G G T C G A	1010	T G C G C T C A T G	619	G C A G G G T C G A	1010	G C A G G G T C G A	677
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					1077
Bellanii-USDA194-nodD1	678	A G G C A T C C C G	1090	A G A A A T G A T C	678	G A G A G T T G G A	1090	T G T G C A G C A G	678	G G C - - - C T C G	1090	C G A A C C C G G A C	678	G C A A C C C G G A C	1090	G C A A C C C G G A C	678	G C A A C C C G G A C	1090	G C A A C C C G G A C	734
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					1130
Bellanii-USDA194-nodD1	735	C C G C A A G C G C	1150	C T C C G C G C G	735	G G T G T C G G G A	1160	C G A G C T C G C G	735	A C C T C T C A T C	1160	G T A A A C A A T T	735	C T G C A A G C G C	1150	T T C C G C G C C A	735	G C A G G G T C G A	1160	G C A G G G T C G A	794
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					281
Bellanii-USDA194-nodD1	795	C A T C G C G A A A	1210	G T A G C T G G G C	795	A G C C G C G - - -	1210	G A T - - - G C A T	795	C G C A G C G C T C	1210	A T G G C G G G G G	795	T G G C T C A T A D A	1210	A T G G C G G G G G	795	T G G C T C A T A D A	1210	A T G G C G G G G G	844
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					300
Bellanii-USDA194-nodD1	845	T G G C T C - - - A G	1270	G T T A A T T T G G	845	G G C G C G G G G	1280	C T G T - - - G A G G T	845	T G C A G C G C T C	1280	A T G G C G G G G G	845	T G G C T C A T A D A	1270	A T G G C G G G G G	845	T G G C T C A T A D A	1270	A T G G C G G G G G	897
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					1309
Bellanii-USDA194-nodD1	898	A G G C A T C G G G	1330	G C A A C G A - - - G	898	A A G G T T T A G A	1340	T G A A G T C C C T	898	T G A A C C G G A T	1340	- - - - -	898	G G C G T C G A G C	1330	G C A A C C G G A C	898	G G C G T C G A G C	1330	G C A A C C G G A C	945
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					1369
Bellanii-USDA194-nodD1	945	.....	1390	.....	1400	.....	1410	.....	1420	.....	1430	.....	1440	.....	.....	.....	.....	.....	.....	.....	945
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					1429
Bellanii-USDA194-nodD1	945	.....	1450	.....	1460	.....	1470	.....	1480	.....	1490	.....	1500	.....	.....	.....	.....	.....	.....	.....	945
USDA110-nodD1																					
Sfreddii-nodD1																					
<i>Chastal Consensus</i>																					1464

### หมายเหตุ

- เครื่องหมาย \* แสดงตำแหน่งนิวคลีโอไทด์ที่เหมือนกันในไรโซบีโยนถ่วงเพิ่มจำนวนช้า
- ผลการทำ multiple alignments พบลำดับนิวคลีโอไทด์ต่างกันของ nodD1 ของไรโซบีโยนถ่วงเพิ่มจำนวนเร็วและเพิ่มจำนวนช้า



