

REFERENCES



- Alberts B, Johnson A, Lewis J, Raff M, Roberts K, Walter P. Cyclic-AMP-dependent Protein Kinase (PKA) Mediates Most of the Effects of Cyclic AMP. *Molecular biology of the cells* 2002.
- Alvaro D, Mancino MG, Onori P, Franchitto A, Alpini G, Francis H, et al. Estrogens and the pathophysiology of the biliary tree. *World J Gastroenterol* 2006; 12: 3537-45.
- Amieux PS, McKnight GS. The essential role of RI alpha in the maintenance of regulated PKA activity. *Ann N Y Acad Sci* 2002; 968: 75-95.
- Ashida K, Terada T, Kitamura Y, Kaibara N. Expression of E-cadherin, alpha-catenin, beta-catenin, and CD44 (standard and variant isoforms) in human cholangiocarcinoma: an immunohistochemical study. *Hepatology* 1998; 27: 974-82.
- Beebe SJ, Corbin J. In: The Enzymes: Control by Phosphorylation, Part A. *Academic Press: New York* 1986; 17: 43-111.
- Bhola NE, Grandis JR. Crosstalk between G-protein-coupled receptors and epidermal growth factor receptor in cancer. *Front Biosci* 2008; 13: 1857-65.
- Bockmann S, Nebe B. The in vitro effects of H-89, a specific inhibitor of protein kinase A, in the human colonic carcinoma cell line Caco-2. *Eur J Cancer Prev* 2003; 12: 469-78.
- Bogorad RL, Ostroukhova TY, Orlova AN, Rubtsov PM, Smirnova OV. Long isoform of prolactin receptor predominates in rat intrahepatic bile ducts and further increases under obstructive cholestasis. *J Endocrinol* 2006; 188: 345-54.
- Bold RJ, Alpard S, Ishizuka J, Townsend CM, Jr., Thompson JC. Growth-regulatory effect of gastrin on human colon cancer cell lines is determined by protein kinase a isoform content. *Regul Pept* 1994; 53: 61-70.
- Bradbury AW, Carter DC, Miller WR, Cho-Chung YS, Clair T. Protein kinase A (PK-A) regulatory subunit expression in colorectal cancer and related mucosa. *Br J Cancer* 1994; 69: 738-42.

- Carlson CC, Chinery R, Burnham LL, Dransfield DT. 8-Cl-adenosine-induced inhibition of colorectal cancer growth in vitro and in vivo. *Neoplasia* 2000; 2: 441-8.
- Chen HX, Marshall JL, Ness E, Martin RR, Dvorchik B, Rizvi N, et al. A safety and pharmacokinetic study of a mixed-backbone oligonucleotide (GEM231) targeting the type I protein kinase A by two-hour infusions in patients with refractory solid tumors. *Clin Cancer Res* 2000; 6: 1259-66.
- Cho YS, Cho-Chung YS. Antisense protein kinase A RIalpha acts synergistically with hydroxycamptothecin to inhibit growth and induce apoptosis in human cancer cells: molecular basis for combinatorial therapy. *Clin Cancer Res* 2003; 9: 1171-8.
- Cho YS, Kim MK, Cheadle C, Neary C, Becker KG, Cho-Chung YS. Antisense DNAs as multisite genomic modulators identified by DNA microarray. *Proc Natl Acad Sci U S A* 2001; 98: 9819-23.
- Cho-Chung YS. Role of cyclic AMP receptor proteins in growth, differentiation, and suppression of malignancy: new approaches to therapy. *Cancer Res* 1990; 50: 7093-100.
- Cho-Chung YS, Clair T, Shepherd C. Anticarcinogenic effect of N6,O2'-dibutyl cyclic adenosine 3':5'-monophosphate on 7,12-dimethylbenz(a)anthracene mammary tumor induction in the rat and its relationship to cyclic adenosine 3':5'-monophosphate metabolism and protein kinase. *Cancer Res* 1983; 43: 2736-40.
- Cho-Chung YS, Clair T, Tagliaferri P, Ally S, Katsaros D, Tortora G, et al. Site-selective cyclic AMP analogs as new biological tools in growth control, differentiation, and proto-oncogene regulation. *Cancer Invest* 1989; 7: 161-77.
- Cho-Chung YS, Nesterova M, Pepe S, Lee GR, Noguchi K, Srivastava RK, et al. Antisense DNA-targeting protein kinase A-RIA subunit: a novel approach to cancer treatment. *Front Biosci* 1999; 4: D898-907.
- Cho-Chung YS, Pepe S, Clair T, Budillon A, Nesterova M. cAMP-dependent protein kinase: role in normal and malignant growth. *Crit Rev Oncol Hematol* 1995; 21: 33-61.

- Chung JY, Hong SM, Choi BY, Cho H, Yu E, Hewitt SM. The expression of phospho-AKT, phospho-mTOR, and PTEN in extrahepatic cholangiocarcinoma. *Clin Cancer Res* 2009; 15: 660-7.
- Ciardiello F, Damiano V, Bianco C, di Isernia G, Ruggiero A, Caraglia M, et al. Cooperative antiproliferative effects of 8-chloro-cyclic AMP and 528 anti-epidermal growth factor receptor monoclonal antibody on human cancer cells. *Clin Cancer Res* 1995; 1: 161-7.
- Ciardiello F, Damiano V, Bianco R, Bianco C, Fontanini G, De Laurentiis M, et al. Antitumor activity of combined blockade of epidermal growth factor receptor and protein kinase A. *J Natl Cancer Inst* 1996; 88: 1770-6.
- Clegg CH, Cadd GG, McKnight GS. Genetic characterization of a brain-specific form of the type I regulatory subunit of cAMP-dependent protein kinase. *Proc Natl Acad Sci U S A* 1988; 85: 3703-7.
- Davies SP, Reddy H, Caivano M, Cohen P. Specificity and mechanism of action of some commonly used protein kinase inhibitors. *Biochem J* 2000; 351: 95-105.
- de Groen PC, Gores GJ, LaRusso NF, Gunderson LL, Nagorney DM. Biliary tract cancers. *N Engl J Med* 1999; 341: 1368-78.
- Dechakhamphu S, Yongvanit P, Nair J, Pinlaor S, Sithithaworn P, Bartsch H. High excretion of etheno adducts in liver fluke-infected patients: protection by praziquantel against DNA damage. *Cancer Epidemiol Biomarkers Prev* 2008; 17: 1658-64.
- Dell'Acqua ML, Scott JD. Protein kinase A anchoring. *J Biol Chem* 1997; 272: 12881-4.
- Doval DC, Sekhon JS, Gupta SK, Fuloria J, Shukla VK, Gupta S, et al. A phase II study of gemcitabine and cisplatin in chemotherapy-naive, unresectable gall bladder cancer. *Br J Cancer* 2004; 90: 1516-20.
- Dumont JE, Jauniaux JC, Roger PP. The cyclic AMP-mediated stimulation of cell proliferation. *Trends Biochem Sci* 1989; 14: 67-71.
- Elkins DB, Mairiang E, Sithithaworn P, Mairiang P, Chaiyakum J, Chamadol N, et al. Cross-sectional patterns of hepatobiliary abnormalities and possible precursor conditions of cholangiocarcinoma associated with *Opisthorchis viverrini* infection in humans. *Am J Trop Med Hyg* 1996; 55: 295-301.

- Farrow B, Rychahou P, Murillo C, O'Connor K L, Iwamura T, Evers BM. Inhibition of pancreatic cancer cell growth and induction of apoptosis with novel therapies directed against protein kinase A. *Surgery* 2003; 134: 197-205.
- Faux MC, Scott JD. More on target with protein phosphorylation: conferring specificity by location. *Trends Biochem Sci* 1996; 21: 312-5.
- Francis H, Glaser S, Ueno Y, Lesage G, Marucci L, Benedetti A, et al. cAMP stimulates the secretory and proliferative capacity of the rat intrahepatic biliary epithelium through changes in the PKA/Src/MEK/ERK1/2 pathway. *J Hepatol* 2004; 41: 528-37.
- Francis SH, Corbin JD. Cyclic nucleotide-dependent protein kinases: intracellular receptors for cAMP and cGMP action. *Crit Rev Clin Lab Sci* 1999; 36: 275-328.
- Gonzalez GA, Yamamoto KK, Fischer WH, Karr D, Menzel P, Biggs W, 3rd, et al. A cluster of phosphorylation sites on the cyclic AMP-regulated nuclear factor CREB predicted by its sequence. *Nature* 1989; 337: 749-52.
- Gu YY, Zhang HY, Zhang HJ, Li SY, Ni JH, Jia HT. 8-Chloro-adenosine inhibits growth at least partly by interfering with actin polymerization in cultured human lung cancer cells. *Biochem Pharmacol* 2006; 72: 541-50.
- Haswell-Elkins MR, Mairiang E, Mairiang P, Chaiyakum J, Chamadol N, Loapaiboon V, et al. Cross-sectional study of *Opisthorchis viverrini* infection and cholangiocarcinoma in communities within a high-risk area in northeast Thailand. *Int J Cancer* 1994; 59: 505-9.
- Hebenstreit D, Horejs-Hoeck J, Duschl A. JAK/STAT-dependent gene regulation by cytokines. *Drug News Perspect* 2005; 18: 243-9.
- Huether A, Hopfner M, Baradari V, Schuppan D, Scherubl H. Sorafenib alone or as combination therapy for growth control of cholangiocarcinoma. *Biochem Pharmacol* 2007; 73: 1308-17.
- Jadeski LC, Hum KO, Chakraborty C, Lala PK. Nitric oxide promotes murine mammary tumour growth and metastasis by stimulating tumour cell migration, invasiveness and angiogenesis. *Int J Cancer* 2000; 86: 30-9.

- Jankovic D, Pesic M, Markovic J, Kanazir S, Markovic I, Rakic L, et al. The combination of sulfinosine and 8-Cl-cAMP induces synergistic cell growth inhibition of the human neuroblastoma cell line in vitro. *Invest New Drugs* 2006; 24: 15-25.
- Jiao X, Katiyar S, Willmarth NE, Liu M, Ma X, Flomenberg N, et al. c-Jun induces mammary epithelial cellular invasion and breast cancer stem cell expansion. *J Biol Chem*; 285: 8218-26.
- Khan SA, Thomas HC, Davidson BR, Taylor-Robinson SD. Cholangiocarcinoma. *Lancet* 2005; 366: 1303-14.
- Kim SN, Kim SG, Park JH, Lee MA, Park SD, Cho-Chung YS, et al. Dual anticancer activity of 8-Cl-cAMP: inhibition of cell proliferation and induction of apoptotic cell death. *Biochem Biophys Res Commun* 2000; 273: 404-10.
- Krebs EG. Protein kinases. *Curr Top Cell Regul* 1972; 5: 99-133.
- Krebs EG, Beavo JA. Phosphorylation-dephosphorylation of enzymes. *Annu Rev Biochem* 1979; 48: 923-59.
- Krishan A, Arya P. Monitoring of cellular resistance to cancer chemotherapy. *Hematol Oncol Clin North Am* 2002; 16: 357-72, vi.
- Lawlor MA, Alessi DR. PKB/Akt: a key mediator of cell proliferation, survival and insulin responses? *J Cell Sci* 2001; 114: 2903-10.
- Ledinko N, Chan IJ. Increase in type I cyclic adenosine 3':5'-monophosphate-dependent protein kinase activity and specific accumulation of type I regulatory subunits in adenovirus type 12-transformed cells. *Cancer Res* 1984; 44: 2622-7.
- Li M, West JW, Numann R, Murphy BJ, Scheuer T, Catterall WA. Convergent regulation of sodium channels by protein kinase C and cAMP-dependent protein kinase. *Science* 1993; 261: 1439-42.
- Liu AY. Differentiation-specific increase of cAMP-dependent protein kinase in the 3T3-L1 cells. *J Biol Chem* 1982; 257: 298-306.
- Lochner A, Moolman JA. The many faces of H89: a review. *Cardiovasc Drug Rev* 2006; 24: 261-74.

- Loilome W, Yongvanit P, Wongkham C, Tepsiri N, Sripa B, Sithithaworn P, et al. Altered gene expression in *Opisthorchis viverrini*-associated cholangiocarcinoma in hamster model. *Mol Carcinog* 2006; 45: 279-87.
- Lonning PE. Study of suboptimum treatment response: lessons from breast cancer. *Lancet Oncol* 2003; 4: 177-85.
- Malik IA, Aziz Z, Zaidi SH, Sethuraman G. Gemcitabine and Cisplatin is a highly effective combination chemotherapy in patients with advanced cancer of the gallbladder. *Am J Clin Oncol* 2003; 26: 174-7.
- Mantovani G, Bondioni S, Lania AG, Rodolfo M, Peverelli E, Polentarutti N, et al. High expression of PKA regulatory subunit 1A protein is related to proliferation of human melanoma cells. *Oncogene* 2008; 27: 1834-43.
- Mao LM, Tang Q, Wang JQ. Protein kinase C-regulated cAMP response element-binding protein phosphorylation in cultured rat striatal neurons. *Brain Res Bull* 2007; 72: 302-8.
- McDaid HM, Cairns MT, Atkinson RJ, McAleer S, Harkin DP, Gilmore P, et al. Increased expression of the RI α subunit of the cAMP-dependent protein kinase A is associated with advanced stage ovarian cancer. *Br J Cancer* 1999; 79: 933-9.
- Miller WR, Watson DM, Jack W, Chetty U, Elton RA. Tumour cyclic AMP binding proteins: an independent prognostic factor for disease recurrence and survival in breast cancer. *Breast Cancer Res Treat* 1993; 26: 89-94.
- Mimeault M, Pommery N, Henichart JP. Synergistic antiproliferative and apoptotic effects induced by epidermal growth factor receptor and protein kinase a inhibitors in human prostatic cancer cell lines. *Int J Cancer* 2003; 106: 116-24.
- Neary CL, Nesterova M, Cho YS, Cheadle C, Becker KG, Cho-Chung YS. Protein kinase A isozyme switching: eliciting differential cAMP signaling and tumor reversion. *Oncogene* 2004; 23: 8847-56.
- Nesterova M, Cho-Chung YS. A single-injection protein kinase A-directed antisense treatment to inhibit tumour growth. *Nat Med* 1995; 1: 528-33.

- Nesterova MV, Cho-Chung YS. Antisense protein kinase A RIalpha inhibits 7, 12-dimethylbenz(a)anthracene-induction of mammary cancer: blockade at the initial phase of carcinogenesis. *Clin Cancer Res* 2004; 10: 4568-77.
- Ohtsuka M, Ito H, Kimura F, Shimizu H, Togawa A, Yoshidome H, et al. Extended hepatic resection and outcomes in intrahepatic cholangiocarcinoma. *J Hepatobiliary Pancreat Surg* 2003; 10: 259-64.
- Olnes MJ, Erlich R. A review and update on cholangiocarcinoma. *Oncology* 2004; 66: 167-79.
- Papazisis KT, Geromichalos GD, Dimitriadis KA, Kortsaris AH. Optimization of the sulforhodamine B colorimetric assay. *J Immunol Methods* 1997; 208: 151-8.
- Patel T. Increasing incidence and mortality of primary intrahepatic cholangiocarcinoma in the United States. *Hepatology* 2001; 33: 1353-7.
- Pinlaor S, Ma N, Hiraku Y, Yongvanit P, Semba R, Oikawa S, et al. Repeated infection with *Opisthorchis viverrini* induces accumulation of 8-nitroguanine and 8-oxo-7,8-dihydro-2'-deoxyguanine in the bile duct of hamsters via inducible nitric oxide synthase. *Carcinogenesis* 2004; 25: 1535-42.
- Pollack A, Bae K, Khor LY, Al-Saleem T, Hammond ME, Venkatesan V, et al. The importance of protein kinase A in prostate cancer: relationship to patient outcome in Radiation Therapy Oncology Group trial 92-02. *Clin Cancer Res* 2009; 15: 5478-84.
- Ramage AD, Langdon SP, Ritchie AA, Burns DJ, Miller WR. Growth inhibition by 8-chloro cyclic AMP of human HT29 colorectal and ZR-75-1 breast carcinoma xenografts is associated with selective modulation of protein kinase A isoenzymes. *Eur J Cancer* 1995; 31A: 969-73.
- Roberson ED, English JD, Adams JP, Selcher JC, Kondratick C, Sweatt JD. The mitogen-activated protein kinase cascade couples PKA and PKC to cAMP response element binding protein phosphorylation in area CA1 of hippocampus. *J Neurosci* 1999; 19: 4337-48.
- Robinson-White AJ, Hsiao HP, Leitner WW, Greene E, Bauer A, Krett NL, et al. Protein kinase A-independent inhibition of proliferation and induction of apoptosis in human thyroid cancer cells by 8-Cl-adenosine. *J Clin Endocrinol Metab* 2008; 93: 1020-9.

- Roesler WJ, Vandenbark GR, Hanson RW. Cyclic AMP and the induction of eukaryotic gene transcription. *J Biol Chem* 1988; 263: 9063-6.
- Rohlf C, Clair T, Cho-Chung YS. 8-Cl-cAMP induces truncation and down-regulation of the RI alpha subunit and up-regulation of the RII beta subunit of cAMP-dependent protein kinase leading to type II holoenzyme-dependent growth inhibition and differentiation of HL-60 leukemia cells. *J Biol Chem* 1993; 268: 5774-82.
- Sathornsumetee S, Reardon DA, Desjardins A, Quinn JA, Vredenburgh JJ, Rich JN. Molecularly targeted therapy for malignant glioma. *Cancer* 2007; 110: 13-24.
- Sawyers C. Targeted cancer therapy. *Nature* 2004; 432: 294-7.
- Scala S, Budillon A, Zhan Z, Cho-Chung YS, Jefferson J, Tsokos M, et al. Downregulation of *mdr-1* expression by 8-Cl-cAMP in multidrug resistant MCF-7 human breast cancer cells. *J Clin Invest* 1995; 96: 1026-34.
- Scheid MP, Woodgett JR. PKB/AKT: functional insights from genetic models. *Nat Rev Mol Cell Biol* 2001; 2: 760-8.
- Schwartz DA, Rubin CS. Regulation of cAMP-dependent protein kinase subunit levels in Friend erythroleukemic cells. Effects of differentiation and treatment with 8-Br-cAMP and methylisobutyl xanthine. *J Biol Chem* 1983; 258: 777-84.
- Scott JD. Cyclic nucleotide-dependent protein kinases. *Pharmacol Ther* 1991; 50: 123-45.
- Settakorn J, Kaewpila N, Burns GF, Leong AS. FAT, E-cadherin, beta catenin, HER 2/neu, Ki67 immuno-expression, and histological grade in intrahepatic cholangiocarcinoma. *J Clin Pathol* 2005; 58: 1249-54.
- Shaib Y, El-Serag HB. The epidemiology of cholangiocarcinoma. *Semin Liver Dis* 2004; 24: 115-25.
- Shaib YH, Davila JA, McGlynn K, El-Serag HB. Rising incidence of intrahepatic cholangiocarcinoma in the United States: a true increase? *J Hepatol* 2004; 40: 472-7.
- Skalhegg BS, Tasken K. Specificity in the cAMP/PKA signaling pathway. Differential expression, regulation, and subcellular localization of subunits of PKA. *Front Biosci* 2000; 5: D678-93.

- Skehan P, Storeng R, Scudiero D, Monks A, McMahon J, Vistica D, et al. New colorimetric cytotoxicity assay for anticancer-drug screening. *J Natl Cancer Inst* 1990; 82: 1107-12.
- Sugimachi K, Taguchi K, Aishima S, Tanaka S, Shimada M, Kajiyama K, et al. Altered expression of beta-catenin without genetic mutation in intrahepatic cholangiocarcinoma. *Mod Pathol* 2001; 14: 900-5.
- Tannock IF. Tumor physiology and drug resistance. *Cancer Metastasis Rev* 2001; 20: 123-32.
- Taylor SS, Buechler JA, Yonemoto W. cAMP-dependent protein kinase: framework for a diverse family of regulatory enzymes. *Annu Rev Biochem* 1990; 59: 971-1005.
- Thamavit W, Bhamarapravati N, Sahaphong S, Vajrasthira S, Angsubhakorn S. Effects of dimethylnitrosamine on induction of cholangiocarcinoma in *Opisthorchis viverrini*-infected Syrian golden hamsters. *Cancer Res* 1978; 38: 4634-9.
- Thanan R, Murata M, Pinlaor S, Sithithaworn P, Khuntikeo N, Tangkanakul W, et al. Urinary 8-oxo-7,8-dihydro-2'-deoxyguanosine in patients with parasite infection and effect of antiparasitic drug in relation to cholangiocarcinogenesis. *Cancer Epidemiol Biomarkers Prev* 2008; 17: 518-24.
- Todoroki T. Chemotherapy for bile duct carcinoma in the light of adjuvant chemotherapy to surgery. *Hepatogastroenterology* 2000; 47: 644-9.
- Tokumoto N, Ikeda S, Ishizaki Y, Kurihara T, Ozaki S, Iseki M, et al. Immunohistochemical and mutational analyses of Wnt signaling components and target genes in intrahepatic cholangiocarcinomas. *Int J Oncol* 2005; 27: 973-80.
- Tortora G, Budillon A, Yokozaki H, Clair T, Pepe S, Merlo G, et al. Retroviral vector-mediated overexpression of the RII beta subunit of the cAMP-dependent protein kinase induces differentiation in human leukemia cells and reverts the transformed phenotype of mouse fibroblasts. *Cell Growth Differ* 1994; 5: 753-9.

- Tortora G, Caputo R, Damiano V, Bianco R, Pepe S, Bianco AR, et al. Synergistic inhibition of human cancer cell growth by cytotoxic drugs and mixed backbone antisense oligonucleotide targeting protein kinase A. *Proc Natl Acad Sci USA* 1997; 94: 12586-91.
- Tortora G, Cho-Chung YS. Type II regulatory subunit of protein kinase restores cAMP-dependent transcription in a cAMP-unresponsive cell line. *J Biol Chem* 1990; 265: 18067-70.
- Tortora G, Ciardiello F. Protein kinase A as target for novel integrated strategies of cancer therapy. *Ann N Y Acad Sci* 2002; 968: 139-47.
- Tortora G, Ciardiello F. Targeting of epidermal growth factor receptor and protein kinase A: molecular basis and therapeutic applications. *Ann Oncol* 2000; 11: 777-83.
- Tortora G, Damiano V, Bianco C, Baldassarre G, Bianco AR, Lanfranccone L, et al. The RIalpha subunit of protein kinase A (PKA) binds to Grb2 and allows PKA interaction with the activated EGF-receptor. *Oncogene* 1997; 14: 923-8.
- Tortora G, di Isernia G, Sandomenico C, Bianco R, Pomatico G, Pepe S, et al. Synergistic inhibition of growth and induction of apoptosis by 8-chloro-cAMP and paclitaxel or cisplatin in human cancer cells. *Cancer Res* 1997; 57: 5107-11.
- Tortora G, Yokozaki H, Pepe S, Clair T, Cho-Chung YS. Differentiation of HL-60 leukemia by type I regulatory subunit antisense oligodeoxynucleotide of cAMP-dependent protein kinase. *Proc Natl Acad Sci USA* 1991; 88: 2011-5.
- Uhler MD, McKnight GS. Expression of cDNAs for two isoforms of the catalytic subunit of cAMP-dependent protein kinase. *J Biol Chem* 1987; 262: 15202-7.
- Uttaravichien T, Bhudhisawasdi V, Pairojkul C, Pugkhem A. Intrahepatic cholangiocarcinoma in Thailand. *J Hepatobiliary Pancreat Surg* 1999; 6: 128-35.
- Vatanasapt V, Sriamporn S, Vatanasapt P. Cancer control in Thailand. *Jpn J Clin Oncol* 2002; 32 Suppl: S82-91.

- Vucic V, Niciforovic A, Adzic M, Radojic MB, Ruzdijic S. The combination of gamma ionizing radiation and 8-Cl-cAMP induces synergistic cell growth inhibition and induction of apoptosis in human prostate cancer cells. *Invest New Drugs* 2008; 26: 309-17.
- Wang HM, Zheng NG, Wu JL, Gong CC, Wang YL. Dual effects of 8-Br-cAMP on differentiation and apoptosis of human esophageal cancer cell line Eca-109. *World J Gastroenterol* 2005; 11: 6538-42.
- West KA, Brognard J, Clark AS, Linnoila IR, Yang X, Swain SM, et al. Rapid Akt activation by nicotine and a tobacco carcinogen modulates the phenotype of normal human airway epithelial cells. *J Clin Invest* 2003; 111: 81-90.
- Yasui W, Tahara E. Effect of gastrin on gastric mucosal cyclic adenosine 3':5'-monophosphate-dependent protein kinase activity in rat stomach carcinogenesis induced by N-methyl-N'-nitro-N-nitrosoguanidine. *Cancer Res* 1985; 45: 4763-7.
- Yokomuro S, Tsuji H, Lunz JG, 3rd, Sakamoto T, Ezure T, Murase N, et al. Growth control of human biliary epithelial cells by interleukin 6, hepatocyte growth factor, transforming growth factor beta1, and activin A: comparison of a cholangiocarcinoma cell line with primary cultures of non-neoplastic biliary epithelial cells. *Hepatology* 2000; 32: 26-35.
- Young MR, Montpetit M, Lozano Y, Djordjevic A, Devata S, Matthews JP, et al. Regulation of Lewis lung carcinoma invasion and metastasis by protein kinase A. *Int J Cancer* 1995; 61: 104-9.

APPENDICES

APPENDIX A

Reagents for laboratory experiments

Reagents for RNA extraction

1. 70% Ethanol	Final conc.	Volume (ml)
Absolute ethanol	70%	35
DW		15

Mix the solution and store at room temperature.

2. Diethylpyrocarbonate (DEPC) treated water	Final conc.	Volume (ml)
DEPC (MW 162.141)	0.1%	1
DW		999

Mix well for at least 1 hr, let the solution stand at room temperature for an overnight then subject to autoclave, and stored at room temperature or aliquots store at -20 °C for longer time.

Reagents for reverse transcription (RT) reaction

1. RT solution I (freshly prepared)	Final amount	Volume (μl)
Total RNA	2.5 μg	11.5
Random hexamer (0.5 μg/μl)	0.5 μg	1

Mix the solution then spin down and incubated at 70 °C for 10 min.

After incubation, keep it on ice. Quickly spin down the solution and keep on ice.

2. RT solution II (freshly prepared)	Final conc.	Volume (μl)
5X RT buffer	20%	4
RNase inhibitor (RNasin [®] ; 40U/μl)	1U/μl	0.5
10 mM dNTP mix	0.5 mM	1
MMuLV (reverse transcriptase)	10U/μl	1
RT solution I		12.5
	Total volume	20

Reagents for TaqMan real time RT-PCR

1. Reaction mixture	Final conc.	Volume (μ l)
2X TaqMan [®] Universal PCR Master Mix (ABI)	50%	10
20X Gene Expression Assay Mix (each target genes; ABI)	5%	1
cDNA	25%	5
DEPC treated water		4
	Total volume	20

Reagents for SDS-PAGE and Western Blotting

1. 30% Acrylamide solution (30% Acrylamide, 0.8% Bis)	Final conc.	Amount
Acrylamide (MW 71.08)	30%	300 g
Bis (N, N,-methylenebisacrylamind, MW 154.17)	0.8%	8 g
Distilled water	up to	1000 ml

Dissolve the component in distilled water and adjust volume (using volumetric flask) upto 1000 ml or 500 ml. Filter through filter paper (Number 1 or 2_ low grade) and store in dark bottle (for light protection). Store at 4°C.

2. 1.5 M Tris-HCL pH 8.8	Final conc.	Amount
Tris (MW 121.14)	1.5 M	181.7 g

Dissolve the component in 800 ml distilled water and adjust pH to 8.8* by using Conc.HCl (~ 20 ml). Then adjust volume upto 1000 ml. Store at 4°C.

3. 0.5 M Tris-HCL pH 6.8	Final conc.	Amount
Tris (MW 121.14)	0.5 M	60.57 g

Dissolve the component in 800 ml distilled water and adjust pH to 6.8* by using Conc.HCl (~ 20 ml). Then adjust volume upto 1000 ml. Store at 4°C

4. 10% (w/v) SDS solution	Final conc.	Amount
SDS (MW 288.38)	10%	5 g
Distilled water	up to	50 ml

Filter with 0.22 μ m filter and keep at room temperature.

5. 10% Ammonium persulfate (APS) (Freshly prepare)	Final conc.	Amount
Ammonium persulfate (MW 71.08)	10%	0.5 g
Distilled water	up to	5 ml

6. Water-saturated buthanol	Final conc.	Volume
N, I or t- buthanol	-	50 ml
Distilled water	-	10 ml

Shake well and store at room temperature.

7. 10 x SDS buffer for electrophoresis	Final conc.	Volume
Trisma base (MW 121.14)	250 mM	30.25 g
Glycine (MW 75.07)	1.92 M	144.10 g
SDS (MW 288.38)	1.0 % (w/v)	10 g
Distilled water	up to	1000 ml

Shake well and store at room temperature

8. 4x SDS loading buffer (4x sample buffer)	Final conc.	Volume
1M Tris- HCL pH 6.8	0.25 M	2.5 ml
SDS	8%	0.8 g
Glycerol	40%	4 ml
2-mercaptoethanol	20%	2 ml
Distilled water	up to 10 ml	< 1.5 ml

Dissolve all of components and add bromophenol blue 0.0001 g (estimate)

Dilute the solution with PBS to 3x, 2x and 1x SDS loading buffer. Aliquot and keep at - 20 °C.

	Final conc.	Volume
9. TBS		
1 M Tris – HCL pH 7.4	30 ml	10 ml
5 M NaCl	60 ml	20 ml
Distilled water	60 ml	1000 ml
Shake well and store at room temperature		

	Final conc.	Volume
10. TBS-T (0.1 % Tween 20 in TBS)		
TBS		500 ml
Tween-20	0.1%	500 μ l
Shake well and store at room temperature		

	Final conc.	Volume
11. Tris-Glycine buffer		
Tris (MW 121.14)	25 mM	3 g
Glycine	192 mM	14.4 g
DW	-	600 ml
Methanol	10%	100 ml
Distilled water	up to	1000 ml
Shake well and store at room temperature		

	Final conc.	Volume
12. 5 % skim milk in TBS (Freshly prepare)		
Skim milk	5%	1.5 g
TBS		30 ml
Shake well and store at room temperature		

	Final conc.	Volume
13. 0.1% CBB (Coomasie Brilliant Blue)		
0.2% CBB stock solution	0.1%	200 ml
20% Acetic acid	10%	200 ml
Shake well and store at room temperature		

14. Destain solution	Final conc.	Volume
Ethanol	25%	750 ml
Acetic acid	8%	240 ml
Distilled water	up to	3000 ml

Shake well and store at room temperature

Reagent for cell lysis buffer

1. RIPA buffer	Final conc.	Volume
1 M Tris-HCl (pH 8.0)	20 mM	0.2 ml
0.5 M EDTA	5 mM	0.1 ml
5 M NaCl	150 mM	0.3 ml
10% NP-40	1%	1 ml
10% SDS	0.025%	0.025 ml
1 M NaF	10 mM	0.1 ml
0.2 M NaVO ₄	2 mM	0.1 ml
Proteinase inhibitor cocktail (100X)	1X	0.1 ml
Distilled water	up to	10 ml

Shake well and store at -20 °C

Reagent for Protein kinase A assay

1. PKA extraction buffer	Final conc.	Volume
1M Tris-HCl (pH 7.4)	25 mM	2.5 ml
EDTA (MW 372.24)	0.5 mM	19 mg
EGTA (MW 380.35)	0.5 mM	19 mg
β-mercaptoethanol	10 mM	0.07 ml
Leupeptin (2.5 mg/ml)	1 µg/ml	0.04 ml
Aprotinin (1 mg/ml)	1 µg/ml	0.1 ml
Distilled water	up to	100 ml

Store at -20 °C. Just before use, add 0.5 ml of PMSF stock solution (100 mM PMSF in

2. PKA dilution buffer	Final conc.	Volume
K ₂ PO ₄ (pH 6.8) (MW 212.27)	350 mM	3.7 g
DTT (MW 154.24)	0.1 mM	0.75 mg
Distilled water	up to	50 ml

Shake well and store at room temperature
100% ethanol) per 100 ml of PKA extraction buffer.

3. 50 mM Tris-HCl (pH 8.0)	Final conc.	Volume
Tris (MW 121.14)	50 mM	6.057 g

Dissolve the component in 800 ml distilled water and adjust pH to 8.0* by using Conc.HCl. Then adjust volume up to 1000 ml. Store at 4°C

Reagent for Sulforhodamin B assay

1. 10% w/v TCA	Final conc.	Amount
Trichloroacetic acid	10%	5 g
DW	up to	50 ml

Store at 4 °C

2. 0.4% Sulforhodamine B in 1% acetic acid	Final conc.	Amount
Sulforhodamine B	0.4%	0.2 g
1% acetic acid		50 ml

3. 1% acetic acid	Final conc.	Volume
100% acetic acid	1%	10 ml
DW	up to	1000 ml

4. 10 mM unbuffered Tris base pH 10.5	Final conc.	Amount
Trisma-base (MW 121.1)	10 mM	121.14 mg

Dissolve the component in 60 ml distilled water and adjust pH to 6.8* by using Conc.HCl. Then adjust volume up to 100 ml. Store at room temperature.

Reagent for cell culture

1. Ham's F12 media	Final conc.	Amount
Ham's F12 Nutrient mixture (Gibco)		10.6 g
Solid NaHCO ₃	14 mM	1.176 g
Sterile distilled water		900 ml
Adjust pH to 7.3-7.4 by using 1N HCl		
Penicillin/streptomycin		10 ml
Fetal bovine serum		100 ml
Stored at 4 °C		

2. 1X Phosphate buffer saline (PBS)	Final conc.	Amount
NaHPO ₄	12 mM	1.48 g
KH ₂ PO ₄	3 mM	0.43 g
NaCl	0.12 M	7.2 g
Adjust pH to 7.4		
DW	up to	1000 ml
Autoclave at 121°C, 15 psi for 20 min and keep at 4 °C		

APPENDIX B
Supplementary data

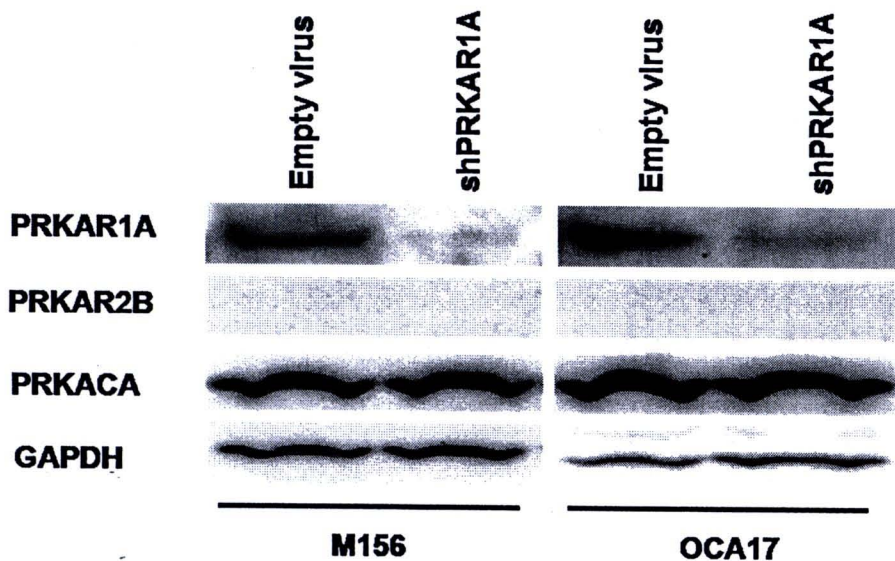


Figure B.1 Effect of shPRKAR1A on PRKACA expression.

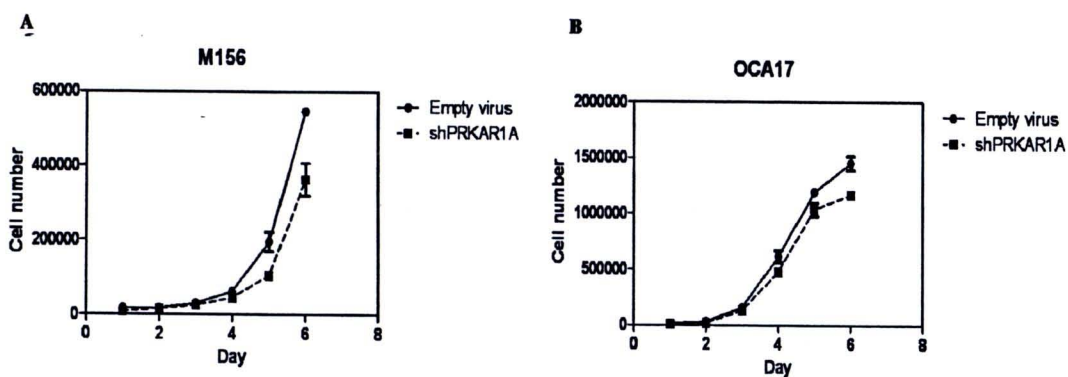


Figure B.2 Growth curve of PRKAR1A silencing cells in M156 (A) and OCA17 (B). The number of cells was determined by cell counting at indicated time. Each point represents average values from triplicates.

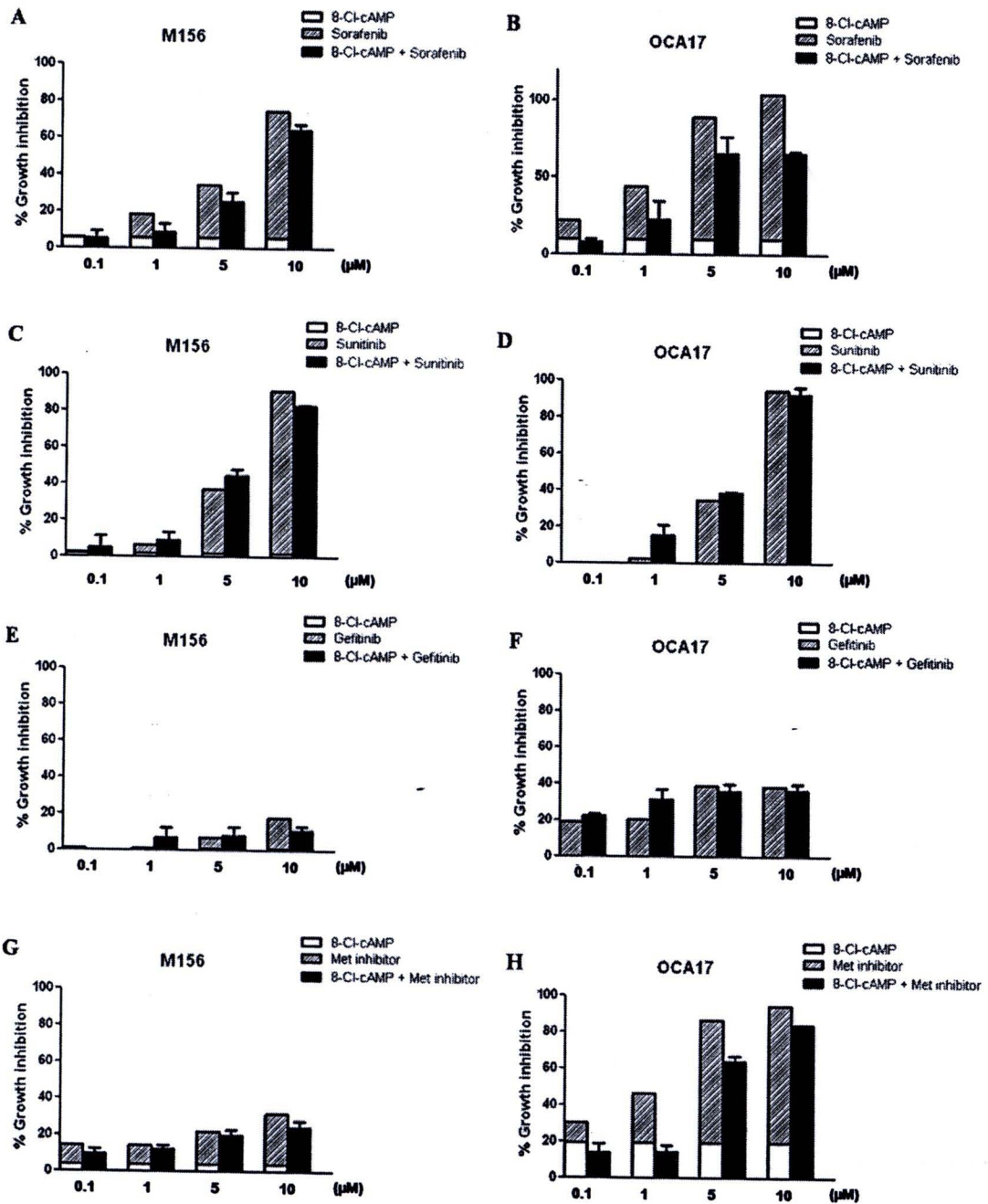


Figure B.3 Antiproliferative effect of 8-Cl-cAMP and protein kinase inhibitors in CCA cell lines. Cells were treated with 100 μM 8-Cl-cAMP plus different concentration of sorafenib (A and B), sunitinib (C and D), gefitinib (E and F) and Met inhibitor (G and H) for 72 h.

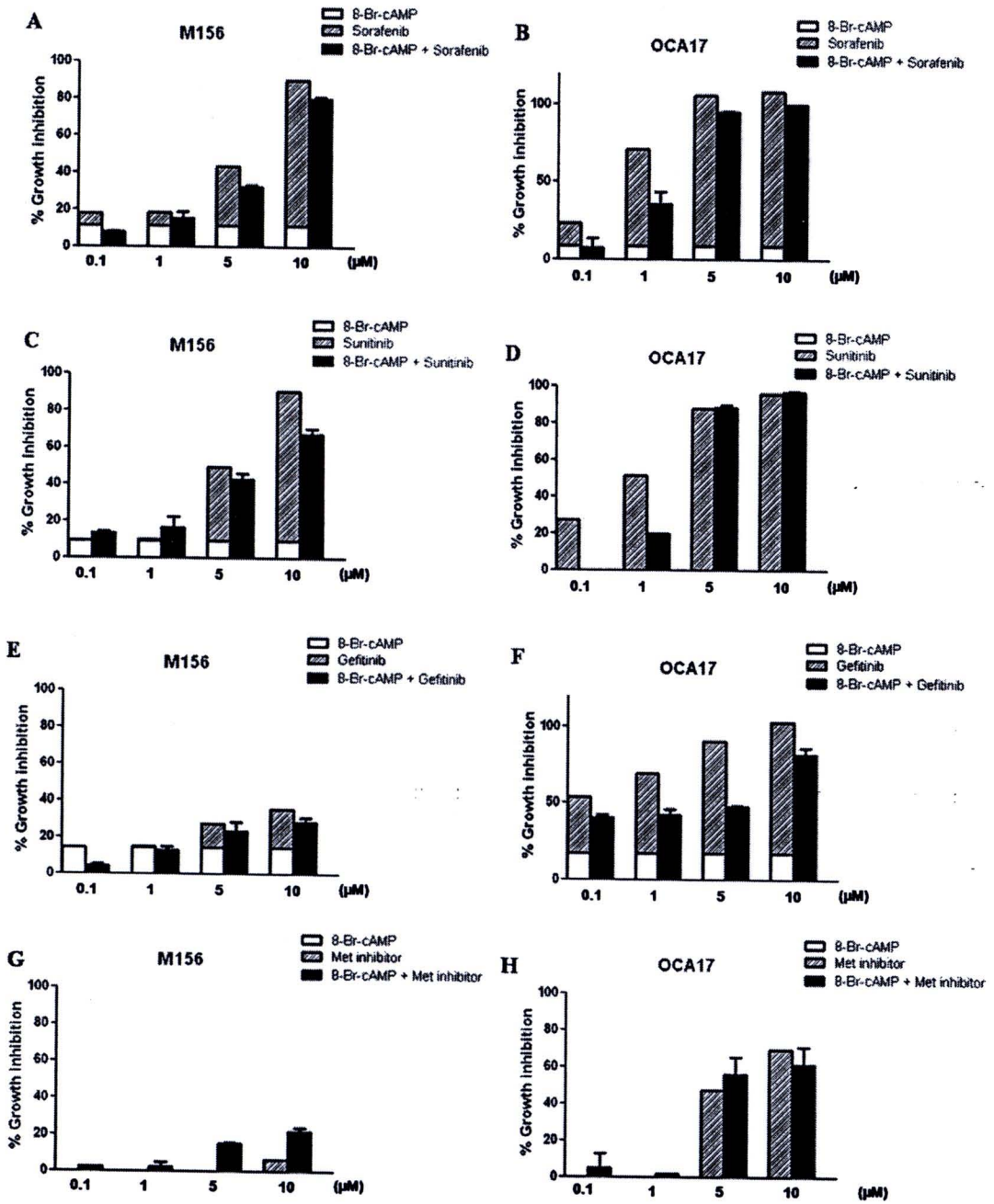


Figure B.4 Antiproliferative effect of 8-Br-cAMP and protein kinase inhibitors in CCA cell lines. Cells were treated with 100 μM 8-Br-cAMP plus different concentration of sorafenib (A and B), sunitinib (C and D), gefitinib (E and F) and Met inhibitor (G and H) for 72 h.

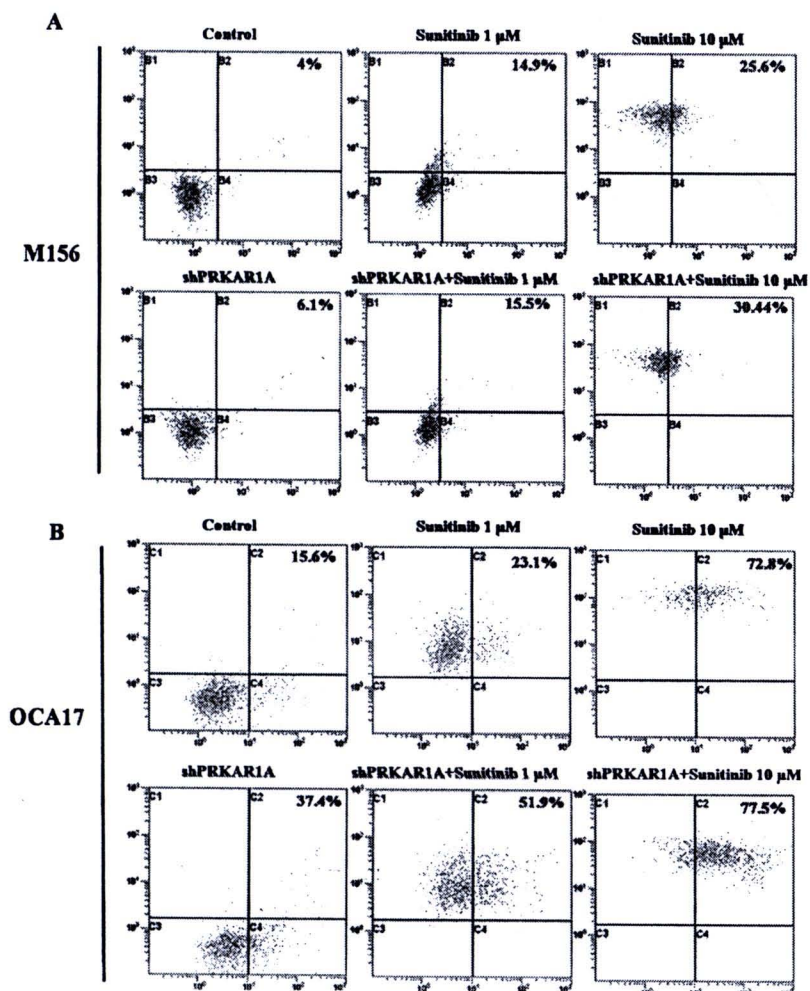


Figure B.5 Effect of shPRKAR1A in combination with 1 μ M and 10 μ M sunitinib on induction of apoptosis in M156(A) and OCA17(B). Annexin-V and PI (propidium iodide) staining was performed, followed by flow cytometry.

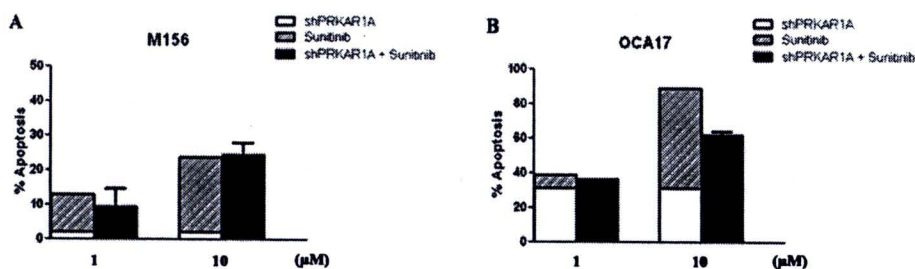


Figure B.6 Percentage of apoptosis of shPRKAR1A in combination with sunitinib in M156(A) and OCA17(B). Annexin-V and PI (propidium iodide) staining was performed, followed by flow cytometry.

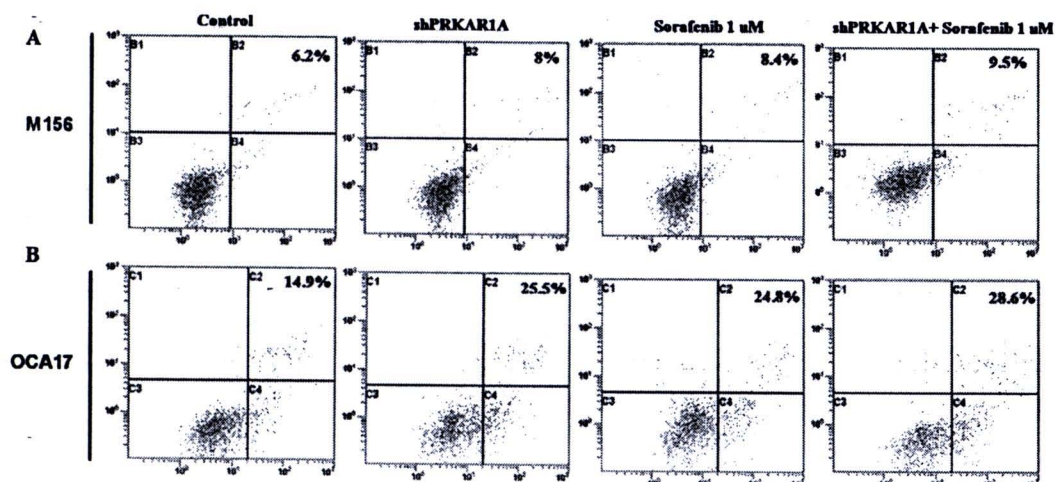


Figure B.7 Effect of shPRKAR1A in combination with 1 μM sorafenib on induction of apoptosis in M156(A) and OCA17(B). Annexin-V and PI (propidium iodide) staining was performed, followed by flow cytometry.

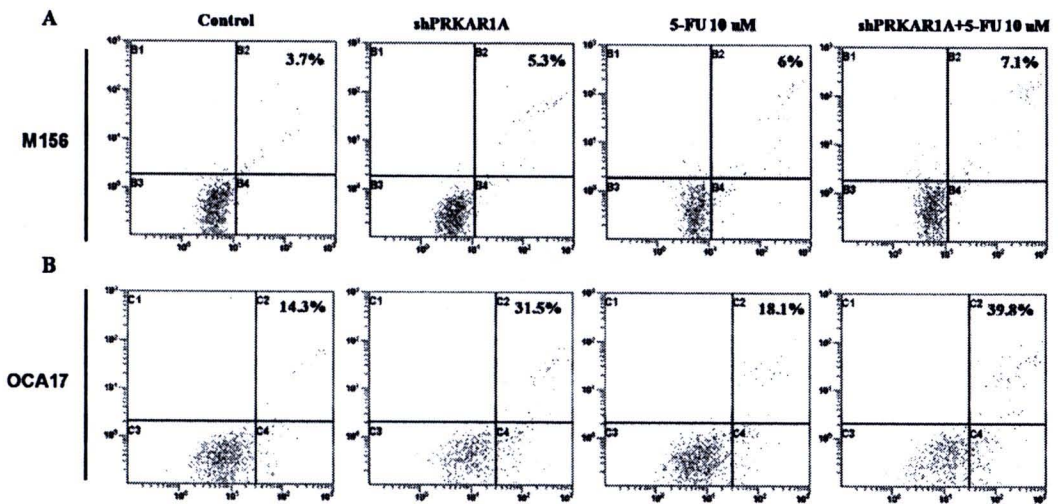


Figure B.8 Effect of shPRKAR1A in combination with 10 μ M 5-FU on induction of apoptosis in M156 (A) and OCA17(B). Annexin-V and PI (propidium iodide) staining was performed, followed by flow cytometry.

Table B.1 Effect of PRKAR1A silencing cells in combination with protein kinase inhibitors or chemotherapeutic drugs on CCA cells growth inhibition. This table represents growth inhibitory effect of either each agent alone or in combination and the expected percentage growth inhibition if drugs are additive when used in the combination.

		% Growth inhibition									
		M156					OCA17				
Drugs	Conc. (μM)	Control	shPRKAR1A	Drug	shPRKAR1A + drug	Expected additive effect	Control	shPRKAR1A	Drug	shPRKAR1A + drug	Expected additive effect
Sorafenib	0	0	29				0	42			
	0.01			8	32	37			5	47	47
	0.1			9	35	38			19	59	61
	1			11	37	40			27	75	69
	10			72	12	101			86	86	128
Sunitinib	0	0	31				0	25			
	0.01			3	33	34			0	30	25
	0.1			4	36	35			0	38	25
	1			8	45	39			7	36	32
	10			50	74	84			73	84	98

Table B.1 Effect of PRKAR1A silencing cells in combination with protein kinase inhibitors or chemotherapeutic drugs on CCA cells growth inhibition. This table represents growth inhibitory effect of either each agent alone or in combination and the expected percentage growth inhibition if drugs are additive when used in the combination (Cont.).

		% Growth inhibition											
		M156					OCA17						
Drugs	Conc. (µM)	Control	shPRKAR1A Drug	shPRKAR1A + drug	Expected additive effect	Control	shPRKAR1A Drug	shPRKAR1A + drug	Expected additive effect	Control	shPRKAR1A Drug	shPRKAR1A + drug	Expected additive effect
Gefitinib	0	0	30			0	23	9	37	32			
	0.01		0	39	30			18	49	41			
	0.1		0	41	30			24	50	47			
	1		1	41	31			35	57	58			
	10		23	54	53								
Met inhibitor	0	0	42			0	30						
	0.01		2	43	44			0	44	30			
	0.1		6	46	48			0	47	30			
	1		7	47	49			10	53	40			
	10		28	56	70			60	85	90			

Table B.1 Effect of PRKAR1A silencing cells in combination with protein kinase inhibitors or chemotherapeutic drugs on CCA cells growth inhibition. This table represents growth inhibitory effect of either each agent alone or in combination and the expected percentage growth inhibition if drugs are additive when used in the combination (Cont.).

		% Growth inhibition									
		M156					OCA17				
Drugs	Conc. (µM)	Control	shPRKAR1A	Drug	shPRKAR1A + drug	Expected additive effect	Control	shPRKAR1A	Drug	shPRKAR1A + drug	Expected additive effect
Paclitaxel	0	0	26				0	40			
	0.01			16	27	42			1	40	41
	0.1			23	32	49			1	46	41
	1			28	40	54			2	48	42
	10			31	48	57			14	53	54
Doxorubicin	0	0	33				0	30			
	0.01			5	32	38			0	30	30
	0.1			6	36	39			1	29	31
	1			31	67	64			35	30	65
	10			63	75	96			29	37	59


Table B.1 Effect of PRKAR1A silencing cells in combination with protein kinase inhibitors or chemotherapeutic drugs on CCA cells growth inhibition. This table represents growth inhibitory effect of either each agent alone or in combination and the expected percentage growth inhibition if drugs are additive when used in the combination (Cont.).

		% Growth inhibition									
		M156					OCA17				
Drugs	Conc. (µM)	Control	shPRKAR1A	Drug	shPRKAR1A + drug	Expected additive effect	Control	shPRKAR1A	Drug	shPRKAR1A + drug	Expected additive effect
5-FU	0	0	35				0	36			
	0.01			0	39	65			0	35	36
	0.1			0	42	65			3	46	39
	1			5	46	40			12	59	48
	10			34	67	69			43	72	79
Gemcitabine	0	0	7				0	6			
	0.01			0	9	7			0	6	6
	0.1			75	78	153			2	6	8
	1			82	81	88			31	37	37
	10			88	89	95			97	100	103

Table B.2 Effect of PRKAR1A silencing cells in combination with protein kinase inhibitors or chemotherapeutic drugs on induction of apoptosis. This table represents percentage apoptosis of either each agent alone or in combination and the expected percentage apoptosis if drugs are additive when used in the combination.

		% Apoptosis									
		M156					OCA17				
Drugs	Conc. (µM)	Control	shPRKAR1A	Drug	shPRKAR1A + Drug	Expected additive effect	Control	shPRKAR1A	Drug	shPRKAR1A + Drug	Expected additive effect
Sorafenib	1	0	1.81	2.19	3.25	4	0	10.62	9.93	13.66	20.55
	10		1.81	6.14	9.28	7.95		10.62	24.53	35.26	35.15
Sunitinib	1	0	2.06	10.91	11.44	12.97	0	31.305	7.48	36.29	38.79
	10		2.06	21.55	26.38	23.6		31.305	57.38	63.41	88.69
5-FU	10	0	1.6	2.36	3.43	3.96	0	17.14	3.8	25.49	20.94
	20		1.6	3.59	4.72	5.19		17.14	10.87	28.46	28.01

1. KKU-M156

Cell Name	KKU-M156	
Species	<i>Homo Sapiens</i>	
Sex	Male	
Age	68-year-old	
Diagnosis	Cholangiocarcinoma	
Histological type	Moderately differentiated	
Establisher	Sripa, B. et al.	
Medium	Ham's F-12 medium with 10% fetal bovine serum	
Passage Method	Cells treated with 0.25% Trypsin-EDTA for 5 min at 37°C	

2. KKU-OCA17

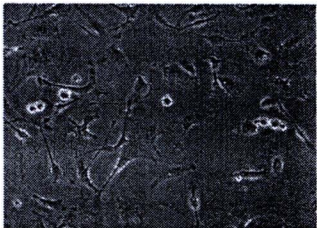
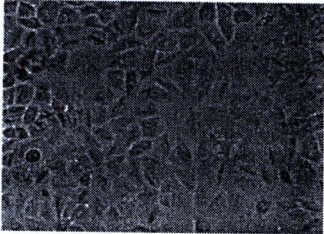
Cell Name	KKU-OCA17	
Species	<i>Homo Sapiens</i>	
Sex	Male	
Age	38-year-old	
Diagnosis	Cholangiocarcinoma	
Histological type	Well differentiated	
Establisher	Sripa, B. et al.	
Medium	Ham's F-12 medium with 10% fetal bovine serum	
Passage Method	Cells treated with 0.25% Trypsin-EDTA for 5 min at 37°C	

Figure B.9 Human CCA cell lines which were used in this study.

3. **KKU100**

Cell Name	KKU-100	
Species	<i>Homo Sapiens</i>	
Sex	Female	
Age	65-year-old	
Diagnosis	Cholangiocarcinoma	
Histological type	Poorly differentiated	
Doubling time	36 h	
Establisher	Sripa, B. et al.	
Medium	Ham's F-12 medium with 10% fetal bovine serum	
Passage Method	Cells treated with 0.25% Trypsin-EDTA for 5 min at 37°C	

4. **KKU-M214**

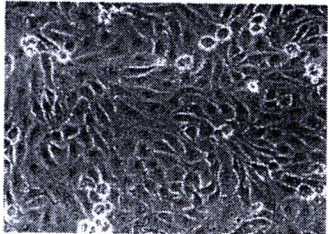
Cell Name	KKU-M214	
Species	<i>Homo Sapiens</i>	
Sex	Male	
Age	52-year-old	
Diagnosis	Cholangiocarcinoma	
Histological type	Well differentiated	
Doubling time	23 h	
Establisher	Sripa, B. et al.	
Medium	Ham's F-12 medium with 10% fetal bovine serum	
Passage Method	Cells treated with 0.25% Trypsin-EDTA for 5 min at 37°C	

Figure B.9 Human CCA cell lines which were used in this study. (Cont.).

APPENDIX C
Research publications & Presentations

1. RESEARCH PUBLICATIONS

1. Sirinun Juntana, Watcharin Loilome, Nisana Namwat, Raynoo Thanan, Apinya Jusakul, Janpen Puetkasichonpasutha, Sasithorn Yooyeun, Gregory J. Riggins, Puangrat Yongvanit. Antiproliferative effect induced by targeting of PRKAR1A/PKAI and chemotherapeutic drug on human cholangiocarcinoma cell lines. Proceeding. The 2nd Biochemistry and Molecular Biology (BMB) Conference, 140-143.
2. Watcharin Loilome, Nisana Namwat, Raynoo Thanan, Anchalee Techasen, Sirinun Juntana, Puangrat Yongvanit. Antiproliferative effect of PRKAR1A gene suppression by RNA interference in cholangiocarcinoma cell line., KKU Res J, 903-910.
3. Watcharin Loilome, Sirinun Juntana, Nisana Namwat, Anucha Puapairoj, Paiboon Sithithaworn, Masanao Miwa, Hideyuki Saya, Gregory J. Riggins and Puangrat Yongvanit. PRKAR1A is over- expressed and represents a possible therapeutic target in-human cholangiocarcinoma. Submitted to *Int J Cancer* (Under revision).

2. RESEARCH PRESENTATIONS

1. Sirinun Juntana *et al.* Antiproliferative effect induced by targeting of PRKAR1A/PKAI and chemotherapeutic drug on human cholangiocarcinoma cell line. The 2nd Biochemistry and Molecular Biology (BMB) Conference : Biochemistry and Molecular Biology for Regional Sustainable Development. Khon Kaen University, Khon Kaen Thailand 2009. (*Poster presentation*) May 7-8.
2. Sirinun Juntana *et al.* PRKAR1A acts as a possible therapeutic target in human cholangiocarcinoma. Annual Meeting of Faculty of Medicine, Khon Kaen University, Khon Kaen, Thailand 2009. (*Oral presentation*) October 15.

VITAE



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2003-2006 Bachelor Degree of Science (Medical technology, Second Class Honor) Faculty of Associated Medical Science, Khon Kaen University, Khon Kaen, Thailand

2007-2010 Graduate student of Master degree in Medical Biochemistry Program, Faculty of Medicine, Khon Kaen University, Khon Kaen, Thailand

Sep 2009-Jan 2010 Exchange research student, Department of Neurological surgery, School of Medicine, Johns Hopkins University, Baltimore, MD, USA

Scholarship:

2007-2008 Liver fluke and Cholangiocarcinoma Research Center Scholarship

Sep 2009- Jan 2010 University Mobility in Asia and the Pacific (UMAP) Scholarship

