

## บทที่ 7

### บรรณานุกรม

- Altmann, S. W., Davis, H. R., Zhu, L. J., Yao, X., Hoos, L. M., Tetzloff, G., Iyer, S. P., Maguire, M., Golovko, A., Zeng, M., Wang, L., Murgolo, N. and Graziano, M. P. (2004). Niemann-Pick C1 Like 1 protein is critical for intestinal cholesterol absorption. *Science*. 303: 1201-1204.
- Anderson, J. W., Johnstone, B. M. and Cook-Newell, M. E. (1995). Meta-analysis of the effects of soy protein intake on serum lipids. *N Engl J Med*. 333: 276-282.
- AOAC. 2000a. Methods 920.36 Dry matter on oven drying : Official methods of analysis. VOI I, Chapter 4. *Association of Official Analytical Chemists International* (17th ed). Gaithersburg, MD.
- AOAC. 2000b. Methods 984.13 Protein (crude) in animal feed and pet food: Official methods of analysis. VOI I, Chapter 4. *Association of Official Analytical Chemists International* (17th ed). Gaithersburg, MD.
- Badger, T. M., Ronis, M. J., Simmen, R. C. and Simmen, F. A. (2005). Soy protein isolate and protection against cancer. *J Am Coll Nutr* 24: 146S-149S.
- Belobrajdic, D. P., McIntosh, G. H. and Owens, J. A. (2003). Whey proteins protect more than red meat against azoxymethane induced ACF in Wistar rats. *Cancer Lett* 198: 43-51.
- Bostick, R. M., Potter, J. D., McKenzie, D. R., Sellers, T. A., Kushi, L. H., Steinmetz, K. A. and Folsom, A. R. (1993). Reduced Risk of Colon Cancer with High Intake of Vitamin E: The Iowa Women's Health Study. *Cancer Res* 53: 4230-4237.
- Bouzourene, H., Chaubert, P., Seelentag, W., Bosman, F. T. and Saraga E. (1999). Aberrant crypt foci in patients with neoplastic and nonneoplastic colonic disease. *Hum Pathol* 30: 66-71.
- Campbell, S., Stone, W., Whaley, S. and Krishnan, K. (2003). Development of gamma (gamma) - tocopherol as a colorectal cancer chemopreventive agent. *Crit Rev Oncol Hematol*. 47: 249-259.

- Cheng, L. and Lai, M. D. (2003). Aberrant crypt foci as microscopic precursors of colorectal cancer. *World J Gastroenterol* 9: 2642-2649.
- Clevers, H. (2006). Wnt/[beta]-Catenin Signaling in Development and Disease. *Cell* 127: 469-480.
- Compher, C. W., Frankel, W. L., Tazelaar, J., Lawson, J. A., McKinney, S., Segall, S., Kinosian, B. P., Williams, N. N. and Rombeau, J. L. (1999). Wheat Bran Decreases Aberrant Crypt Foci, Preserves Normal Proliferation, and Increases Intraluminal Butyrate Levels in Experimental Colon Cancer. *JPNEN J Parenter Enteral Nutr* 23: 269-278.
- DeMarini D. M. (1998). Dietary interventions of human carcinogenesis. *Mutat Res.* 400: 457-465.
- FAO. (2008). Modern techniques for feed analysis - I. MUELLER - HARVEY [On - line]. Available: <http://www.fao.org/docrep/007/y5159e/y5159e03.htm>.
- Ferguson, L. R. (2005). Does a diet rich in dietary fibre really reduce the risk of colon cancer. *Dig Liver Dis.* 37: 139-141.
- Ferris, R. A., Flores, R. A., Shanklin, C. W. and King, M. K. (1995). A proximate analysis of food service waste. *App Eng Agric.* 11: 567 - 572.
- Freddi, G., Mossotti, R. and Innocenti, R. (2003). Degumming of silk fabric with several proteases. *J Biotechnol.* 106: 101-112.
- Gamo, T., Inokuchi, T. and Laufer, H. (1977). Polypeptides of fibroin and sericin secreted from the different sections of the silk gland in *Bombyx mori*. *Insect Biochemistry*. 7: 285-295.
- Giovannucci, E., Rimm, E. B., Ascherio, A., Stampfer, M. J., Colditz, G. A. and Willett, W. C. (1995). Alcohol, Low-Methionine-Low-Folate Diets, and Risk of Colon Cancer in Men. *J. Natl. Cancer Inst.* 87: 265-273.
- Grosch, W. and Belitz, H. -D. (1987). *Food Chem.* Springer - Verlag Berlin: Germany.

Gross, M. D. (2005). Vitamin D and Calcium in the Prevention of Prostate and Colon Cancer: New Approaches for the Identification of Needs. *J. Nutr.* 135: 326-331.

Hakkak, R., Korourian, S., Ronis, M. J. J., Johnston, J. M. and Badger, T. M. (2001a). Dietary Whey Protein Protects against Azoxymethane-induced Colon Tumors in Male Rats. *Cancer Epidemiol Biomarkers Prev.* 10: 555-558.

Hakkak, R., Korourian, S., Ronis, M. J. J., Johnston, J. M. and Badger, T. M. (2001b). Soy protein isolate consumption protects against azoxymethane-induced colon tumors in male rats. *Cancer Lett.* 166: 27-32.

Harris, D. M. and Go, V. L. W. (2004). Vitamin D and Colon Carcinogenesis. *J. Nutr.* 13: 3463-3471.

Harris, P. J. and Ferguson, L. R. (1999). Dietary fibres may protect or enhance carcinogenesis. *Mutat Res.* 443: 95-110.

Hidalgo, I. J., Raub, T. J. and Borchardt, R. T. (1989). Characterization of the human colon carcinoma cell line (Caco-2) as a model system for intestinal epithelial permeability. *Gastroenterology.* 96: 736-749.

Ishida, H., Miwa, H., Tatsuta, M., Masutani, S., Imamura, H., Shimizu, J., Ezumi, K., Kato, H., Kawasaki, T., Furukawa, H. and Kawakami, H. (2004). Ki-67 and CEA expression as prognostic markers in Dukes' C colorectal cancer. *Cancer Lett.* 207: 109-115.

Institute for Laboratory Animal Research (ILAR). 1995. Nutritional requirements of laboratory animals, 4<sup>th</sup> ed [On-line]. Available: [http://books.nap.edu/openbook.php?record\\_id=4758&page=11.html](http://books.nap.edu/openbook.php?record_id=4758&page=11.html).

James, C.S. (1995). *Analytical chemistry of foods*. Blackie Academic & Professional: London.

Jemal, A., Murray, T., Ward, E., Samuels, A., Tiwari, R. C., Ghafoor, A., Feuer, E. J., Thun, M. J. (2005). Cancer statistics. *CA Cancer J Clin.* 55: 10 - 30.

Jemal, A., Siegel, R., Ward, E., Hao, Y., Xu, J., Murray, T. and Thun, M. J. (2008). Cancer statistics, 2008. *CA Cancer J Clin.* 58: 71-96.

- Janne, P. A. and Mayer, R. J. (2000). Chemoprevention of Colorectal Cancer. *N Engl J Med.* 342, 1960-1968.
- Johnson, I. T. (2004). New approaches to the role of diet in the prevention of cancers of the alimentary tract. *Mutat Res.* 551: 9-28.
- Kato, N., Sato, S., Yamanaka, A., Yamada, H., Fuwa, N. and Nomura, M. (1998). Silk protein, sericin, inhibits lipid peroxidation and tyrosinase activity. *Biosci Biotechnol Biochem.* 62: 145-147.
- Kayashita, J., Shimaoka, I., Nakajoh, M., Yamazaki, M. and Kato, N. (1997). Consumption of buckwheat protein lowers plasma cholesterol and raises fecal neutral sterols in cholesterol-Fed rats because of its low digestibility. *J Nutr.* 127: 1395-1400.
- Khuhaprema, T. and Srivatanakul, P. (2008). Colon and Rectum Cancer in Thailand: An Overview. *Jpn. J. Clin. Oncol.* 38: 237-243.
- Kundu, S. C., Dash, B. C., Dash, R. and Kaplan, D. L. (2008). Natural protective glue protein, sericin bioengineered by silkworms: Potential for biomedical and biotechnological applications. *Progress in Polymer Science.* 33: 998-1012.
- Kurioka, A., Kurioka, F. and Yamazaki, M. (2004). Characterization of Sericin Powder Prepared from Citric Acid-degraded Sericin Polypeptides of the Silkworm, *Bombyx Mori*. *Biotechnol Biochem.* 68: 774-780.
- Kwon, D. Y., Oh, S. W., Lee, J. S., Yang, H. J., Lee, S. H. and Lee, J. H. (2002). Amino acid substitution of hypocholesterolemic peptide originated from glycinin hydrolyzate. *Food Sci Biotech.* 11:55-61.
- Labianca, R., Beretta, G., Gatta, G., de Braud, F. and Wils, J. (2004). Colon cancer. *Crit Rev Oncol Hematol.* 51: 145-170.
- Liu, Z., Ishikawa, W., Huang, X., Tomotake, H., Kayashita, J., Watanabe, H. and Kato, N. (2001). A Buckwheat Protein Product Suppresses 1,2-Dimethylhydrazine-Induced Colon Carcinogenesis in Rats by Reducing Cell Proliferation. *J. Nutr.* 131: 1850-1853.

- Liyانage, R., Han, K.-H., Watanabe, S., Shimada, K.-i., Sekikawa, M., Ohba, K., Tokuji, Y., Ohnishi, M., Shibayama, S., Nakamori, T. and Fukushima, M. (2008). Potato and Soy Peptide Diets Modulate Lipid Metabolism in Rats. *Biosci Biotechnol Biochem.* 72: 943-950.
- Martinez, M. E. (2005). Primary prevention of colorectal cancer: lifestyle, nutrition, exercise. *Recent Results Cancer Res.* 166: 177 - 211.
- Megías, C., Pedroche, J., del Mar Yust, M., Alaiz, M., Girón-Calle, J., Millán, F. and Vioque, J. (2009). Sunflower Protein Hydrolysates Reduce Cholesterol Micellar Solubility. *Plant Foods Hum Nutr.* 64: 86-93.
- Morita, T., Oh-hashi, A., Takei, K., Ikai, M., Kasaoka, S. and Kiriyma, S. (1997). Cholesterol-Lowering Effects of Soybean, Potato and Rice Proteins Depend On Their Low Methionine Contents In Rats Fed a Cholesterol-Free Purified Diet. *J. Nutr.* 127: 470-477.
- Nagaoka, S., Miwa, K., Eto, M., Kuzuya, Y., Hori, G. and Yamamoto, K. (1999). Soy Protein Peptic Hydrolysate with Bound Phospholipids Decreases Micellar Solubility and Cholesterol Absorption in Rats and Caco-2 Cells. *J. Nutr.* 129: 1725-1730.
- Nagaoka, S., Futamura, Y., Miwa, K., Awano, T., Yamauchi, K., Kanamaru, Y., Tadashi, K. and Kuwata, T. (2001). Identification of Novel Hypocholesterolemic Peptides Derived from Bovine Milk [beta]-Lactoglobulin. *Biochem Biophys Res Commun.* 281: 11-17.
- National Laboratory Animal Centre, Mahidol University. 2008. Environmental monitoring of NLAC- MU [On-line]. Available: [http://www.nlac.mahidol.ac.th/nalcwwwtha/pa\\_environment.html](http://www.nlac.mahidol.ac.th/nalcwwwtha/pa_environment.html).
- Organization of Economic Co - operation and development. 2008. Manual for investigation of HPV chemicals: Annex 1: OECD Test guidelines for studies included in the SIDS [On-line]. Available: [http://www.oecd.org/document/23/0.2340.en\\_2649\\_34379\\_1948503\\_1\\_1\\_1\\_1.00.html](http://www.oecd.org/document/23/0.2340.en_2649_34379_1948503_1_1_1_1.00.html).
- Ponz de Leon, M. (2002). Prevention and chemoprevention of colorectal neoplasms. *Dig Liver Dis.* 34: 59-69.



- Potter, J. D., Slattery, M. L., Bostick, R. M. and Gapstur, S. M. (2004). Colon Cancer: A Review of the Epidemiology. *Epidemiol Rev.* 5: 499 - 545.
- Rogers, A. E., Zeisel, S. H. and Groopman, J. (1993). Diet and carcinogenesis. *Carcinogenesis.* 14: 2205-2217.
- Roynette, C. E., Calder, P. C., Dupertuis, Y. M. and Pichard, C. (2004). n-3 Polyunsaturated fatty acids and colon cancer prevention. *Clin Nutr.* 23: 139-151.
- Sasaki, M., Yamada, H. and Kato, N. (2000). A Resistant Protein, Sericin Improves Atropine-Induced Constipation in Rats. *Food Science and Technology Research.* 6: 280-283.
- Sasaki, M., Yamada, H. and Kato, N. (2000). Consumption of silk protein. sericin elevates intestinal absorption of zinc, iron, magnesium and calcium in rats. *Nutr Res.* 20: 1505-1511.
- Scholzen, T. and Gerdes, J. (2000). The Ki-67 protein: from the known and the unknown. *J Cell Physiol.* 182: 311-322.
- Sugano, M., Goto, S., Yamada, Y., Yoshida, K., Hashimoto, Y., Matsuo, T. and Kimoto, M. (1990). Cholesterol-Lowering Activity of Various Undigested Fractions of Soybean Protein in Rats. *J. Nutr.* 120: 977-985.
- Takahashi, M., Tsujimoto, K., Yamada, H., Takagi, H. and Nakamori, S. (2003). The silk protein, sericin, protects against cell death caused by acute serum deprivation in insect cell culture. *Biotechnol Lett.* 25: 1805-1809.
- Takahashi, T., Satou, M., Watanabe, N., Sakaitani, Y., Takagi, A., Uchida, K., Ikeda, M., Moriyama, R., Matsumoto, K. and Morotomi, M. (1999). Inhibitory effect of microfibril wheat bran on azoxymethane-induced colon carcinogenesis in CF1 mice. *Cancer Lett.* 141: 139-146.
- Takasu, Y., Yamada, H. and Tsubouchi, K. (2002). Isolation of three main sericin components from the cocoon of the silkworm, *Bombyx mori*. *Biosci Biotechnol Biochem.* 66: 2715-2718.

- Tamada, Y., Sano, M., Niwa, K., Imai, T. and Yoshino, G. (2004). Sulfation of silk sericin and anticoagulant activity of sulfated sericin. *J Biomater Sci Polym Ed.* 15: 971-980.
- Tanaka, T., Yasui, Y., Tanaka, M., Tanaka, T., Oyama, T. and Rahman, K. M. W. (2009). Melatonin suppresses AOM/DSS-induced large bowel oncogenesis in rats. *Chem Biol Interact.* 177: 128-136.
- Tatsuta, M., Iishi, H., Baba, M., Uedo, N., Ishihara, R., Higashino, K., Mukai, M. and Ishiguro, S. (2005). Induction by lysophosphatidic acid of peritoneal and pleural metastases of intestinal cancers induced by azoxymethane in Wistar rats. *Cancer Lett.* 219: 137-145.
- Tokutake, S. (1980). Isolation of the smallest component of silk protein. *J Biochem.* 187: 413-417.
- Tsujimoto, K., Takagi, H., Takahashi, M., Yamada, H. and Nakamori, S. (2001). Cryoprotective Effect of the Serine-Rich Repetitive Sequence in Silk Protein Sericin. *J Biochem.* 129: 979-986.
- van Breda, S. G., de Kok, T. M. and van Delft, J. H. (2007). Mechanisms of colorectal and lung cancer prevention by vegetables: a genomic approach. *J Nutr Biochem.* 19: 139-157.
- van de Wetering, M., Sancho, E., Verweij, C., de Lau, W., Oving, I., Hurlstone, A., van der Horn, K., Batlle, E., Coudreuse, D., Haramis, A. P., Tjon-Pon-Fong, M., Moerer, P., van den Born, M., Soete, G., Pals, S., Eilers, M., Medema, R. and Clevers, H. (2002). The beta-catenin/TCF-4 complex imposes a crypt progenitor phenotype on colorectal cancer cells. *Cell.* 111: 241-250.
- Walstra, P. (2003). *Physical chemistry of foods*. Marcel Dekker. New York.
- Weinglass, A. B., Kohler, M., Schulte, U., Liu, J., Nketiah, E. O., Thomas, A., Schmalhofer, W., Williams, B., Bildl, W., McMasters, D. R., Dai, K., Beers, L., McCann, M. E., Kaczorowski, G. J. and Garcia, M. L. (2008). Extracellular loop C of NPC1L1 is important for binding to ezetimibe. *Proc Natl Acad Sci USA.* 105: 11140-11145.
- Zhang, X. and Beynen, A. C. (1995). Influence of dietary fish proteins on plasma and liver cholesterol concentrations in rats. *Br J Nutr.* 69: 767-777.

- Zhang, Y. Q. 2002. Applications of natural silk protein sericin in biomaterials. *Biotechnol. Adv.* 20: 91-100.
- Zhaorigetu, S., Sasaki, M. and Kato, N. (2007). Consumption of sericin suppresses colon oxidative stress and aberrant crypt foci in 1,2-dimethylhydrazine-treated rats by colon undigested sericin. *J Nutr Sci Vitaminol (Tokyo)*. 53: 297-300.
- Zhaorigetu, S., Sasaki, M., Watanabe, H. and Kato, N. (2001). Supplemental silk protein, sericin, suppresses colon tumorigenesis in 1,2-dimethylhydrazine-treated mice by reducing oxidative stress and cell proliferation. *Biosci Biotechnol Biochem*. 65: 2181-2186.

## บทที่ 8

### ต้นฉบับงานวิจัยเพื่อตีพิมพ์ในวารสารต่างประเทศ

#### **1. การทดสอบฤทธิ์ของซิริซินในการป้องกันมะเร็งลำไส้ใหญ่และการหนอก**

**Sericin Consumption Suppresses Development and Progression of Colon Tumorigenesis in  
1,2-Dimethylhydrazine-Treated Rats**

Waraporn Kaewkon<sup>a</sup>, Waree Tiyaboonchai<sup>b</sup>, Sutatip Pongcharoen<sup>c</sup> Manote Sutheerawattananonda<sup>d</sup> and  
Nanteetip Limpeanchob<sup>a</sup>

<sup>a</sup>Departments of Pharmacy Practice, <sup>b</sup> Pharmaceutical Technology, Faculty of Pharmaceutical Sciences, <sup>c</sup>Department of Medicine, Faculty of Medicine, Naresuan University, Phitsanulok 65000 and <sup>d</sup>School of Food Technology, Institute of agricultural Technology, Suranaree University of Technology, Nakhon Ratchasima 30000, Thailand.

Corresponding author: Nanteetip Limpeanchob

Faculty of Pharmaceutical Sciences, Naresuan University,

Phitsanulok 65000, Thailand

Tel.: +6655261000 ext 1822

Fax: +6655261923

E-mail: Nanteetipl@nu.ac.th

### Abstract

Colon cancer is one of the most common cancers in many countries. Dietary factor is a major risk for developing this type of cancer, thus dietary interventions could perhaps prevent the diseases. This study was conducted to examine the chemopreventive effect of sericin against 1,2-dimethylhydrazine (DMH)-induced colon tumorigenesis in rats. Rats were fed with 4% sericin or 4% casein diet for 20 weeks, and given 20 mg/kg DMH subcutaneously injection once a week for the initial ten weeks. Supplementation of sericin did not affect the incidence rate of aberrant crypt foci (ACF) formation, but effectively suppressed tumor development compared to casein supplement. Consumption of sericin prior to carcinogen exposure appeared to reduce the number and multiplicity of ACF indicating its ability to suppress the initiation and promotion stages of tumorigenesis. Sericin supplement after being exposed to DMH could also reduce the development of ACF, while crypt multiplicity was not affected. Although, it's not statistically significance, the level of colonic lipid peroxidation in rats fed sericin diet showed a tendency to be lower than that of casein diet. From Ki67 immunohistochemistry and colonic morphological analysis, sericin seemed to delay tumorigenesis process since colonic crypts of sericin fed rats were mostly in the hyperplastic stage whereas those of casein fed rats showed abnormal and dysplastic morphology indicating more advanced stage of tumorigenesis. This finding suggests that consumption of sericin could reduce the risk of developing colon tumor by suppressing the initiation and progression of tumorigenesis.

**Key words:** colon cancer, sericin, 1,2-dimethylhydrazine, aberrant crypt foci

## 1. Introduction

Colon cancer is one of the most common cancers affecting people worldwide. The incidence is high in high income countries, but it is now increasing in middle- and low income countries (Labianca et al. 2010). The incidence rate of colon cancer in Thailand is low when compared with other countries and the highest incidence is seen in Bangkok (Khuhaprema and Srivatanakul 2008). This rate is expected to be rapidly increased in the next decade probably due to the acquisition of Western lifestyle. Western style diet (high levels of fat and red meat, and low dietary fiber) is the major risk factor of colorectal cancer (Rogers et al. 1993; van Breda et al. 2007). Since diet is definitely important for colon cancer development, dietary interventions are received much attention as one of the approaches to prevent this type of cancer (Rogers et al. 1993; DeMarini 1998; Johnson 2004).

Several experimental studies have proposed that the protective effects of diets rich in fruits and vegetables against colon carcinogenesis are thought to be due to their content of anti-oxidant vitamins and fibers (Wogan 1985; Levi et al. 2001; Campos et al. 2005). Many types of dietary proteins such as wheat bran, soy bean, buckwheat, and whey proteins were also found to reduce the development of colon cancer in animals (Compher et al. 1999; Takahashi et al. 1999; Hakkak et al. 2001a; Liu et al. 2001; McIntosh and Le Leu 2001; Belobrajdic et al. 2003; Badger et al. 2005). Most studies have been focused on proteins isolated from regular diets. The present study was aimed to investigate sericin, an eatable protein isolated from silk cocoon.

Sericin is a glue protein that secreted from the middle part of silk gland of silkworm, *Bombyx mori* (Gamo et al. 1977; Grzelak 1995; Kundu et al. 2008). Its molecular weight is ranging from 20 to 400 kDa with high content of serine, glycine and aspartic acid (Tokutake 1980; Zhang 2002). Many pharmacological activities of sericin including anti-oxidation (Kato et al. 1998; Dash et al. 2008), tyrosinase inhibition (Kato et al. 1998), anti-coagulation (Tamada et al. 2004), chemoprevention (Zhaorigetu et al. 2001; Zhaorigetu et al. 2003a; Zhaorigetu et al. 2003b; Zhaorigetu et al. 2007), and liver

and gastric protection (Li et al. 2008) were reported. Regarding to its chemopreventive effect, sericin consumption significantly suppress colon tumorigenesis in 1,2-dimethylhydrazine (DMH)-treated animals (Zhaorigetu et al. 2001; Zhaorigetu et al. 2007). The mechanisms of its chemoprevention are associated with the ability to reduce colonic oxidative stress and cell proliferation (Zhaorigetu et al. 2001; Zhaorigetu et al. 2007). It should be noted that protein preparation of sericin is generally heterogeneous consisting of various sizes of polypeptides depending on separation techniques. Due to sericin's heterogeneity caused by the degumming procedure, different sericin preparations may differently display their pharmacological activities. Small sericin (5-100 kDa) was found to promote mammalian cell proliferation better than large sericin (50-200 kDa) (Terada et al. 2005). Thus, the present study was aimed to test the chemopreventive effect against colon tumorigenesis of sericin isolated by xxxx techniques developing by Institution of Agricultural Technology, Suranaree University of Technology, Thailand. The molecular weight of sericin used in this study was 76-132 kDa.

Ki67, a cellular marker of cell proliferation, was also investigated to determine the potential mechanism of action of sericin.

## 2. Materials and methods

### *Animals and Chemicals*

Male Sprague-Dawley rats (120-160 g) were purchased from the National Laboratory Animal Centre, Mahidol University, Thailand. All experimental procedures were approved by the Animal Research Ethics Committee, Naresuan University, Thailand. The animals were housed in stainless steel cages at temperature of  $24 \pm 2$  °C with a 12 hours light-dark cycle. All rats were fed the assigned experimental diet and water ad libitum. DMH, methylene blue and EDTA were purchased from Sigma Chemical Company (St. Louis, MO, USA). All other chemicals and reagents used were of analytical grade. Sericin was isolated by Institution of Agricultural Technology, Suanaree University of Technology, Nakhon Ratchasima, Thailand.

### *Experimental protocol*

After 1 week acclimatization period, the rats were divided into 5 groups ( $n = 6/\text{group}$ ). The experimental diets were prepared by mixing commercial diet with casein or sericin at the level of 4%. The experimental design is shown in Fig. 1. Group 1 and 3 were fed casein diet whereas group 2 and 4 were fed sericin diet. The last group (post-sericin) was received casein diet for the first 5 weeks and then fed sericin diet until the end of experiment. Group 3, 4 and 5 were subcutaneously injected with DMH (20 mg/kg bodyweight) once a week for 10 weeks. Group 1 and 2 were control groups injected with 1 mM EDTA pH 6.5 used to dissolve DMH for the same period of time. Food consumption and body weight were recorded weekly throughout the experimental period. Ten weeks after the last DMH injection, all rats were sacrificed and colons were removed and flushed with cold phosphate buffer saline (pH 7.4). The colon was divided into three equal parts, proximal, middle and distal. Each part was then cut into three segments for evaluation of aberrant crypt foci (ACF), lipid peroxidation and Ki67 immunohistochemistry.

### **Determination of aberrant crypt foci (ACF)**

The colons were flushed with cold PBS, opened along the longitudinal median axis and fixed flat between two pieces of filter paper in 10% neutral formalin buffer for 6 hours. The colonic tissues were stained with 0.2% methylene blue in PBS for 5 minutes. The colons were placed on microscopic slides and observed ACF formation under a microscope. (Bird and Good 2000). Aberrant crypts were distinguished from the surrounding normal crypts by their increased size and had thicker epithelial lining compare to normal crypts. They usually gathered into a focus, consisting of one or more aberrant crypts (AC). (Bird 1987; McLellan and Bird 1988) The number of ACF in each part of colon was recorded. Crypt multiplicity was analyzed by determining the number of crypts in each focus and was categorized as 1, 2, 3, 4 and 5 or more aberrant crypts per focus or AC/ACF.

### **Lipid peroxidation of colon tissue**

Lipid peroxidation was estimated by measuring thiobarbituric acid reactive substances (TBARs) assay. Briefly, colon was homogenized with ice-cold PBS pH 7.4 containing 1 mM phenylmethanesulfonylfluoride (PMSF), incubated with TBARs reagent (40% TCA: 1.4% TBA: 8% HCl) and heated at 90°C for 1 hour. After, centrifugation, the fluorescent intensity was measured (Ex 535 nm, Em 595 nm). The protein content of colon homogenate was determined using BCA protein assay kit (Pierce).

### **Immunohistochemical analysis of Ki67**

After fixing colon segments in 10% neutral formalin buffer for 48 hours, they were cut into serial strips, processed for paraffin-embedding and cut into 3 µm-thick sections. The expression of Ki67 was detected by immunohistochemistry using avidin-biotin complex method (Vectastain Elite ABC kit, Vector Laboratories, USA). The procedure was performed as the manufacturer recommendation. Colon sections were deparaffinized with xylene, dehydrated through a graded ethanol series. Hydrogen peroxide at 0.5% was administered to the slides for 10 minutes. Antigen retrieval was accomplished by

heating the sections in 10 mM citrate buffer pH 6.0 for 10 minutes. The slides were then incubated with the rabbit monoclonal antibody Ki67 (Abcam, Cambridge, UK) for 1 hour. Biotinylated secondary antibody was applied for 30 minutes, followed by incubation with DAB peroxidase substrate, counterstained with hematoxylin.

The numbers of Ki67 positive cells in the individual crypt were recorded. In each group, the average number of Ki67 positive cells per crypt was calculated from the total 60 crypts (10 crypts /rat).

#### *Statistical analysis*

All data are expressed as means  $\pm$  standard deviation (SD). The data were analyzed by one-way analysis of variance (ANOVA) and student t-test. Differences were considered to be significance when p value  $\leq 0.05$ .

### 3. Results

#### *Body weight and food consumption*

Initial and final body weights of all groups were not significantly different (Table 2). The initial diet intake of group 4 was unexpectedly lower than the others, probably because rats in this group were slightly smaller than the rest. Food consumption at the end of the experiment significantly increased in DMH-treated groups (group 3-5) compared to control groups (group 1 and 2). Type of protein did not affect body weight and food consumption.

#### *Incidences of ACF and tumor development*

Table 3 summarizes the incidence of ACF which is considered as the early sign of colon tumorigenesis and tumor formation. There was no ACF and tumors found in the colon of control rat in group 1 and 2. All DMH-treated rats showed the incidence of ACF development. Two rats in casein fed group appeared to have tumors in their colons, whereas none of rats in sericin diet groups showed sign of tumor. This observation suggests that consumption of sericin containing diet did not completely block the DMH-induced ACF formation, but effectively suppressed tumor development. The density of ACF in casein diet (group 3) was higher than that of sericin diet (group 4 and 5), although it was not statistically different. It should be noted that two of six rats in casein diet group developed tumors in which the number of ACF reported in the result was likely to be underestimated due to the interference from tumorigenesis. Thus, sericin diet appears to suppress or decelerate colon tumorigenesis process whether or not it is intake before or after the exposure of carcinogen.

#### *ACF distribution and multiplicity*

ACF found in DMH-treated rats were mostly localized at the distal part of colon (Table 4). The density of ACF in the middle and distal colon of sericin fed rats was lower than that of casein fed rats, although it was not statistically significant. Distribution of ACF in the proximal colon was observed only

in casein diet group. This observation suggests that sericin diet could retard the expanding of ACF formation referring to its ability to delay the progression of colon cancer pathogenesis.

Crypt multiplicity was also analyzed and represent as number of aberrant crypt (AC) per ACF (Table 4). Multiplicity of ACF was analyzed by dividing into five categories depending on the number of AC/ACF (1, 2, 3, 4 and  $\geq 5$  AC/ACF). Distal colon showed the highest multiplicity, approximately 3 AC/ACF, indicating larger size of ACF compared to ACF in other parts of colon. The multiplicity of the average number of AC/ACF in the distal part of colon was not affected by type of protein supplement. The degree of multiplicity of ACF in distal colon was further analyzed since most ACF were distributed in this segment. The density of ACF containing  $\leq 4$  AC was not different among three groups (table 4). In both sericin fed groups, the densities of large ACF containing  $\geq 5$  AC ( $28.79 \pm 15.79$  and  $21.43 \pm 9.03$ ) were lower than that of casein diet group ( $37.66 \pm 10.12$ ). Because of high variation in the actual size among large ACF ( $\geq 5$  AC/ACF), area analysis of individual ACF was performed. Average size of large ACF in casein diet was  $0.14 \pm 0.06$  mm<sup>2</sup> and it was smaller ( $0.11 \pm 0.05$  mm<sup>2</sup>) in rats fed sericin except, rats in post-sericin group. Taken all data together, sericin consumption prior to carcinogen exposure appears to reduce ACF density and crypt multiplicity suggesting the ability to suppress both ACF development and progression. Sericin could only suppress ACF formation when it was consumed after carcinogen administration.

#### ***Lipid peroxidation of distal colon***

Lipid peroxidation of the colon was measured by using TBARs assay as showed in table 5. The production of malondialdehyde (MDA) was referred to the intensity of lipid peroxidation reaction. Since sericin diet exhibited beneficial effect in the distal part of rat colon, lipid peroxidation in this segment was firstly determined. Although the lipid peroxidation products in distal colon of sericin diet groups seemed to be lower than those of casein diets (group 1 vs 2 and group 3 vs 4, 5), no significant difference was observed between the two protein types because of high variation among individual rat.



***Immunohistochemical and morphological analysis***

The marker of cell proliferation, Ki67 was determined in this study. Our finding showed no difference in the average number of Ki67 positive cells presenting in each colonic crypt among groups 1-3 (table 6). Unexpectedly the number of Ki67 positive cells in group 4 and 5 were higher than the first three groups. Compared to DMH-treated rat fed casein diet (group 3), the length of colonic crypt in DMH-treated rat fed sericin diet (group 4 and 5) were higher. In addition, Ki67 staining cells of the last two groups were mostly distributed in the top zone of the crypt. Taken all data together, sericin diet seemed to promote colonic cell proliferation. However, we observed dramatically changes in morphology of colonic crypt of group 3 rats such as abnormal mucosal lining and dysplastic crypts (figure 1C, D). In sericin fed groups, the appearance of colonic crypts was similar to normal but longer in length and hyperplastic morphology was observed (figure 1E, F).

#### 4. Discussions

The incidence of colon cancer is rapidly rising in many areas where Western lifestyle especially type of diets is being immersed into their eating habits. Thus, dietary interventions seem to have a great influence on preventing this kind of cancer. Many animal studies indicated that supplement with dietary proteins including wheat bran, soy bean, buckwheat, and whey proteins could protect against the development of colon cancer (Compher et al. 1999; Takahashi et al. 1999; Hakkak et al. 2001b, 2001a; Liu et al. 2001; McIntosh and Le Leu 2001; Belobrajdic et al. 2003; Badger et al. 2005). Sericin is one of eatable protein that has received of much attention since it was demonstrated to be beneficial for human health in many aspects. The present study was therefore interested to evaluate the chemopreventive effect of sericin against DMH-induced colon cancer in rats.

Sericin is a dietary fiber-like protein with very low digestibility (Sasaki et al. 2000) . Undigested sericin was found to play a role in the mechanism of its chemopreventive effect against colon tumorigenesis (Zhaorigetu et al. 2007). Sericin is a complex mixture of polypeptides differing in size, chemical and structural properties (Gamo et al. 1977). Degumming processes generally affect certain properties of sericin preparations (Freddi et al. 2003; Kurioka et al. 2004), and subsequently could showed to be affect their pharmacological activities, for example 5-100 kDa sericin was superior to 50-200 kDa sericin in accelerating cell proliferation in culture (Terada et al. 2005). Therefore, this study was conducted to test sericin (MW 76-132 kDa) prepared by degumming process developed by Suranaree University of Technology. Our result indicate that sericin diet showed could suppress the development and delay progression of tumorigenesis whether or not sericin is consumed before or after the exposure of DMH carcinogen.

DMH, an alkylating agent, is metabolized in the liver by conjugating with glucuronic acid. This glucuronide is excreted in the bile and then enters to the gut where microbial beta-glucuronidase releases the active metabolite to mediate its carcinogenesis activities on the mucosa (Reddy et al. 1974;

Fiala 1975; Emerich 1977; Chipman 1982). Thus, DMH seems to be highly specific to initiate colon tumorigenesis. In this study, DMH administration effectively induced the development of ACF with 100% incidence rate. Food consumption in DMH-treated rats was higher than control rats, possible due to the reduction in nutrient absorbability. Consistent with previous studies (Zhaorigetu et al. 2001; Zhaorigetu et al. 2007), this study indicated that sericin diet suppressed the development of ACF, an early marker of colon tumorigenesis. Sericin did not provide statistically significant reduction in the density of ACF in the present study, probably because of under estimation of the number of ACF in casein diet group due to tumor development. There was no sign of tumor developed after sericin supplement indicating less tumorigenesis progression than casein supplement.

According to previous reports, the increased number of ACF seemed to reflect the initiation step of colon carcinogenesis, whereas the increase in number of aberrant crypts (AC) per ACF or crypt multiplicity might correspond to the progression or promotion step of the disease (Zhang et al. 1992; Magnuson et al. 1993; Fenoglio-Preiser and Noffsinger 1999) Consumption of sericin prior to the exposure of carcinogen appeared to suppress both initiation and promotion stages. Even though sericin diet was consumed after receiving carcinogen, it could still reduce the number of ACF formation. Post-sericin might somehow affect the progression of colon tumorigenesis despite crypt multiplicity is not obviously reduced but tumor development was not occurred in this group of rat.

Consumption of sericin could reduce cell proliferation possibly by suppressing *c-myc* and *c-fos* proliferation-related genes (Zhaorigetu et al. 2001). Sericin diet also suppressed oxidative stress in the colon mucosa that might at least in part lead to the reduction of cell proliferation (Zhaorigetu et al. 2001). The reduction of colon oxidative stress by sericin supplement in the present study was not clearly demonstrated as the previous report. The Ki67, a nuclear protein associated with cell proliferation (Pollack et al. 2004), was determined in this study. In normal colonic mucosa, the proliferating cells is

localized to the basal and lower one third of the crypt, then the cells migrate from the base of the crypts upwards towards the luminal surface, where they are sloughed off (Ishida et al. 2004).

Ki67 is a biomarker of proliferation presenting in all phases of cell cycle except for G0 phase. Ki67 positive cells were stained along whole crypts up to the mucosal surface. Unexpectedly, sericin diet in DMH-treated rats appeared to increase the number of Ki67 positive cells and the length of colonic crypts indicating its cell proliferation promoting properties. Colonic crypts from these rats were also appeared in hyperplastic morphology. In tumorigenesis process, ACF show variable histological features, ranging from hyperplasia to dysplasia (Cheng and Lai 2003). Colonic crypts of casein diet DMH-treated rats were histologically heterogeneous, hyperplastic and dysplastic crypts were observed together with abnormal mucosal lining indicating more advanced stage of tumorigenesis than sericin diet groups. Therefore, sericin consumption might not directly promote colonic cell proliferation but instead delay the progression of colon tumorigenesis.

High content of arginine and glycine in buckwheat protein was suggested to be responsible for its chemopreventive effect against colon tumorigenesis (Liu et al. 2001). Supplementation of glycine was also found to inhibit the growth of B16 melanoma tumors in mice (Rose et al. 1999). Sericin protein contains higher concentration of glycine (13.5% w/w) compared to casein (1.6% w/w) (Morita et al. 1997). It is possible that glycine component is involved in the protective effect of sericin against colon tumorigenesis but further study is necessary before coming to this conclusion.

In conclusion, the chemopreventive effect of sericin consumption on ACF development occurred in both initiation and promotion stages. Sericin diet could suppress or decelerate colon tumorigenesis process whether or not it is intake before or after the exposure of carcinogen. Thus, sericin is one of the interesting food supplements exhibiting various beneficial effects for human health, particularly its preventive effect to colon tumorigenesis.

## References

- Badger T. M., Ronis M. J., Simmen R. C. and Simmen F. A. (2005) Soy protein isolate and protection against cancer. *J Am Coll Nutr* **24**, 146S-149S.
- Belobrajdic D. P., McIntosh G. H. and Owens J. A. (2003) Whey proteins protect more than red meat against azoxymethane induced ACF in Wistar rats. *Cancer Letters* **198**, 43-51.
- Bird R. P. (1987) Observation and quantification of aberrant crypts in the murine colon treated with a colon carcinogen: Preliminary findings. *Cancer Letter* **37**, 147 - 151.
- Bird R. P. and Good C. K. (2000) The significance of aberrant crypt foci in understanding the pathogenesis of colon cancer. *Toxicology Letters* **112-113**, 395-402.
- Campos F. G., Logullo Waitzberg A. G., Kiss D. R., Waitzberg D. L., Habr-Gama A. and Gama-Rodrigues J. (2005) Diet and colorectal cancer: current evidence for etiology and prevention. *Nutr Hosp* **20**, 18-25.
- Chipman J. K. (1982) Bile as a source of potential reactive metabolites. *Toxicology* **25**, 99-111.
- Compher C. W., Frankel W. L., Tazelaar J., Lawson J. A., McKinney S., Segall S., Kinosian B. P., Williams N. N. and Rombeau J. L. (1999) Wheat Bran Decreases Aberrant Crypt Foci, Preserves Normal Proliferation, and Increases Intraluminal Butyrate Levels in Experimental Colon Cancer. *JPNEN J Parenter Enteral Nutr* **23**, 269-278.
- Dash R., Acharya C., Bindu P. C. and Kundu S. C. (2008) Antioxidant potential of silk protein sericin against hydrogen peroxide-induced oxidative stress in skin fibroblasts. *BMB Rep* **41**, 236-241.
- DeMarini D. M. (1998) Dietary interventions of human carcinogenesis. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis* **400**, 457-465.
- Emerich S. F. (1977) Investigations into the metabolism and mode of action of the colon carcinogens 1,2-dimethylhydrazine and azoxymethane. *Cancer* **40**, 2436-2445.
- Fenoglio-Preiser C. M. and Noffsinger A. (1999) Aberrant Crypt Foci: a Review. *Toxicol Pathol* **27**, 632 - 642.
- Fiala E. (1975) Investigations into the metabolism and mode of action of the colon carcinogen 1,2-dimethylhydrazine. *Cancer* **36**, 2407-2412.
- Freddi G., Mossotti R. and Innocenti R. (2003) Degumming of silk fabric with several proteases. *Journal of Biotechnology* **106**, 101-112.
- Gamo T., Inokuchi T. and Laufer H. (1977) Polypeptides of fibroin and sericin secreted from the different sections of the silk gland in Bombyx mori. *Insect Biochemistry* **7**, 285-295.
- Grzelak K. (1995) Control of expression of silk protein genes. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology* **110**, 671-681.

- Hakkak R., Korourian S., Ronis M. J. J., Johnston J. M. and Badger T. M. (2001a) Dietary Whey Protein Protects against Azoxymethane-induced Colon Tumors in Male Rats. *Cancer Epidemiol Biomarkers Prev* **10**, 555-558.
- Hakkak R., Korourian S., Ronis M. J. J., Johnston J. M. and Badger T. M. (2001b) Soy protein isolate consumption protects against azoxymethane-induced colon tumors in male rats. *Cancer Letters* **166**, 27-32.
- Ishida H., Miwa H., Tatsuta M., Masutani S., Imamura H., Shimizu J., Ezumi K., Kato H., Kawasaki T., Furukawa H. and Kawakami H. (2004) Ki-67 and CEA expression as prognostic markers in Dukes' C colorectal cancer. *Cancer Letters* **207**, 109-115.
- Johnson I. T. (2004) New approaches to the role of diet in the prevention of cancers of the alimentary tract. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis Nutrition and Carcinogenesis* **551**, 9-28.
- Kato N., Sato S., Yamanaka A., Yamada H., Fuwa N. and Nomura M. (1998) Silk protein, sericin, inhibits lipid peroxidation and tyrosinase activity. *Biosci Biotechnol Biochem* **62**, 145-147.
- Khuhaprema T. and Srivatanakul P. (2008) Colon and Rectum Cancer in Thailand: An Overview. *Jpn. J. Clin. Oncol.* **38**, 237-243.
- Kundu S. C., Dash B. C., Dash R. and Kaplan D. L. (2008) Natural protective glue protein, sericin bioengineered by silkworms: Potential for biomedical and biotechnological applications. *Progress in Polymer Science* **33**, 998-1012.
- Kurioka A., Kurioka F. and Yamazaki M. (2004) Characterization of Sericin Powder Prepared from Citric Acid-degraded Sericin Polypeptides of the Silkworm, *Bombyx Mori*. *Bioscience, Biotechnology, and Biochemistry* **68**, 774-780.
- Labianca R., Beretta G. D., Kildani B., Milesi L., Merlin F., Mosconi S., Pessi M. A., Prochilo T., Quadri A., Gatta G., de Braud F. and Wils J. Colon cancer. *Critical Reviews in Oncology/Hematology In Press, Corrected Proof*.
- Levi F., Pasche C., Lucchini F. and La Vecchia C. (2001) Dietary fibre and the risk of colorectal cancer. *European Journal of Cancer* **37**, 2091-2096.
- Li Y. G., Ji D. F., Lin T. B., Zhong S., Hu G. Y. and Chen S. (2008) Protective effect of sericin peptide against alcohol-induced gastric injury in mice. *Chin Med J (Engl)* **121**, 2083-2087.
- Liu Z., Ishikawa W., Huang X., Tomotake H., Kayashita J., Watanabe H. and Kato N. (2001) A Buckwheat Protein Product Suppresses 1,2-Dimethylhydrazine-Induced Colon Carcinogenesis in Rats by Reducing Cell Proliferation. *J. Nutr.* **131**, 1850-1853.
- Magnuson B. A., Carr I. and Bird R. P. (1993) Ability of Aberrant Crypt Foci Characteristics to Predict Colonic Tumor Incidence in Rats Fed Cholic Acid. *Cancer Res* **53**, 4499-4504.

- McIntosh G. H. and Le Leu R. K. (2001) The influence of dietary proteins on colon cancer risk. *Nutrition Research* **21**, 1053-1066.
- McLellan E. A. and Bird R. P. (1988) Aberrant crypts: potential preneoplastic lesions in the murine colon. *Cancer Res* **48**, 6187-6192.
- Morita T., Oh-hashi A., Takei K., Ikai M., Kasaoka S. and Kiriyma S. (1997) Cholesterol-Lowering Effects of Soybean, Potato and Rice Proteins Depend On Their Low Methionine Contents In Rats Fed a Cholesterol-Free Purified Diet. *J. Nutr.* **127**, 470-477.
- Pollack A., DeSilvio M., Khor L. Y., Li R., Al-Saleem T. I., Hammond M. E., Venkatesan V., Lawton C. A., Roach M., III, Shipley W. U., Hanks G. E. and Sandler H. M. (2004) Ki-67 Staining Is a Strong Predictor of Distant Metastasis and Mortality for Men With Prostate Cancer Treated With Radiotherapy Plus Androgen Deprivation: Radiation Therapy Oncology Group Trial 92-02. *J Clin Oncol* **22**, 2133-2140.
- Reddy B. S., Weisburger J. H. and Wynder E. L. (1974) Fecal Bacterial beta-Glucuronidase: Control by Diet. *Science* **183**, 416-417.
- Rogers A. E., Zeisel S. H. and Groopman J. (1993) Diet and carcinogenesis. *Carcinogenesis* **14**, 2205-2217.
- Rose M. L., Madren J., Bunzendahl H. and Thurman R. G. (1999) Dietary glycine inhibits the growth of B16 melanoma tumors in mice. *Carcinogenesis* **20**, 793-798.
- Sasaki M., Yamada H. and Kato N. (2000) A Resistant Protein, Sericin Improves Atropine-Induced Constipation in Rats. *Food Science and Technology Research* **6**, 280-283.
- Takahashi T., Satou M., Watanabe N., Sakaitani Y., Takagi A., Uchida K., Ikeda M., Moriyama R., Matsumoto K. and Morotomi M. (1999) Inhibitory effect of microfibril wheat bran on azoxymethane-induced colon carcinogenesis in CF1 mice. *Cancer Letters* **141**, 139-146.
- Tamada Y., Sano M., Niwa K., Imai T. and Yoshino G. (2004) Sulfation of silk sericin and anticoagulant activity of sulfated sericin. *J Biomater Sci Polym Ed* **15**, 971-980.
- Terada S., Sasaki M., Yanagihara K. and Yamada H. (2005) Preparation of silk protein sericin as mitogenic factor for better mammalian cell culture. *J Biosci Bioeng* **100**, 667-671.
- Tokutake S. (1980) Isolation of the smallest component of silk protein. *Biochem J* **187**, 413-417.
- van Breda S. G., de Kok T. M. and van Delft J. H. (2007) Mechanisms of colorectal and lung cancer prevention by vegetables: a genomic approach. *J Nutr Biochem*.
- Wogan G. N. (1985) Diet and nutrition as risk factors for cancer. *Princess Takamatsu Symp* **16**, 3-10.
- Zhang X. M., Stamp D., Minkin S., Medline A., Corpet D. E., Bruce W. R. and Archer M. C. (1992) Promotion of Aberrant Crypt Foci and Cancer in Rat Colon by Thermolyzed Protein. *J. Natl. Cancer Inst.* **84**, 1026-1030.

- Zhang Y.-Q. (2002) Applications of natural silk protein sericin in biomaterials. *Biotechnology Advances* **20**, 91-100.
- Zhaorigetu S., Sasaki M. and Kato N. (2007) Consumption of sericin suppresses colon oxidative stress and aberrant crypt foci in 1,2-dimethylhydrazine-treated rats by colon undigested sericin. *J Nutr Sci Vitaminol (Tokyo)* **53**, 297-300.
- Zhaorigetu S., Sasaki M., Watanabe H. and Kato N. (2001) Supplemental silk protein, sericin, suppresses colon tumorigenesis in 1,2-dimethylhydrazine-treated mice by reducing oxidative stress and cell proliferation. *Biosci Biotechnol Biochem* **65**, 2181-2186.
- Zhaorigetu S., Yanaka N., Sasaki M., Watanabe H. and Kato N. (2003a) Inhibitory effects of silk protein, sericin on UVB-induced acute damage and tumor promotion by reducing oxidative stress in the skin of hairless mouse. *Journal of Photochemistry and Photobiology B: Biology* **71**, 11-17.
- Zhaorigetu S., Yanaka N., Sasaki M., Watanabe H. and Kato N. (2003b) Silk protein, sericin, suppresses DMBA-TPA-induced mouse skin tumorigenesis by reducing oxidative stress, inflammatory responses and endogenous tumor promoter TNF-alpha. *Oncol Rep* **10**, 537-543.

**Table 1**

Amino acid composition of the sericin

Amino acid	% w/w
Serine	33.40
Aspartate	16.70
Glutamate	4.40
Glycine	13.50
Threonine	9.70
Lysine	3.30
Tyrosine	2.60
Arginine	3.10
Alanine	6.00
Valine	2.80
Histidine	1.30
Luecine	1.10
Isoluecine	0.70
Phenylalanine	0.50
Trytophan	0.20
Proline	0.70
Cystine	0.20
Methionine	0.04

**Table 2**

## Body weight and food consumption

Group	Treatment groups	Initial body weight (g)	Final body weight (g)	Body-weight gain (g)	Initial food consumption n (g/rat/day)	Final food consumption (g/rat/day)
1	Casein	237.38 ± 11.61	502.75 ± 28.25	265.37 ± 16.64	10.73 ± 2.76	18.21 ± 0.90
2	Sericin	234.67 ± 30.53	488.83 ± 40.81	254.16 ± 10.28	12.52 ± 2.40	19.37 ± 2.70
3	Casein + DMH	228.54 ± 37.24	498.08 ± 29.73	269.54 ± 7.54	11.84 ± 2.70	22.38 ± 2.70*
4	Sericin + DMH	206.04 ± 25.24	483.38 ± 24.84	277.34 ± 0.40	8.41 ± 3.00#	21.89 ± 2.70**#
5	Post-sericin + DMH	229.00 ± 35.06	486.83 ± 48.42	257.83 ± 13.36	12.37 ± 2.80	21.21 ± 1.00*

Values are mean ± SD of 6 rats

\* Significantly different from casein diet by Student's t-test ( $p < 0.05$ )# Significantly different from sericin diet by Student's t-test ( $p < 0.05$ )

**Table 3**

Incidences of aberrant crypt foci (ACF) and tumor formation in rat colon

Group	Treatment groups	No. of rats	ACF and tumor formation		
			Tumor	ACF	Number of
			Incidence (%)	Incidence (%)	ACF/cm <sup>2</sup> /rat
1	Casein	6	0	0	0
2	Sericin	6	0	0	0
3	Casein + DMH	6	2 (33)	6 (100)	116.01 ± 48.52
4	Sericin + DMH	6	0	6 (100)	79.96 ± 29.04
5	Post-sericin + DMH	6	0	6 (100)	82.35 ± 39.83

Values are mean ± SD

**Table 4**

Distribution and multiplicity of aberrant crypt foci (ACF) in rat colon

	Treatment groups		
	DMH	Sericin + DMH	Post-sericin + DMH
<i>ACF distribution (No. ACF/cm<sup>2</sup>)</i>			
Proximal colon	3.90 ± 5.30	0	0
Middle colon	37.99 ± 27.50	21.00 ± 18.47	28.35 ± 29.78
Distal colon	73.38 ± 27.02	57.79 ± 26.38	53.46 ± 22.78
<i>Average ACF multiplicity (AC/ACF)</i>			
Proximal colon	0.44 ± 0.44	0	0
Middle colon	0.78 ± 0.17	0.67 ± 0.16	0.60 ± 0.13
Distal colon	3.57 ± 0.17	3.53 ± 0.57	3.25 ± 0.28
<i>Type of ACF in distal colon (No. ACF/cm<sup>2</sup>)</i>			
1 AC/ACF	6.49 ± 6.27	6.06 ± 7.69	5.84 ± 6.96
2 AC/ACF	12.01 ± 8.70	12.34 ± 10.48	11.26 ± 7.19
3 AC/ACF	9.74 ± 6.49	7.58 ± 6.49	9.52 ± 7.47
4 AC/ACF	7.47 ± 7.14	3.68 ± 1.73	5.41 ± 4.89
≥ 5 AC/ACF	37.66 ± 10.12	28.79 ± 15.79	21.43 ± 9.03*
(Area of individual ACF, mm <sup>2</sup> )	(0.14 ± 0.06)	(0.11 ± 0.05*)	(0.14 ± 0.06)

Values are mean ± SD

\* Significantly different from casein diet by Student's *t*-test (*p* < 0.05)

**Table 5**

## Lipid peroxidation of distal part of colon

Group	Treatment groups	MDA concentration ( $\mu\text{M}$ )
1	Casein	1.26 $\pm$ 0.51
2	Sericin	0.81 $\pm$ 0.21
3	Casein + DMH	1.77 $\pm$ 1.26
4	Sericin + DMH	1.08 $\pm$ 0.71
5	Post-sericin + DMH	1.15 $\pm$ 0.64

Values are mean  $\pm$  SD

**Table 6**

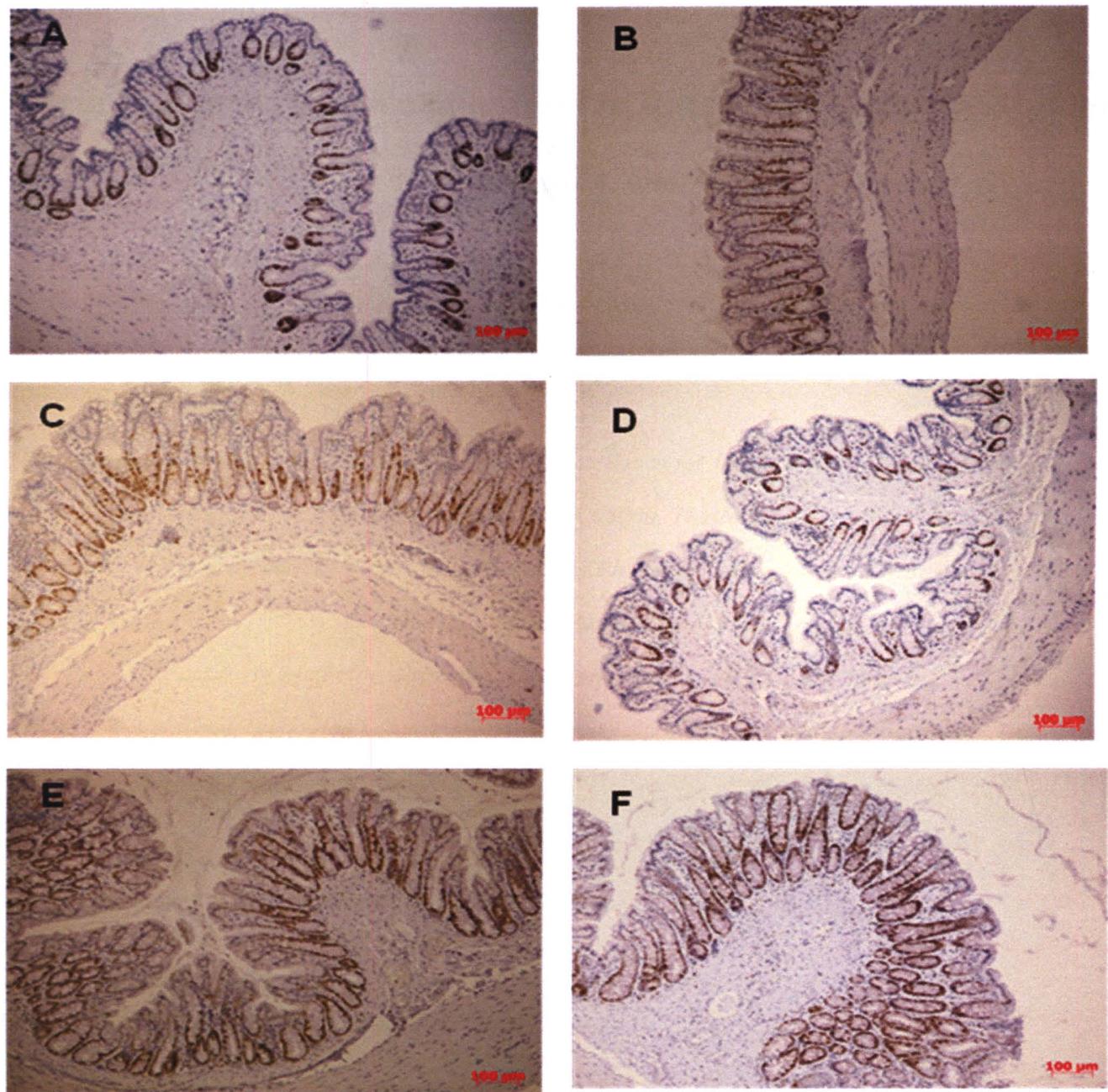
## Immunohistochemical analysis of Ki67 proliferation marker

	Treatment groups				
	Casein	Sericin	Casein + DMH	Sericin + DMH	Post-sericin + DMH
<i>ki67 positive cells</i>					
Total number (cells/crypts)	15.78 ± 3.00	18.25 ± 6.97	17.08 ± 4.42	28.55 ± 10.32 *†	29.82 ± 9.55 *†#
<i>length of crypt (μm)</i>					
Total length (μm)	220.62 ± 34.90	228.64 ± 34.90	224.42 ± 19.55	237.30 ± 46.67	270.08 ± 13.83 *†#
Ki67 length (μm)	102.93 ± 18.08	107.61 ± 29.74	101.66 ± 16.14	147.83 ± 46.58	174.12 ± 27.45 *†#
Relative distance of Ki67	0.46 ± 0.05	0.47 ± 0.08	0.45 ± 0.05	0.62 ± 0.10 *†#	0.64 ± 0.10 *†#
Relative total number of Ki67	0.08 ± 0.02	0.08 ± 0.04	0.08 ± 0.02	0.12 ± 0.03 *#	0.11 ± 0.04
<i>distribution of Ki67 positive cells</i>					
Top zone	11.63 ± 2.30	14.05 ± 5.36	12.33 ± 3.39 *	17.63 ± 3.22 *†#	17.67 ± 5.23 *†
Middle zone	5.02 ± 2.17	4.52 ± 2.17	4.73 ± 2.30	9.70 ± 5.89	10.85 ± 4.45 *†#
Basal zone	0.07 ± 0.10	0.05 ± 0.12	0.03 ± 0.05	1.17 ± 1.79	1.30 ± 1.35 †

Values are mean ± SD

Data were analyzed from 60 crypts (10 crypts/rat)

\* Significantly different from casein diet by Student's t-test ( $p < 0.05$ )† Significantly different from sericin diet by Student's t-test ( $p < 0.05$ )# Significantly different from casein+ DMH by Student's t-test ( $p < 0.05$ )



**Figure 1.** Morphology of colonic crypt from normal rats fed casein diet (A) and sericin diet (B) as well as DMH-treated rats fed with casein diet (C, D), sericin diet (E) and post sericin diet (F)



2. การทดสอบฤทธิ์ลดการดูดซึมโคเลสเทอโรลในเซลล์กำไส้เพาะเลี้ยงและในสัตว์ทดลอง

## Sericin Reduces Plasma Cholesterol in Rats and Cholesterol Uptake in Caco-2 Cells

Nanteetip Limpeanchob<sup>a,\*</sup>, Kanithaporn Trisat<sup>a</sup>, Waree Tiyaboonchai<sup>b</sup>, Sutatip Pongcharoen<sup>c</sup>, Manote Sutheerawattananonda<sup>d</sup>

<sup>a</sup> Department of Pharmacy Practice and Center of Excellence for Innovation in Chemistry, Naresuan University, Phitsanulok 65000, Thailand

<sup>b</sup> Department of Pharmaceutical Technology, Faculty of Pharmaceutical Sciences, Naresuan University, Phitsanulok 65000, Thailand

<sup>c</sup> Faculty of Medicine, Naresuan University, Phitsanulok 65000, Thailand

<sup>d</sup> Department of Food Technology, Suranaree University of Technology, Nakonrachasrima, Thailand

\* Corresponding author;

Name              Nanteetip Limpeanchob

Address          Faculty of Pharmaceutical Sciences, Naresuan University,  
                      Phitsanulok 65000, Thailand

Tel                +66-81-554-3013,        Fax                +66-55-963-731

Email              [nanteetipl@yahoo.com](mailto:nanteetipl@yahoo.com), [nanteetipl@nu.ac.th](mailto:nanteetipl@nu.ac.th)

Research funding: Thailand Research Fund (TRF)

**Keywords:** hypocholesterolemic effect, sericin, cholesterol uptake, cholesterol, protein

## ABSTRACT

Sericin is an eatable protein isolated from silk cocoon and its consumption has been suggested to be beneficial for health. Many dietary proteins have been shown to reduce serum lipids, but this lipid lowering effect of sericin has not been studied. The purpose of this study was to evaluate the hypocholesterolemic effect of sericin in rats and its potential mechanism of actions *in vitro* models. The hypocholesterolemic effect was tested in rats fed a high cholesterol diet for 14 days. Total cholesterol and non-high density lipoprotein (non-HDL) cholesterol were significantly reduced in rats fed high-cholesterol diet together with all three tested doses of sericin (10, 100 and 1000 mg/kg/day). HDL cholesterol and triglyceride levels were not affected by sericin supplement. The effect of sericin on cholesterol absorption was determined by measuring the uptake of radio-labeled cholesterol into differentiated Caco-2 cells. Low concentrations of sericin (25 and 50 µg/ml) inhibited cholesterol uptake about 30%, whereas higher concentrations showed no effect. Cholesterol micellar solubility was also reduced in the presence of sericin. This study is the first evidence suggesting the hypocholesterolemic of sericin is partially the result of its inhibitory activity on cholesterol absorption in intestinal cells.

**Keywords:** hypocholesterolemic effect, sericin, cholesterol uptake, cholesterol, protein

## 1. Introduction

There is growing interest not only in functional foods but also dietary supplements and nutraceutical products to improve human health, especially those that promote prevention of certain diseases. Since high cholesterol level increases risk of many cardiovascular diseases, the cholesterol lowering agents have become one of the major targets for prevention of those diseases. Several reports have indicated that various types of dietary proteins could affect serum cholesterol levels [1-5]. The most extensively studied proteins are soybean and milk proteins. Some other sources of proteins such as rice, buckwheat, potato and sunflower appeared to have cholesterol lowering effect [1,2,6,7]. It should be noted that most studies have been focused on proteins isolated from regular diets. The present study was aimed to investigate cholesterol lowering effect of sericin, an eatable protein found in silk cocoon.

Silk proteins produced by silkworm are composed of fibroin and sericin. Sericin contributes 25-30% of the total cocoon weight. It has high serine content (30-33%) and its role is to help the formation of a cocoon by gluing the fibroin together. Sericin is a mixture of macromolecule polypeptides, its molecular weight ranges widely from about 10 to over 300 kDa [8]. In general silk processing, sericin is removed and mostly discarded in the wastewater. In the past decade, there are attempts to discover its biological activities which then make it a valuable natural ingredient particularly in food and cosmetic purposes. Sericin's biological functions include anti-oxidation [9], tyrosinase inhibition [9], chemoprevention [10-12], and protection against alcohol-induced liver and gastric injuries [13], UV-induced keratinocyte apoptosis [14].

Since sericin is protease-resistant with low digestibility property [15], it was received much attention to be beneficial for the health of intestines. Consumption of sericin suppressed colon tumorigenesis in animals treated with 1,2-dimethylhydrazine-treated (DMH), a potent colon carcinogen [10,11]. Consumption of sericin could have more interesting undiscovered advantages, such as cholesterol lowering effect as above mentioned dietary proteins. Thus, the present study was carried out to obtain the hypcholesterolemic activity of sericin in rats fed high cholesterol diet and its potential mechanism of actions to inhibit cholesterol absorption in differentiated Caco-2 cells and cholesterol micellar solubility.

## 2. Materials and Methods

### 2.1. Materials

Dulbecco's modified Eagle's medium (DMEM)/F12 and all the materials used in cell culture and cholesterol, phosphatidylcholine and sodium taurocholate were purchased from Sigma Chemical Co. (St. Louis, MO). Fetal bovine serum (FBS) was purchased from Gibco. [ $1\alpha,2\alpha(n)$ - $^3$ H]cholesterol was purchased from PerkinElmer. Sericin was provided by the Institution of Agricultural Technology, Suanaree University of Technology, Nakhon Ratchasima, Thailand. The analysis of amino acid composition is showed in Table 1.

### 2.2. Animal experiment

Twenty five male Sprague-Dawley rats (200–250 g) were obtained from the National Laboratory Animal Centre, Mahidol University, Thailand. All animal experimental procedures were approved by the Animal Research Ethics Committee, Naresuan University, Thailand. Animals were housed in stainless steel cages at  $22 \pm 2$  °C and 12 h light/dark cycle and had free access to water and food for 1 week of acclimatization before the beginning experiment. Rats were randomly divided into four groups consisting of 5 animals each. The animals were fed a standard diet with high-cholesterol supplement (cholesterol, bile extract, and coconut oil at 1.5, 0.75 and 0.75 g/kg body weight/day, respectively). The amount of cholesterol each animal received was approximately 2% of daily diet. The cholesterol mixtures were daily administered with and without sericin solutions by gastric tube for 14 days. Fasting blood was collected at day 0 (baseline), 4, 8, 11 and 14 from the tail vein to measure plasma lipid levels. During the experimental period, body weight and food intake were regularly recorded.

### 2.3. Measurement of plasma lipid levels

Plasma total cholesterol, triglyceride and HDL-cholesterol levels were measured with commercial enzymatic assay kits (HUMAN GmbH, Germany). The procedures were conducted as manufacturer recommendation. The levels of non-HDL cholesterol were calculated by subtracting HDL-cholesterol from total cholesterol.

#### **2.4. Cell culture preparation**

Caco-2 cells were obtained from the American Type Culture Collection (ATCC). Cells were grown in Dulbecco's modified Eagle's medium (DMEM)/F12 containing 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin. Cells were maintained at 37 °C in CO<sub>2</sub> incubator in a saturated humidity atmosphere containing 95% air and 5% CO<sub>2</sub>. All cells were propagated in culture flasks and subsequently plated in 24-well plates for cholesterol uptake experiment.

#### **2.5. Cholesterol micelle preparation**

Briefly, stock solutions of [1 $\alpha$ ,2 $\alpha$ (n)-<sup>3</sup>H]cholesterol, cholesterol, phosphatidylcholine were dissolved in chloroform. A stock solution of sodium taurocholate was prepared in methanol. The lipid and bile salt solutions were mixed and evaporated under a stream of N<sub>2</sub>. The lipid film was stored under N<sub>2</sub> at -20 °C until use. The micelle solutions were freshly prepared by hydrating the lipid film in serum-free DMEM/F12 so that the final concentrations of the micelle were 1 μM cholesterol, 2 mM sodium taurocholate, 50 μM phosphatidylcholine, and 1 μCi/ml [1 $\alpha$ ,2 $\alpha$ (n)-<sup>3</sup>H]cholesterol. The micelle solutions were sonicated and passed through 0.2 μm syringe filters and kept at 37 °C before adding to the cells.

#### **2.6. Cholesterol uptake assay**

Caco-2 cells were seeded on 24-well plate at a cell density 50,000 cells/well and cultured for 14 days to allow them to differentiate. During this period cells were fed with fresh medium every 2 days. After 14 days, cells were incubated with serum-free medium overnight. Cells were treated with sericin or ezetimibe (positive control) for 1 h before adding [1 $\alpha$ ,2 $\alpha$ (n)-<sup>3</sup>H]cholesterol-micelles. After 3 h incubation, medium were removed and cells were washed twice with ice-cold PBS. The cells were disrupted with 0.2 N NaOH and 0.1 % Triton-X 100. One part of the aliquot was added to scintillation cocktail (MicroScint™-20; PerkinElmer), the other part was taken for protein determination by BCA protein assay kit (Pierce). The radioactivities of cell lysates were measured in a Packard β-counter.

### **2.7. Micellar cholesterol solubility assay**

Cholesterol micelles were prepared with ultrasonication of a 7ml micellar dispersion containing 10 mM sodium taurocholate, 2 mM cholesterol, 5 mM oleic acid, 132 mM NaCl and 15 mM sodium phosphate (pH 7.4). Various concentration of sericin or BSA (500-2000 $\mu$ g/ml) were added to the mixed micelle solution before and after micelle preparation and incubated for 24 h at 37°C. The precipitated cholesterol was separated from the intermicellar cholesterol by filtering through a 0.22  $\mu$ m. The intermicellar cholesterol in the filtrates were determined by cholesterol assay kit (Human).

### **2.8. Statistical analysis**

All data from animal experiments are expressed as means  $\pm$  standard deviation (SD). Means  $\pm$  standard error of means (SEM) were used for the *in vitro* experiments. The data were analyzed by one-way analysis of variance (ANOVA). Differences were considered to be significance when p value  $\leq$  0.05.

### 3. Results

#### 3.1. Lipid levels in rat plasma

The mean of total cholesterol levels of four groups are showed in Fig. 1. Before feeding high-cholesterol mixture, the baseline levels of plasma lipids were measured. Since the entire experimental period is 14 days in which the increasing age was expected not to influence the lipid profiles, therefore plasma lipid levels at day 0 (baseline) were used as control levels. Rats fed normal diet were not included in this study in order to minimize the number of animals. After cholesterol administration, plasma cholesterol level was continuously increased and a significant increase was observed after days 4 of feeding. Plasma cholesterol levels were significantly reduced in rats fed high-cholesterol mixtures with sericin. All tested doses of sericin, 10, 100 and 1000 µg/kg body weight/day, effectively reduced cholesterol levels after days 8 of feeding but dose-dependent pattern is not clearly established. This data suggest that sericin supplement could suppress plasma cholesterol level in rats given high-cholesterol diet.

The plasma levels of high density lipoprotein cholesterol (HDL-C), non-HDL cholesterols and triglyceride (TG) were showed in Fig. 2. The levels of plasma HDL cholesterol in all groups were not much different across 14 days of administrations, although there was slightly increase at days 4 in sericin fed groups. Non-HDL cholesterol (VLDL and LDL cholesterols) levels were appeared in the similar pattern to total cholesterol levels, but the dose-dependent effect of sericin to reduce these types of lipoproteins was apparently demonstrated at days 11 and 14. Plasma TG levels were also slightly increased from baseline in all groups over treatment period, and sericin showed no TG lowering effect (Fig. 2). Taken all data together, sericin supplement effectively reduce plasma levels of total cholesterol and non-HDL cholesterol in rats fed high-cholesterol diet.

#### 3.2. Cholesterol uptake in differentiated Caco-2 cells

To examine the effect of the sericin on cholesterol absorption in the intestine, differentiated Caco-2 cells were used as an *in vitro* model. The level of [ $1\alpha,2\alpha(n)-^3H$ ] cholesterol captured in Caco-2 cells was measured and calculated as the amount of tritium cholesterol per mg

protein of cell lysates. In this study, ezetimibe was used as a positive control since it is a cholesterol absorption inhibitor by blocking cholesterol transporter protein. Bovine serum albumin (BSA) was used as a control protein. The results showed the significant decreases in uptake of cholesterol micelles into differentiated Caco-2 cells pre-treated with sericin at 25 and 50 µg/ml (Fig. 3). Unexpectedly, cholesterol uptake inhibitory activity was decreasing with increasing concentration of sericin. Sericin at 1000 µg/ml somehow increased the uptake of cholesterol. This pattern of cholesterol uptake was also observed with BSA, but BSA at 25 and 50 µg/ml did not significantly reduced cholesterol uptake whereas its higher concentrations (100-1000 µg/ml) slightly increased the uptake of cholesterol. This data suggest that sericin could either reduce or promote cholesterol uptake into intestinal cells depending on its concentrations.

### *3.3. Lipid micellar cholesterol solubility*

The effect of sericin on the micellar solubility of cholesterol is shown in Table 3. The cholesterol solubility in mixed lipid micelles was significantly decreased in the presence of sericin particularly before and after micelle preparation. The inhibitory activity of sericin on cholesterol solubility was dose-dependent. BSA as control protein showed no reduction of cholesterol micellar solubility.

#### 4. Discussion

To our knowledge, this is the first study showing that silk protein, sericin, lowers cholesterol levels in rats. Our data provide evidence that sericin significantly suppresses the plasma levels of total cholesterol and non-HDL cholesterol in high-cholesterol diet-induced hypercholesterolemia in rats. This cholesterol lowering effect is dose-dependent. Although its effect is significant, sericin could not bring cholesterol levels back to the baseline.

The present study choose to reports non-HDL cholesterol since calculated LDL-cholesterol is still controversial especially in rat serum. In human, serum LDL cholesterol can be calculated by Friedewald equation,  $[LDL\text{-Cholesterol}] = [Total\text{-Cholesterol}] - [HDL\text{-Cholesterol}] - [Triglyceride]/5$  [16]. In rat, this equation however seemed not to be suitable for certain circumstances, particularly in hypercholesterolaemic rats [17]. Non-HDL cholesterol in the present study mainly represents LDL and partially VLDL cholesterols.

Because a strong positive correlation was observed between serum cholesterol concentration and dietary methionine concentration or methionine:glycine ratio [6], the low methionine content in dietary proteins was though to be involved in their cholesterol lowering effects [6,18]. Sericin is one of the protein that contains low methionine concentration (<0.05 mol %) [11,13]. The glycine content in sericin is about 200 times higher than methionine, which makes methionine:glycine ratio to 0.005 [11,13]. The amino acid composition of the sericin protein used in the present study is similar to previous reports [11,13]. Our sericin contains 0.04% w/w methionine and 13.5% w/w glycine which makes its ratio about 0.003. The decreases in apoprotein A-I and HDL secretion from rat livers were suggested to partially responsible for cholesterol lowering effect of low methionine protein diet [19,20]. In this study, the increases in HDL cholesterol were observed at days 4 of sericin administration, but this effect was not clear at longer periods of treatment. Although the exact mechanism to lower serum cholesterol level is not clearly identified, low-methionine content seems be involved in the hypocholesterolemic effect of some dietary proteins such as soybean, potato and rice [6]. It should be noted that the amount of methionine and

methionine:glycine ratio in sericin protein are far lower than those dietary proteins. Methionine and glycine contents in these dietary proteins are 1-2 and 3-4.5 % w/w, respectively [6,21].

Sericin is a dietary fiber-like protein with very low digestibility [15]. Undigested sericin was found to play a role in the mechanism of its chemopreventive effect against colon tumorigenesis [10]. According to its protease resistance property, undigested sericin might affect the absorption of some substances in the intestinal lumen. Sericin consumption was showed to enhance intestinal absorption of zinc, iron, magnesium and calcium in rats [22]. Disruption of cholesterol absorption of undigested sericin might responsible for its cholesterol lowering effect. However, the *in vivo* experiment did not allow determination the effect of sericin on cholesterol absorption. To investigate this mechanism of action, differentiated Caco-2 cell cultures were therefore used. Caco-2 cells at 2-3 weeks in culture were proliferated and differentiated to become the monolayer intestinal epithelium [23]. The present study demonstrated that sericin reduced cholesterol uptake into differentiated Caco-2 but surprisingly showed inverse dose-dependent activity. In cellular experiment, sericin protein was tested in the comparison with BSA. High concentration of both proteins seem to promote cholesterol uptake. The explanation for this observation have not been thoroughly investigated at this time, it at least give us the direction for further investigation.

There are evidences indicated that dietary proteins decreased cholesterol absorption in Caco-2 cell model [21,24]. Disruption of cholesterol solubility referring to incorporation of cholesterol into micelles is one of the mechanisms of plant phytosterols (Jesch 2006) and some proteins [2,24] to reduce cholesterol absorption in animal models. Alteration of cholesterol solubility in lipid micelles may affect the efficacy to micelles to be taken up by Caco-2 cells. Micellar cholesterol solubility *in vitro* was significantly lower in the presence of soy protein derivatives and possibly lead to cholesterol absorption inhibition in rats and in caco-2 cells [21]. Buckwheat protein was shown to reduce cholesterol solubility in lipid micelles only when it was added prior to micelle formation but this effect was not observed if buckwheat protein was

added after lipid micelle formation [24]. In the current study, cholesterol solubility in mixed lipid micelles was significantly reduced in the presence of increasing concentration of sericin either before and after micelle formation.

We have found for the first time that protein from silk cocoon, sericin, has cholesterol lowering effect which partially due to its inhibitory activity against cholesterol absorption silk cocoon. Although higher concentration of sericin unexpectedly exhibits lesser inhibitory activity against cholesterol absorption in cell culture models, this inverse correlation between dose and activity is not observed in animal experiment.

#### **Acknowledgements**

The authors would like to thank xxx for suggestions in preparing the manuscript. This study was financial supported by Thailand Research Fund and partially supported of the graduate program by the Center of Excellence for Innovation in Chemistry (PERCH-CIC).

## References

- [1] Liyanage R, Han KH, Watanabe S, Shimada K, Sekikawa M, Ohba K, Tokuji Y, Ohnishi M, Shibayama S, Nakamori T, Fukushima M. Potato and soy peptide diets modulate lipid metabolism in rats. *Biosci Biotechnol Biochem* 2008;72 (4):943-50.
- [2] Megías C, Pedroche J, Del Mar Yust M, Alaiz M, Girón-Calle J, Millán F, Vioque J. Sunflower protein hydrolysates reduce cholesterol micellar solubility. *Plant Foods Hum Nutr* 2009;64 (2):86-93.
- [3] Nagaoka S, Futamura Y, Miwa K, Awano T, Yamauchi K, Kanamaru Y, Tadashi K, Kuwata T. Identification of novel hypocholesterolemic peptides derived from bovine milk beta-lactoglobulin. *Biochem Biophys Res Commun* 2001;281 (1):11-7.
- [4] Zhang X, Beynen AC. Influence of dietary fish proteins on plasma and liver cholesterol concentrations in rats. *Br J Nutr* 1993;69 (3):767-77.
- [5] Zhong F, Liu J, Ma J, Shoemaker CF. Preparation of hypocholesterol peptides from soy protein and their hypocholesterolemic effect in mice. *Food Res Int* 2007;40 (6):661-7.
- [6] Morita T, Oh-hashi A, Takei K, Ikai M, Kasaoka S, Kiriyama S. Cholesterol-lowering effects of soybean, potato and rice proteins depend on their low methionine contents in rats fed a cholesterol-free purified diet. *J Nutr* 1997;127 (3):470-7.
- [7] Kayashita J, Shimaoka I, Nakajoh M, Yamazaki M, Kato N. Consumption of buckwheat protein lowers plasma cholesterol and raises fecal neutral sterols in cholesterol-Fed rats because of its low digestibility. *J Nutr* 1997;127 (7):1395-400.
- [8] Zhang YQ. Applications of natural silk protein sericin in biomaterials. *Biotechnol Adv* 2002;20 (2):91-100.
- [9] Kato N, Sato S, Yamanaka A, Yamada H, Fuwa N, Nomura M. Silk protein, sericin, inhibits lipid peroxidation and tyrosinase activity. *Biosci Biotechnol Biochem* 1998;62 (1):145-7.
- [10] Zhaorigetu S, Sasaki M, Kato N. Consumption of sericin suppresses colon oxidative stress and aberrant crypt foci in 1,2-dimethylhydrazine-treated rats by colon undigested sericin. *J Nutr Sci Vitaminol (Tokyo)* 2007;53 (3):297-300.
- [11] Zhaorigetu S, Sasaki M, Watanabe H, Kato N. Supplemental silk protein, sericin, suppresses colon tumorigenesis in 1,2-dimethylhydrazine-treated mice by reducing oxidative stress and cell proliferation. *Biosci Biotechnol Biochem* 2001;65 (10):218-6.
- [12] Zhaorigetu S, Yanaka N, Sasaki M, Watanabe H, Kato N. Silk protein, sericin, suppresses DMBA-TPA-induced mouse skin tumorigenesis by reducing oxidative stress, inflammatory responses and endogenous tumor promoter TNF-alpha. *Oncol Rep* 2003;10 (3):537-43.
- [13] Li YG, Ji DF, Chen S, Hu GY. Protective effects of sericin protein on alcohol-mediated liver damage in mice. *Alcohol Alcohol* 2008;43 (3):246-53.
- [14] Dash R, Mandal M, Ghosh SK, Kundu SC. Silk sericin protein of tropical tasar silkworm inhibits UVB-induced apoptosis in human skin keratinocytes. *Mol Cell Biochem* 2008;311 (1-2):111-9.
- [15] Sasaki M, Yamada H, Kato N. A resistant protein: sericin improves atropine-induced constipation in rats. *Food Sci Technol Res* 2000;6:280-3.
- [16] Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clin Chem* 1972;18 (6):499-502.
- [17] Sanchez-Muniz FJ, Bastida S. Do not use the Friedewald formula to calculate LDL-cholesterol in hypercholesterolaemic rats. *Eur J Lipid Sci Technol* 2008;110 (4):295 - 301.

- [18] Gudbrandsen OA, Wergedahl H, Liaset B, Espe M, Berge RK. Dietary proteins with high isoflavone content or low methionine-glycine and lysine-arginine ratios are hypocholesterolaemic and lower the plasma homocysteine level in male Zucker fa/fa rats. *Br J Nutr* 2005;94 (3):321-30.
- [19] Oda H, Fukui H, Hitomi Y, Yoshida A. Alteration of serum lipoprotein metabolism by polychlorinated biphenyls and methionine in rats fed a soybean protein diet. *J Nutr* 1991;121 (7):925-33.
- [20] Oda T, Aoe S, Sanada H, Ayano Y. Effects of soluble and insoluble fiber preparations isolated from oat, barley, and wheat on liver cholesterol accumulation in cholesterol-fed rats. *J Nutr Sci Vitaminol (Tokyo)* 1993;39 (1):73-9.
- [21] Nagaoka S, Miwa K, Eto M, Kuzuya Y, Hori G, Yamamoto K. Soy protein peptic hydrolysate with bound phospholipids decreases micellar solubility and cholesterol absorption in rats and caco-2 cells. *J Nutr* 1999;129 (9):1725-30.
- [22] Sasaki M, Yamada H, Kato N. Consumption of silk protein, sericin elevates intestinal absorption of zinc, iron, magnesium and calcium in rats. *Nutri Res* 2000;20 (10):1505-11.
- [23] Hidalgo IJ, Raub TJ, Borchardt RT. Characterization of the human colon carcinoma cell line (Caco-2) as a model system for intestinal epithelial permeability. *Gastroenterology* 1989;96 (3):736-49.
- [24] Metzger BT, Barnes DM, Reed JD. Insoluble fraction of buckwheat (*Fagopyrum esculentum* Moench) protein possessing cholesterol-binding properties that reduce micelle cholesterol solubility and uptake by Caco-2 cells. *J Agric Food Chem* 2007;55 (15):6032-8.

**Table 1**

Amino acid composition of the sericin

<b>Amino acid</b>	<b>% w/w</b>
Serine	33.40
Aspartate	16.70
Glutamate	4.40
Glycine	13.50
Threonine	9.70
Lysine	3.30
Tyrosine	2.60
Arginine	3.10
Alanine	6.00
Valine	2.80
Histidine	1.30
Luecine	1.10
Isoluecine	0.70
Phenylalanine	0.50
Tryptophan	0.20
Proline	0.70
Cystine	0.20
Methionine	0.04



**Table 2**

Changes of plasma lipid levels and amount of food intake after 14 days of treatment

Groups	Changes from day 0 (mg/dl)				Food intake (g/day/100g BW)
	Total cholesterol	HDL-C	Non-HDL-C	Triglyceride	
High cholesterol	97.45±31.52	-0.38±1.49	97.83±31.01	56.05±40.70	6.56±1.34
High cholesterol+ Sericin 10 mg/kg/day	41.06±7.59**	1.80±3.87	39.25±4.26**	30.71±29.00	6.18±0.29
High cholesterol+ Sericin 100 mg/kg/day	38.61±12.45**	2.64±4.80	35.97±16.84**	72.79±43.02	8.43±1.86
High cholesterol+ Sericin 1000 mg/kg/day	49.05±13.02*	-2.70±9.54	51.75±9.84*	97.87±16.61	7.52±0.51

Values are expressed as mean ± SD (n = 5)

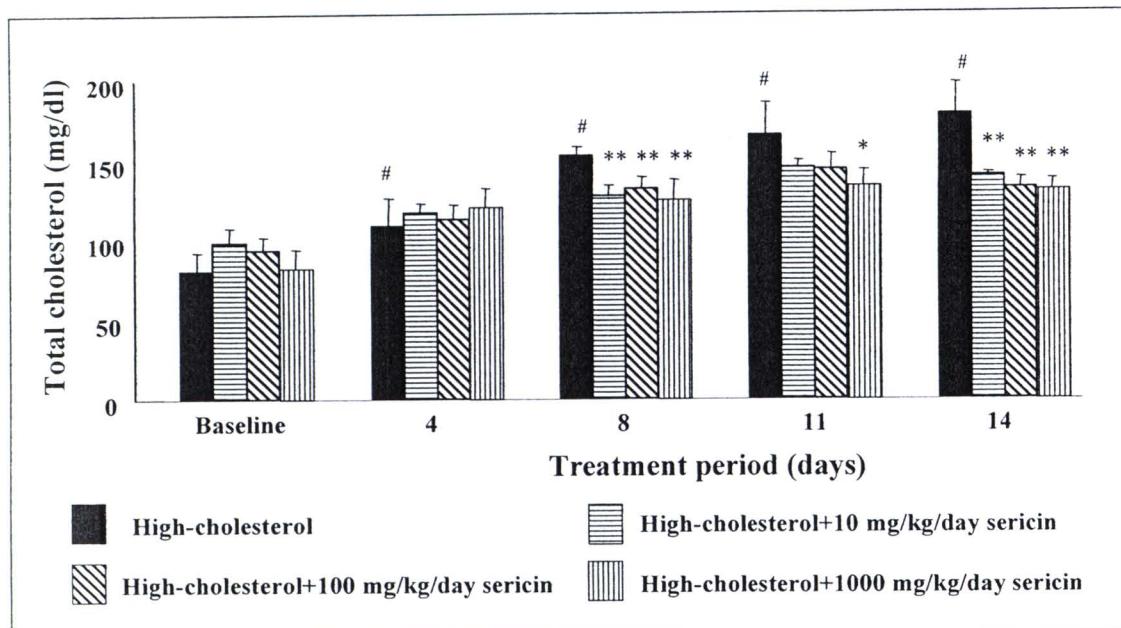
Significantly different from high cholesterol group (\* p ≤ 0.05, \*\* p ≤ 0.01)

**Table 3.**  
Inhibition of cholesterol micellar solubility of sericin

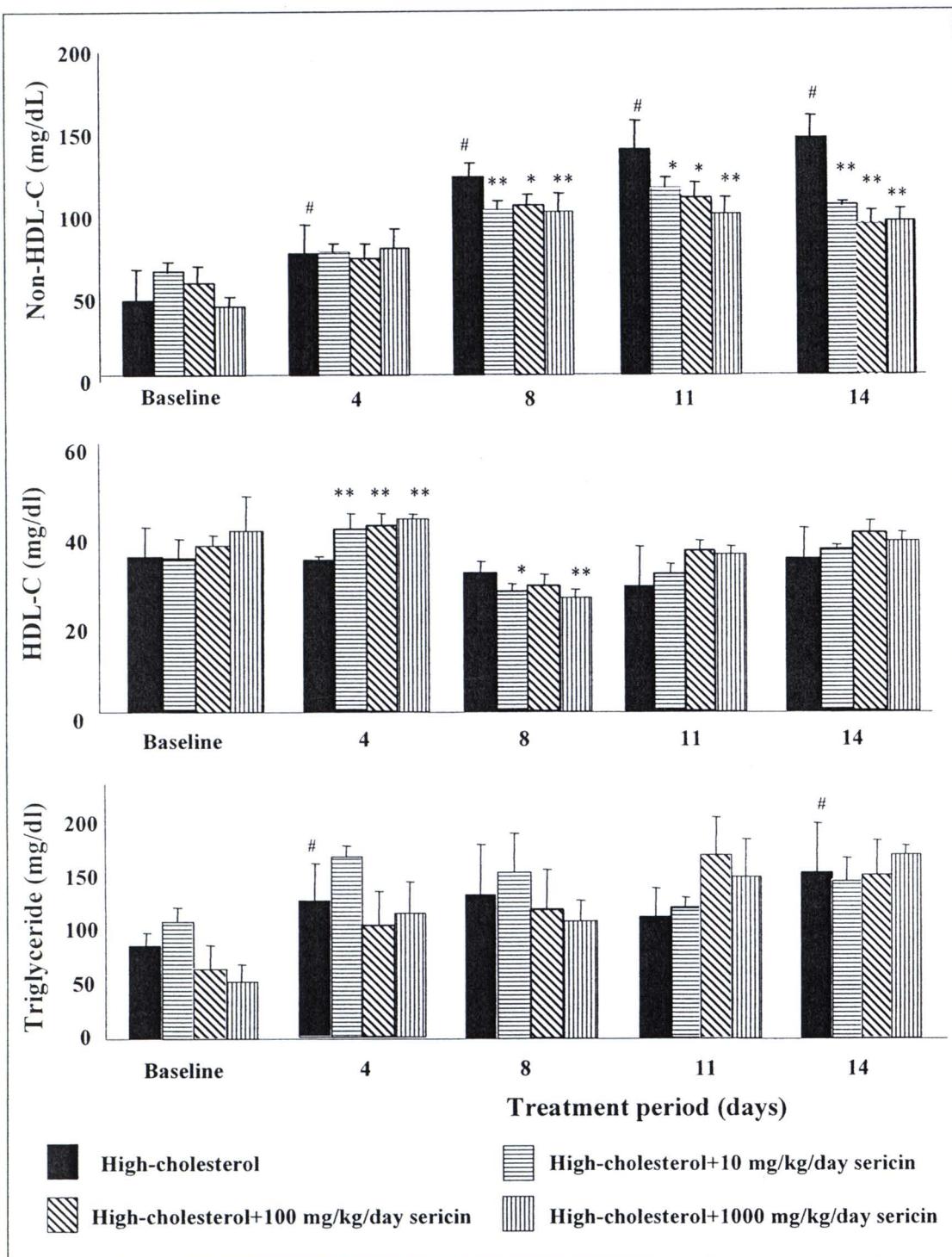
Protein concentration ( $\mu\text{g/ml}$ )	Before micelle preparation		After micelle preparation	
	Sericin	BSA	Sericin	BSA
Control	94.46 $\pm$ 3.34	94.46 $\pm$ 3.34	83.24 $\pm$ 12.02	83.24 $\pm$ 12.02
0	92.11 $\pm$ 7.80	92.11 $\pm$ 7.80	85.01 $\pm$ 8.86	85.01 $\pm$ 8.86
500	76.72 $\pm$ 7.98*	87.96 $\pm$ 4.37	77.84 $\pm$ 4.14	85.46 $\pm$ 5.39
1000	71.78 $\pm$ 9.76*	85.92 $\pm$ 1.62	66.67 $\pm$ 3.45	84.65 $\pm$ 7.52
2000	71.30 $\pm$ 3.39*	87.36 $\pm$ 6.14	64.52 $\pm$ 1.70	86.31 $\pm$ 6.20

Values are expressed as mean  $\pm$  SD ( $n = 3$ )

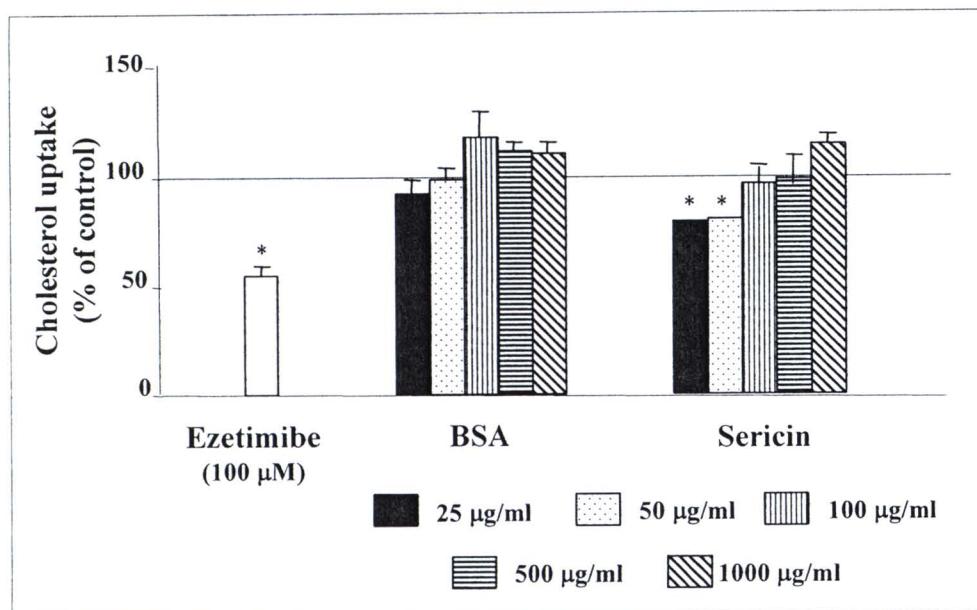
Significantly different from control (\*  $p \leq 0.05$ )



**Fig. 1.** Plasma total cholesterol in rats fed high-cholesterol diet. Rats were fed cholesterol (2% cholesterol of total daily diet) with and without sericin at 10, 100 and 1000 mg/kg for 14 days. The data represent mean  $\pm$  SD. (# p < 0.05 compared to baseline; \* p < 0.05, \*\* p < 0.01 compared to high-cholesterol, n = 5)



**Fig. 2.** Plasma HDL cholesterol (HDL-C), non-HDL cholesterol (non-HDL-C) and triglyceride in rats fed high-cholesterol diet. Rats were fed cholesterol (2% cholesterol of total daily diet) with and without sericin at 10, 100 and 1000 mg/kg for 14 days. The data represent mean  $\pm$  SD. (# p < 0.05 compared to baseline; \* p < 0.05, \*\* p < 0.01 compared to high-cholesterol, n = 5)



**Fig. 3.** Cholesterol uptake in differentiated Caco-2 cells. Differentiated Caco-2 cells were pretreated with various concentrations of either sericin or BSA (25-1000 µg/ml) for 1 h before incubating with [ $1\alpha,2\alpha(n)$ -<sup>3</sup>H]cholesterol-micelles for 3 h. Ezetimibe was used as positive control. The data represent mean  $\pm$  SEM from 5-6 experiments. \* indicates significant difference between treatment and control or PBS-treated groups with  $p < 0.05$ .

# บทที่ ๙

ภาคผนวก

## 1. การผลิตอาหารสัตว์ทดลองเสริมชีวิชิน

### 1.1. ข้อมูลผลการวิเคราะห์องค์ประกอบทางเคมี

#### 1.) ความชื้น (Moisture content)

ตารางที่ 43 ปริมาณความชื้นที่เป็นองค์ประกอบในตัวอย่างสูตรอาหารหนูเสริมชีวิชิน

ตัวอย่าง	Rep.	น้ำหนัก		น้ำหนักตัวอย่าง		น้ำหนักตัวอย่าง		ความชื้น		
		ถัวย	ตัวอย่างเริ่มต้น	และถัวเหลือง	เหลือง	(moisture content)				
						(%)	Ave.	SD		
COM	1	12.4725	2.0205	14.3536	1.8811	6.90	<b>6.74</b>	0.136		
	2	13.8974	2.0170	15.7796	1.8822	6.68				
	3	12.4820	2.0062	14.3548	1.8728	6.65				
CN5	1	14.0073	2.0090	15.9079	1.9006	5.40	<b>5.44</b>	0.106		
	2	12.7835	2.0103	14.6821	1.8986	5.56				
	3	13.8369	2.0236	15.7521	1.9152	5.36				
CN6	1	13.7775	2.0205	15.6549	1.8774	7.08	<b>7.13</b>	0.137		
	2	13.6533	2.0066	15.5191	1.8658	7.02				
	3	13.8920	2.0194	15.7644	1.8724	7.28				
CN7	1	12.4225	2.0138	14.3674	1.9449	3.42	<b>3.94</b>	0.875		
	2	14.4931	2.0052	16.4293	1.9362	3.44				
	3	14.7884	2.0196	16.7081	1.9197	4.95				
CC7	1	15.3824	2.0219	17.3220	1.9396	4.07	<b>4.20</b>	0.115		
	2	13.5662	2.0199	15.5002	1.9340	4.25				
	3	13.5102	2.0150	15.4389	1.9287	4.28				
CS7	1	13.3306	2.0188	15.2990	1.9684	2.50	<b>2.52</b>	0.068		
	2	13.6843	2.0181	15.6526	1.9683	2.47				
	3	13.2191	2.0254	15.1919	1.9728	2.60				
ST5	1	13.2949	2.0080	15.1904	1.8955	5.60	<b>5.61</b>	0.079		
	2	13.1869	2.0113	15.0838	1.8969	5.69				
	3	13.9811	2.0252	15.8943	1.9132	5.53				
ST6	1	11.8656	2.0276	13.7754	1.9098	5.81	<b>5.88</b>	0.068		
	2	12.5311	2.0185	14.4309	1.8998	5.88				
	3	11.5725	2.0219	13.4742	1.9017	5.94				
ST7	1	12.6640	2.0256	14.5779	1.9139	5.51	<b>5.56</b>	0.071		
	2	11.4922	2.0118	13.3905	1.8983	5.64				
	3	11.8648	2.0066	13.7606	1.8958	5.52				
SS7	1	12.3935	2.0233	14.3627	1.9692	2.67	<b>2.64</b>	0.041		
	2	11.8586	2.0221	13.8282	1.9696	2.60				
	3	12.6481	2.0200	14.6144	1.9663	2.66				
SA7	1	12.0472	2.0106	13.9768	1.9296	4.03	<b>4.06</b>	0.028		
	2	11.5541	2.0251	13.4965	1.9424	4.08				
	3	12.6988	2.0243	14.6410	1.9422	4.06				

ตารางที่ 44 ปริมาณน้ำอิสระและความชื้นที่เป็นองค์ประกอบในสูตรอาหารหนูเสริมชีริชินและสูตรที่มีปริมาณโภเดสเตรอรอลสูง

ตัวอย่าง	Rep.	น้ำหนัก		น้ำหนัก		น้ำหนัก		ความชื้น		อุณหภูมิ		น้ำอิสระ	
		ถัววัย	ตัวอย่าง	ตัวอย่างและเริ่มต้น	ถัววัยหลังอบ	หลังอบ		(moisture content)		ขณะวัด	(%)	Ave	SD
			(g)	(g)	(g)	(g)	(%)	Ave	SD	(°C)	(%)	Ave	SD
CC	1	11.5725	2.0698	13.5294	1.9569	5.45	<b>5.32</b>	0.120	30.1	0.428	<b>0.36</b>	0.058	
	2	13.3318	2.0658	15.2896	1.9578	5.23			30.4	0.314			
	3	13.5119	2.0333	15.4380	1.9261	5.27			30.8	0.350			
SA	1	12.4219	2.0728	14.4154	1.9935	3.83	<b>3.82</b>	0.070	29.6	0.230	<b>0.25</b>	0.024	
	2	13.7797	2.0040	15.7059	1.9262	3.88			30.0	0.243			
	3	13.9826	2.0328	15.9393	1.9567	3.74			30.5	0.277			
SB	1	13.1884	2.1590	15.1988	2.0104	6.88	<b>6.86</b>	0.024	30.9	0.499	<b>0.50</b>	0.005	
	2	12.1018	2.0399	14.0021	1.9003	6.84			31.0	0.496			
	3	14.0087	2.1141	15.9782	1.9695	6.84			31.0	0.505			
SC	1	13.6550	2.0443	15.5879	1.9329	5.45	<b>5.43</b>	0.025	31.1	0.351	<b>0.35</b>	0.003	
	2	11.6550	2.0822	-	-	-			31.2	0.345			
	3	12.6424	2.0338	14.5661	1.9237	5.41			31.3	0.347			
HC	1	13.2122	2.1082	15.2397	2.0275	3.83	<b>4.78</b>	1.831	29.8	0.265	<b>0.27</b>	0.020	
	2	13.2958	2.0707	15.2238	1.9280	6.89			30.2	0.289			
	3	12.6437	2.0707	14.6394	1.9957	3.62			30.9	0.250			
HB	1	12.7842	2.0712	14.7343	1.9501	5.85	<b>5.69</b>	0.168	31.4	0.401	<b>0.39</b>	0.008	
	2	12.5321	2.0940	14.5107	1.9786	5.51			31.5	0.386			
	3	11.5129	2.0674	13.4625	1.9496	5.70			31.6	0.392			



## 2.) ปริมาณถ้า (Ash content)

ตารางที่ 45 ปริมาณถ้าที่เป็นองค์ประกอบในตัวอย่างสูตรอาหารหนูเสริมชิริซิน

ตัวอย่าง	Rep.	Crucible	ตัวอย่างเสริม		ตัวอย่างและ		ตัวอย่าง		ปริมาณถ้าที่เป็นองค์ประกอบ			
					crucible		หลังเผา		หลังเผา		(ash content)	
			(g)	(g-wet)	(g-dry)	(g)	(g)	(%) <sub>wet</sub>	Ave.	SD	(%) <sub>dry</sub>	Ave.
COM	1	29.3183	2.1762	2.0294	29.4520	0.1337	6.14	<b>6.16</b>	0.015	6.59	<b>6.60</b>	0.017
	2	27.7448	2.0198	1.8836	27.8695	0.1247	6.17			6.62		
	3	23.4940	2.1763	2.0295	23.6279	0.1339	6.15			6.60		
CN5	1	35.7927	2.0685	1.9561	35.9246	0.1319	6.38	<b>6.36</b>	0.027	6.74	<b>6.72</b>	0.028
	2	28.6663	2.1066	1.9921	28.7996	0.1333	6.33			6.69		
	3	24.9811	2.1663	2.0485	25.1191	0.1380	6.37			6.74		
CN6	1	26.2434	2.2347	2.0755	26.3742	0.1308	5.85	<b>5.91</b>	0.071	6.30	<b>6.36</b>	0.076
	2	30.1829	2.2401	2.0805	30.3170	0.1341	5.99			6.45		
	3	23.4667	2.0776	1.9295	23.5888	0.1221	5.88			6.33		
CN7	1	36.4905	2.0365	1.9563	36.6192	0.1287	6.32	<b>6.32</b>	0.019	6.58	<b>6.58</b>	0.019
	2	25.9935	2.0495	1.9688	26.1233	0.1298	6.33			6.59		
	3	28.6180	2.1853	2.0993	28.7556	0.1376	6.30			6.55		
CC7	1	29.7542	2.0470	1.9610	29.8819	0.1277	6.24	<b>6.19</b>	0.050	6.51	<b>6.47</b>	0.053
	2	26.4047	2.0637	1.9770	26.5314	0.1267	6.14			6.41		
	3	26.8830	2.1818	2.0901	27.0184	0.1354	6.21			6.48		
CS7	1	29.7981	2.0184	1.9675	29.9525	0.1544	7.65	<b>7.64</b>	0.005	7.85	<b>7.84</b>	0.005
	2	32.2167	2.0982	2.0453	32.3771	0.1604	7.64			7.84		
	3	26.6211	2.0379	1.9865	26.7768	0.1557	7.64			7.84		
ST5	1	34.4002	2.1079	1.9897	34.5256	0.1254	5.95	<b>5.95</b>	0.009	6.30	<b>6.31</b>	0.009
	2	28.8864	2.1434	2.0232	29.0139	0.1275	5.95			6.30		
	3	29.6803	2.0139	1.9010	29.8004	0.1201	5.96			6.32		
ST6	1	36.7154	2.1850	2.0566	36.8514	0.1360	6.22	<b>6.20</b>	0.041	6.61	<b>6.58</b>	0.044
	2	28.5276	2.1042	1.9805	28.6570	0.1294	6.15			6.53		
	3	28.5858	2.1925	2.0636	28.7221	0.1363	6.22			6.60		
ST7	1	23.8227	2.0362	1.9230	23.9470	0.1243	6.10	<b>6.05</b>	0.062	6.46	<b>6.41</b>	0.065
	2	25.8902	2.1488	2.0293	26.0188	0.1286	5.98			6.34		
	3	29.8387	2.1319	2.0134	29.9681	0.1294	6.07			6.43		
SS7	1	26.3892	2.0144	1.9613	26.5357	0.1465	7.27	<b>7.31</b>	0.036	7.47	<b>7.50</b>	0.037
	2	28.0776	2.1215	2.0656	28.2334	0.1558	7.34			7.54		
	3	30.6504	2.1824	2.1249	30.8097	0.1593	7.30			7.50		
SA7	1	28.7596	2.0679	1.9840	28.8836	0.1240	6.00	<b>6.15</b>	0.231	6.25	<b>6.41</b>	0.241
	2	32.0228	2.0615	1.9779	32.1472	0.1244	6.03			6.29		
	3	23.0386	2.0157	1.9339	23.1679	0.1293	6.41			6.69		

ตารางที่ 46 ปริมาณถ้าที่เป็นองค์ประกอบในสูตรอาหารหนูเสริมชิริชินและสูตรที่มีปริมาณโคลเลสเตอรอลสูง

ตัวอย่าง	Rep.	Crucible	ตัวอย่างเริ่มต้น		ตัวอย่างและ		ตัวอย่าง		ปริมาณถ้าที่เป็นองค์ประกอบ					
					crucible หลังเผา		หลังเผา		(ash content)					
			(g)	(g-wet)	(g-dry)	(g)	(g)	(%) <sub>wet</sub>	Ave.	SD	(%) <sub>dry</sub>	Ave.	SD	
CC	1	19.8737	2.0563	1.9469	20.0092	0.1355	6.59	<b>6.57</b>	0.036	6.96	<b>6.94</b>	<b>0.038</b>		
	2	21.4807	2.0118	1.9048	31.6121	0.1314	6.53					6.90		
	3	20.6791	2.0875	1.9765	20.8168	0.1377	6.60					6.97		
SA	1	19.3323	2.0222	1.9147	19.4685	0.1269	6.28	<b>6.37</b>	0.152	6.63	<b>6.73</b>	<b>0.161</b>		
	2	21.5631	2.0533	1.9441	21.6933	0.1344	6.55					6.91		
	3	19.9278	2.0272	1.9194	20.0594	0.1275	6.29					6.64		
SB	1	22.1589	2.0555	1.9462	22.2845	0.1256	6.11	<b>6.08</b>	0.023	6.45	<b>6.43</b>	<b>0.025</b>		
	2	22.8482	2.0408	1.9323	22.9721	0.1239	6.07					6.41		
	3	21.1522	2.0943	1.9829	21.2793	0.1271	6.07					6.41		
SC	1	20.1308	2.0774	1.9669	20.2580	0.1272	6.12	<b>6.08</b>	0.043	6.47	<b>6.42</b>	<b>0.046</b>		
	2	20.3058	2.0286	1.9207	20.4283	0.1225	6.04					6.38		
	3	21.9837	2.0529	1.9437	22.1082	0.1245	6.06					6.41		
HC	1	20.3969	2.0916	1.9804	20.5186	0.1362	6.51	<b>6.47</b>	0.040	6.88	<b>6.83</b>	<b>0.042</b>		
	2	20.4659	2.0185	1.9112	20.5912	0.1302	6.45					6.81		
	3	22.4845	2.0445	1.9358	22.6118	0.1316	6.44					6.80		
HB	1	21.3829	2.0051	1.8985	21.5098	0.1217	6.07	<b>6.07</b>	0.020	6.41	<b>6.41</b>	<b>0.021</b>		
	2	20.4134	2.0696	1.9595	20.5478	0.1253	6.05					6.39		
	3	20.7008	2.0890	1.9779	20.8283	0.1273	6.09					6.44		

### 3.) ปริมาณทราย (Sand content)

ตารางที่ 47 ปริมาณทรายที่เป็นองค์ประกอบในตัวอย่างสูตรอาหารหนูเสริมชีริชิน

ตัวอย่าง	Rep.	Crucible	ตัวอย่างเริ่มต้น			ตัวอย่างและ		ตัวอย่าง		ปริมาณทรายที่เป็นองค์ประกอบ			
						crucible หลังเผา		หลังเผา		(sand content)			
			(g)	(g- <sub>wet</sub> )	(g- <sub>dry</sub> )	(g)	(g)	(%) <sub>wet</sub>	Ave.	SD	(%) <sub>dry</sub>	Ave.	SD
COM	1	29.3183	2.1762	2.0294	29.3273	0.0090	0.41	0.29	0.109	0.44	0.31	0.117	
	2	27.7448	2.0198	1.8836	27.7494	0.0049	0.24			0.26			
	3	23.4940	2.1763	2.0295	23.4986	0.0046	0.21			0.23			
CN5	1	35.7927	2.0685	1.9561	35.8031	0.0104	0.50	0.53	0.048	0.53	0.56	0.051	
	2	28.6663	2.1066	1.9921	28.6767	0.0104	0.49			0.52			
	3	24.9811	2.1663	2.0485	24.9937	0.0126	0.58			0.62			
CN6	1	26.2434	2.2347	2.0755	26.2508	0.0074	0.33	0.35	0.012	0.36	0.37	0.013	
	2	30.1829	2.2401	2.0805	30.1908	0.0079	0.35			0.38			
	3	23.4667	2.0776	1.9295	23.4740	0.0073	0.35			0.38			
CN7	1	36.4905	2.0365	1.9563	36.4996	0.0091	0.45	0.42	0.033	0.47	0.43	0.035	
	2	25.9935	2.0495	1.9688	26.0013	0.0078	0.38			0.40			
	3	28.6180	2.1853	2.0993	28.6272	0.0092	0.42			0.44			
CC7	1	29.7542	2.0470	1.9610	29.7615	0.0073	0.36	0.34	0.040	0.37	0.35	0.042	
	2	26.4047	2.0637	1.9770	26.4123	0.0076	0.37			0.38			
	3	26.8830	2.1818	2.0901	26.8894	0.0064	0.29			0.31			
CS7	1	29.7981	2.0184	1.9675	29.8072	0.0091	0.45	0.45	0.043	0.46	0.46	0.044	
	2	32.2167	2.0982	2.0453	32.2253	0.0086	0.41			0.42			
	3	26.6211	2.0379	1.9865	26.6312	0.0101	0.50			0.51			
ST5	1	34.4002	2.1079	1.9897	34.4106	0.0104	0.49	0.48	0.032	0.52	0.51	0.033	
	2	28.8864	2.1434	2.0232	28.8971	0.0107	0.50			0.53			
	3	29.6803	2.0139	1.9010	29.6892	0.0089	0.44			0.47			
ST6	1	36.7154	2.1850	2.0566	36.7253	0.0099	0.45	0.40	0.045	0.48	0.43	0.048	
	2	28.5276	2.1042	1.9805	28.5355	0.0079	0.38			0.40			
	3	23.8227	2.1925	2.0636	28.5940	0.0082	0.37			0.40			
ST7	1	23.8227	2.0362	1.9230	23.8303	0.0076	0.37	0.34	0.030	0.40	0.36	0.031	
	2	25.8902	2.1488	2.0293	25.8972	0.0070	0.33			0.34			
	3	29.8387	2.1319	2.0134	29.8455	0.0068	0.32			0.34			
SS7	1	26.3892	2.0144	1.9613	26.3978	0.0086	0.43	0.39	0.043	0.44	0.40	0.044	
	2	28.0776	2.1215	2.0656	28.0849	0.0073	0.34			0.35			
	3	30.6504	2.1824	2.1249	30.6592	0.0088	0.40			0.41			
SA7	1	28.7596	2.0679	1.9840	28.7689	0.0093	0.45	0.30	0.128	0.47	0.31	0.133	
	2	32.0228	2.0615	1.9779	32.0274	0.0046	0.22			0.23			
	3	23.0386	2.0157	1.9339	23.0433	0.0047	0.23			0.24			

#### 4.) ปริมาณโปรตีนทั้งหมด (Total nitrogen and protein content)

ตารางที่ 48 ปริมาณ โปรตีนทั้งหมดที่เป็นองค์ประกอบในตัวอย่างสูตรอาหารหนูเสริมชิริซิน

ตัวอย่าง	Rep.	ตัวอย่างเริ่มต้น	0.1 N HCl	N HCl	ปริมาณโปรตีนที่เป็นองค์ประกอบ								
					(as total nitrogen content)								
					(g <sub>wet</sub> )	(g <sub>wet</sub> )	(ml)	Actual	(%) <sub>wet</sub>	Ave.	SD	(%) <sub>dry</sub>	Ave.
COM	1	2.0211	1.8848	53.30	0.1061	23.96	24.62	1.198	25.69	26.40	1.285		
	2	2.0222	1.8858	57.80	0.1061	26.01			27.89				
	3	2.0288	1.8920	53.40	0.1061	23.91			25.64				
CN5	1	2.0218	1.9119	57.25	0.1061	25.76	26.36	0.868	27.24	27.88	0.918		
	2	2.0245	1.9144	58.05	0.1056	25.97			27.47				
	3	2.0013	1.8925	60.40	0.1056	27.36			28.93				
CN6	1	2.0269	1.8825	60.55	0.1061	27.21	27.70	0.861	29.29	29.83	0.927		
	2	2.0120	1.8686	60.10	0.1061	27.20			29.29				
	3	2.0144	1.8708	63.70	0.1506	28.70			30.90				
CN7	1	2.0279	1.9481	55.25	0.1061	24.77	25.49	0.727	25.78	26.54	0.757		
	2	2.0283	1.9485	58.70	0.1056	26.22			27.30				
	3	2.0138	1.9345	56.70	0.1056	25.49			26.54				
CC7	1	2.0222	1.9372	60.80	0.1061	27.39	25.36	1.818	28.59	26.47	1.900		
	2	2.0131	1.9285	54.95	0.1061	24.81			25.90				
	3	2.0187	1.9339	53.30	0.1056	23.87			24.92				
CS7	1	2.0064	1.9558	62.45	0.1061	28.36	29.36	1.000	29.10	30.12	1.026		
	2	2.0148	1.9640	67.35	0.1056	30.36			31.15				
	3	2.0095	1.9589	65.00	0.1056	29.36			30.12				
ST5	1	2.0173	1.9042	56.50	0.1061	25.47	28.31	2.572	26.99	29.99	2.720		
	2	2.0125	1.8997	67.25	0.1061	30.49			32.30				
	3	2.0029	1.8906	63.90	0.1056	28.95			30.67				
ST6	1	2.0167	1.8981	67.90	0.1061	30.59	28.61	2.613	32.50	30.39	2.776		
	2	2.0100	1.8918	65.20	0.1061	29.58			31.43				
	3	2.0035	1.8857	56.75	0.1056	25.65			27.25				
ST7	1	2.0060	1.8945	65.50	0.1061	29.78	29.81	0.176	31.53	31.56	0.186		
	2	2.0307	1.9178	66.00	0.1061	29.65			31.39				
	3	2.0195	1.9072	66.70	0.1056	30.00			31.76				
SS7	1	2.0027	1.9499	65.80	0.1056	29.83	29.90	0.097	30.64	30.71	0.100		
	2	2.0278	1.9744	66.70	0.1061	30.01			30.82				
	3	2.0098	1.9561	66.05	0.1056	29.85			30.66				
SA7	1	2.0277	1.9455	69.40	0.1061	31.25	30.34	0.979	32.57	31.62	1.021		
	2	2.0279	1.9456	67.70	0.1061	30.47			31.76				
	3	2.0153	1.9336	65.05	0.1056	29.30			30.54				

ตารางที่ 49 ปริมาณโปรตีนที่เป็นองค์ประกอบในสูตรอาหารหมูเสริมชีวิตชิ้นและสูตรที่มีปริมาณโภเดสเทอโรลดูง

ตัวอย่าง	Rep.	ตัวอย่างเริ่มต้น	0.1N HCl	ปริมาณโปรตีนที่เป็นองค์ประกอบ						
				(as total nitrogen content)						
				(g - <sub>wet</sub> )	(g - <sub>dry</sub> )	(ml)	(% <sub>wet</sub> )	Ave.	SD	
CC	1	0.5392	0.5105	17.05	25.88	<b>26.14</b>	0.289	27.34	<b>27.60</b>	0.305
	2	0.5118	0.4846	16.35	26.07			27.54		
	3	0.5111	0.4839	16.55	26.45			27.94		
SA	1	0.5217	0.5018	16.80	26.33	<b>27.19</b>	1.216	27.38	<b>28.27</b>	1.264
	2	-	-	-	-			-		
	3	0.5521	0.5310	18.80	28.05			29.17		
SB	1	0.5168	0.4814	-	-	<b>29.70</b>	0.278	-	<b>31.89</b>	0.299
	2	0.5709	0.5318	20.35	29.50			31.68		
	3	0.5546	0.5166	20.05	29.90			32.10		
SC	1	0.5517	0.5217	20.30	30.45	<b>29.14</b>	1.859	32.20	<b>30.81</b>	1.965
	2	0.5252	0.4967	17.80	27.82			29.42		
	3	-	-	-	-			-		
HC	1	0.5316	0.5062	18.30	28.31	<b>28.05</b>	0.704	29.73	<b>29.46</b>	0.739
	2	0.5395	0.5137	18.73	28.59			30.03		
	3	0.5104	0.4860	17.00	27.26			28.63		
HB	1	0.5418	0.5110	18.30	27.78	<b>26.95</b>	1.164	29.45	<b>28.58</b>	1.234
	2	0.5221	0.4924	-	-			-		
	3	0.5324	0.5021	17.00	26.13			27.51		

### 5.) ปริมาณไขมันทั้งหมด (Fat content)

ตารางที่ 50 ปริมาณไขมันที่เป็นองค์ประกอบในตัวอย่างสูตรอาหารหนูเสริมชีวิตนิ

ตัวอย่าง	Rep.	กระดาษ	ตัวอย่างเริ่มต้น	ตัวอย่างแลบ	ไขมัน		ปริมาณไขมันที่เป็นองค์ประกอบ					
					กรอง		กระดาษกรอง		(fat content)			
					(g)	(g <sub>wet</sub> )	(g <sub>wet</sub> )	หลังถักดัด (g)	(g)	(% <sub>wet</sub> )	Ave.	SD
COM	1	2.2192	3.0270	2.8229	5.1092	0.1370	4.53	4.78	0.323	4.85	<b>5.12</b>	0.346
	2	2.2073	3.0702	2.8631	5.1343	0.1432	4.66			5.00		
	3	2.0170	3.0281	2.8239	4.8894	0.1557	5.14			5.51		
CN5	1	2.2139	3.0225	2.8582	5.1206	0.1157	3.83	<b>4.28</b>	0.449	4.05	<b>4.53</b>	0.475
	2	2.2066	3.0241	2.8597	5.1010	0.1297	4.29			4.53		
	3	2.0125	3.0404	2.8751	4.9091	0.1438	4.73			5.00		
CN6	1	2.1663	3.0067	2.7924	5.0613	0.1117	3.72	<b>4.46</b>	0.665	4.00	<b>4.80</b>	0.716
	2	2.1680	3.0147	2.7999	5.0422	0.1405	4.66			5.02		
	3	1.9869	3.0351	2.8188	4.8702	0.1518	5.00			5.38		
CN7	1	2.1611	3.0184	2.8996	5.0311	0.1484	4.92	<b>5.12</b>	0.173	5.12	<b>5.33</b>	0.180
	2	2.1452	3.0195	2.9006	5.0069	0.1578	5.22			5.44		
	3	2.1704	3.0212	2.9023	5.0342	0.1574	5.21			5.42		
CC7	1	2.1764	3.0297	2.9024	5.0711	0.1350	4.46	<b>4.54</b>	0.379	4.65	<b>4.74</b>	0.396
	2	2.1944	3.0089	2.8825	5.0767	0.1266	4.21			4.39		
	3	1.9574	3.0429	2.9150	4.4896	0.1507	4.95			5.17		
CS7	1	2.2384	3.0250	2.9488	5.1784	0.0850	2.81	<b>2.72</b>	0.120	2.88	<b>2.79</b>	0.123
	2	2.1764	3.0174	2.9413	5.1159	0.0779	2.58			2.65		
	3	1.6965	3.0515	2.9746	4.6638	0.0842	2.76			2.83		
ST5	1	2.1631	3.0047	2.8362	5.0665	0.1013	3.37	<b>3.61</b>	0.364	3.57	<b>3.82</b>	0.386
	2	2.2032	3.0180	2.8488	5.1180	0.1032	3.42			3.62		
	3	1.7049	3.0468	2.8760	4.6290	0.1227	4.03			4.26		
ST6	1	2.2600	3.0039	2.8273	5.1342	0.1297	4.32	<b>4.51</b>	0.197	4.59	<b>4.97</b>	0.210
	2	2.2748	3.0082	2.8314	5.1480	0.1350	4.49			4.77		
	3	1.7183	3.0477	2.8685	4.6224	0.1436	4.71			5.01		
ST7	1	2.1647	3.0204	2.8525	5.0782	0.1069	3.54	<b>3.69</b>	0.131	3.75	<b>3.91</b>	0.139
	2	2.2074	3.0045	2.8375	5.0982	0.1137	3.78			4.01		
	3	1.7066	3.0323	2.8637	4.6252	0.1137	3.75			3.97		
SS7	1	2.2090	3.0095	2.9302	5.1333	0.0852	2.83	<b>2.98</b>	0.275	2.91	<b>3.06</b>	0.283
	2	2.1755	3.0168	2.9373	5.1076	0.0847	2.81			2.88		
	3	1.6958	3.0253	2.9456	4.6214	0.0997	3.30			3.38		
SA7	1	2.1574	3.0135	2.8913	5.0993	0.0716	2.38	<b>2.63</b>	0.216	2.48	<b>2.74</b>	0.225
	2	1.7060	3.0099	2.8878	4.6340	0.0819	2.72			2.84		
	3	1.6884	3.0358	2.9127	4.6399	0.0843	2.78			2.89		

ตารางที่ 51 ปริมาณไขมันที่เป็นองค์ประกอบในสูตรอาหารหนูเสริมชีวิตชิ้นและสูตรที่มีปริมาณโคลเลสเตอรอลสูง

ตัวอย่าง	Rep.	กระดาษ	ตัวอย่างเริ่มต้น	ตัวอย่างและ ไขมัน	ปริมาณไขมันที่เป็นองค์ประกอบ							
					กรอง			กระดาษกรอง			(fat content)	
					(g)	(g-wet)	(g-wet)	หลังสกัด (g)	(g)	(%wet)	Ave.	SD
CC	1	2.2037	3.0639	3.0639	2.9010	5.1350	4.33	4.30	0.124	4.57	4.54	0.130
	2	2.1929	3.0060	3.0606	2.8978	5.1188	4.40			4.65		
	3	2.2511	3.0064	3.1369	2.9701	5.2575	4.16			4.39		
SA	1	2.2454	3.0062	3.0062	2.8914	5.1439	3.58	3.45	0.084	3.72	3.68	0.087
	2	2.2323	3.0081	3.0081	2.8933	5.1368	3.44			3.58		
	3	2.2538	3.0704	3.0704	2.9532	5.2138	3.60			3.74		
SB	1	2.2310	3.0150	3.0152	2.8085	5.0962	4.97	4.78	0.281	5.34	5.13	0.301
	2	2.1728	3.0364	3.0364	2.8282	5.0702	4.58			4.91		
	3	2.1949	3.0900	3.0062	2.8001	-	-			-		
SC	1	2.1870	3.0792	3.0792	2.9120	5.1375	4.18	4.54	0.503	4.42	4.80	0.532
	2	2.8590	3.0590	2.3859	2.2563	-	-			-		
	3	2.7630	3.0032	2.4165	2.2853	5.0613	4.89			5.17		
HC	1	2.2257	3.0215	3.0215	2.8771	5.0956	5.02	5.15	0.253	5.27	5.41	0.266
	2	2.1884	3.0889	3.0889	2.9412	5.1230	5.00			5.25		
	3	2.1710	3.0621	3.0621	2.9157	5.0664	5.44			5.72		
HB	1	2.2016	3.0881	3.0881	2.9125	5.1180	5.56	5.43	0.361	5.90	5.75	0.383
	2	2.1911	3.0802	3.0502	2.8768	5.0674	5.70			6.04		
	3	2.1689	3.0571	3.0571	2.8833	5.0726	5.02			5.32		

### 6.) ปริมาณเส้นใยหิน (Crude fiber)

ตารางที่ 52 | ปริมาณเส้นใยหินที่ใช้ทดสอบในตัวอย่างที่ได้รับจากอบไนต์ของยาสูตรอาหารหนูบลูเบิร์ชัน

ตัวอย่าง	Rep.	Crucible	กรด tam	ตัวอย่างเริ่มต้น	กาก	กาก หลังเผา	ถ้า หลังเผา	ปริมาณเส้นใยหินที่ประเมินจาก粗纤维 (crude fiber)				
								(g)	(g-wet)	(g-wet)	คงค่ากราฟเรซิวัล	
COM	1	24.8197	1.1880	3.0270	2.8229	26.0668	24.8253	0.0591	0.0056	1.77	1.66	0.347
	2	28.4581	1.1702	3.0702	2.8631	29.6765	28.4673	0.0482	0.0092	1.27	1.27	0.372
CN5	3	26.4052	1.1468	3.0281	2.8239	27.6161	26.4106	0.0641	0.0054	1.94	2.32	0.670
	1	26.1081	1.1501	3.0225	2.8582	27.3024	25.1353	0.0442	0.0070	2.79	2.32	0.709
CN6	2	23.9395	1.2001	3.0241	2.8597	25.2310	23.9465	0.0914	0.0070	2.95	2.95	
	3	26.2449	1.1888	3.0404	2.8751	27.4977	26.2529	0.0640	0.0080	1.84	1.91	0.391
CN7	1	29.3192	1.1835	3.0067	2.7924	30.5657	29.3273	0.0630	0.0081	1.82	1.91	0.421
	2	28.7598	1.1723	3.0147	2.7999	29.9876	28.7680	0.0555	0.0082	1.57	1.69	
CC7	3	29.7539	1.1687	3.0351	2.8188	31.0009	29.7613	0.0783	0.0074	2.34	2.34	2.52
	1	28.8887	1.1967	3.0184	2.8996	30.1415	28.9009	0.0561	0.0122	1.45	2.08	0.668
CS7	2	27.7293	1.1865	3.0195	2.9006	28.9887	27.7407	0.0729	0.0114	2.04	2.08	1.51
	3	31.5155	1.1566	3.0212	2.9023	32.7631	31.5238	0.0910	0.0083	2.74	2.74	2.85
ST5	1	23.8237	1.1692	3.0297	2.9024	25.0036	23.8507	0.0107	0.0041	0.94	0.99	0.084
	2	32.2176	1.1630	3.0089	2.8825	33.4128	32.2217	0.0322	0.0044	1.05	0.99	0.98
ST6	3	29.6811	1.1787	3.0429	2.9150	30.8963	29.6855	0.0365	0.0044	1.10	1.46	0.399
	1	28.3172	1.1630	3.0250	2.9488	29.5397	28.3240	0.0595	0.0068	1.74	1.74	1.50
ST7	2			3.0174	2.9413							0.410
	3	28.5280	1.1754	3.0515	2.9746	29.7500	28.5391	0.0466	0.0111	1.18	1.64	0.648
SS7	1	29.7994	1.1907	3.0047	2.8362	31.0651	29.8140	0.0750	0.0146	2.01	1.90	2.13
	2	28.6186	1.1922	3.0180	2.8488	29.8805	28.6288	0.0697	0.0102	1.97	1.97	2.09
ST8	3	25.8908	1.1469	3.0468	2.8760	27.0751	25.8998	0.0374	0.0090	0.93	0.93	0.99
	1	30.1825	1.1804	3.0039	2.8273	31.4593	30.2220	0.0964	0.0395	1.90	1.75	2.01
ST9	2	27.2032	1.1568	3.0082	2.8314	28.0480	27.2120	0.0614	0.0051	1.87	1.90	0.332
	3	28.5865	1.1708	3.0047	2.8685	29.8145	28.5950	0.0572	0.0085	1.60	1.92	1.70
ST10	1	27.6020	1.1408	3.0204	2.8525	28.8104	27.6116	0.0676	0.0096	1.92	1.90	2.03
	2	32.0383	1.1403	3.0045	2.8375	33.2400	32.0434	0.0614	0.0051	1.87	1.75	2.01
SS8	3	29.0874	1.1781	3.0095	2.9302	30.3346	29.1015	0.0691	0.0141	1.83	1.47	0.510
	2	25.9949	1.1918	3.0168	2.9373	27.2296	26.0044	0.0429	0.0095	1.11	1.11	1.14
SA7	1	24.9841	1.1515	3.0135	2.8913	26.1974	24.9954	0.0618	0.0113	1.68	1.58	0.256
	2	25.6836	1.1725	3.0099	2.8878	26.1988	25.6930	0.0627	0.0094	1.77	1.75	0.267
	3	28.7879	1.1726	3.0358	2.9127	30.0068	28.7951	0.0463	0.0072	1.29	1.29	1.34

### 7.) ปริมาณสตาร์ช (Starch content)

ตารางที่ 53 | ริบบินสตาร์ชที่เพื่อนองค์ประกอบในตัวอย่างสูตรอาหารห่านสตาร์ชรีซิป

ตัวอย่าง	Rep.	Absorbance (620m)						Carbohydrate conc. (ug/ml)						Total carbohydrate (g)						Total carbohydrate (%)					
		1	2	3	Ave	SD	1	2	Ave	SD	1	2	3	Ave	SD	1	2	3	Ave	Ave	SD				
COM 1	1	0.156	0.156	0.158	0.157	0.147	0.012	297.94	301.76	299.21	280.04	22.98	0.745	0.745	0.748	0.700	0.057	37.24	37.72	37.40	35.01	2.87			
	2	0.150	0.153	0.149	0.151			286.48	292.21	284.57	287.75		0.716	0.731	0.711	0.719		35.81	36.53	35.57	35.97				
	3	0.157	0.150	0.157	0.155			299.85	286.48	299.85	295.39		0.750	0.716	0.750	0.738		37.48	35.81	37.48	36.92				
	2	1	0.130	0.137	0.139	0.135		248.28	261.65	265.47	258.47		0.621	0.654	0.664	0.646		31.04	32.71	33.18	32.31				
	2	0.134	0.133	0.129	0.132			255.92	254.01	246.37	252.10		0.640	0.635	0.616	0.630		31.99	31.75	30.80	31.51				
	3	0.125	0.125	0.125	0.125			238.73	238.73	238.73	238.73		0.597	0.597	0.597	0.597		29.84	29.84	29.84	29.84				
CNS 1	3	1	0.156	0.158	0.156	0.157		297.94	301.76	297.94	299.21		0.745	0.754	0.745	0.748		37.24	37.72	37.72	37.40				
	2	0.154	0.150	0.154	0.153			294.12	285.48	294.12	291.57		0.735	0.716	0.735	0.729		36.76	35.81	35.81	36.45				
	3	0.156	0.156	0.156	0.156			297.94	297.94	297.94	297.94		0.745	0.745	0.745	0.745		37.24	37.24	37.24	37.42				
	1	0.192	0.193	0.194	0.193	0.196	0.004	366.69	368.60	370.51	368.60	374.61	7.63	0.917	0.922	0.926	0.922	0.937	0.019	45.84	46.08	46.31	46.83	0.95	
	2	0.192	0.197	0.191	0.193			366.69	376.24	364.78	369.24		0.917	0.941	0.912	0.923		45.84	47.03	45.60	46.15				
	3	0.196	0.193	0.197	0.195			374.33	368.60	376.24	373.06		0.936	0.922	0.941	0.933		46.79	46.08	47.03	46.63				
CN5	2	1	0.202	0.203	0.202	0.202		385.79	385.79	387.70	386.43		0.964	0.964	0.969	0.966		48.22	48.22	48.46	48.30				
	2	0.198	0.196	0.196	0.197			378.15	374.33	374.33	375.60		0.945	0.936	0.936	0.939		47.27	46.79	46.79	46.95				
	3	0.202	0.202	0.202	0.202			385.79	385.79	385.79	385.79		0.964	0.964	0.964	0.964		48.22	48.22	48.22	48.22				
	3	1	0.197	0.197	0.197	0.197		376.24	376.24	376.24	376.24		0.941	0.941	0.941	0.941		47.03	47.03	47.03	47.03				
	2	0.191	0.190	0.190	0.190			364.78	362.87	362.87	363.51		0.912	0.907	0.907	0.909		45.60	45.36	45.36	45.44				
	3	0.196	0.195	0.195	0.195			374.33	372.42	372.42	373.06		0.936	0.931	0.931	0.933		46.79	46.55	46.55	46.63				
CN6 1	1	0.177	0.176	0.176	0.176	0.223	0.029	338.04	336.13	336.13	336.77	426.46	54.68	0.845	0.840	0.840	0.842	1.066	0.137	42.26	42.02	42.02	42.10	53.31	6.84
	2	0.175	0.176	0.178	0.176			334.22	336.13	339.95	336.77		0.836	0.840	0.850	0.842		41.78	42.02	42.49	42.10				
	3	0.207	0.243	0.245	0.232			395.34	464.09	467.91	442.45		0.988	1.160	1.170	1.106		49.42	58.01	58.49	55.31				
	2	1	0.225	0.224	0.223	0.224		429.72	427.81	425.90	427.81		1.074	1.070	1.065	1.070		53.71	53.48	53.24	53.48				
	2	0.225	0.224	0.224	0.224			429.72	427.81	425.90	427.81		1.074	1.070	1.065	1.070		53.71	53.48	53.24	53.48				
	3	0.223	0.227	0.223	0.224			425.90	433.54	425.90	428.44		1.065	1.084	1.065	1.071		53.24	54.19	53.24	53.56				
CN7 1	3	1	0.250	0.252	0.252	0.251		477.46	481.28	481.28	480.01		1.194	1.203	1.203	1.200		59.68	60.16	60.16	60.00				
	2	0.249	0.248	0.252	0.250			475.55	473.64	481.28	476.83		1.189	1.184	1.203	1.192		59.44	59.21	60.16	59.60				
	3	0.251	0.252	0.253	0.252			479.37	481.28	483.19	481.28		1.198	1.203	1.208	1.203		59.92	60.16	60.40	60.16				
	1	0.238	0.231	0.232	0.234	0.237	0.004	454.55	441.18	441.18	446.27	452.85	7.25	1.136	1.103	1.108	1.116	1.132	0.018	56.82	55.15	55.39	55.78	56.61	0.91
	2	0.236	0.236	0.234	0.235			450.73	450.73	450.73	449.45		1.127	1.127	1.117	1.124		56.34	56.34	55.86	56.18				
	3	0.231	0.234	0.232	0.232			441.18	446.91	443.72	440.72		1.103	1.117	1.108	1.109		55.15	55.86	55.39	55.47				
CN7 2	1	0.236	0.237	0.235	0.236			450.73	452.64	452.64	450.73		1.127	1.132	1.122	1.127		56.34	56.58	56.10	56.34				
	2	0.237	0.239	0.240	0.239			452.64	456.46	456.46	455.82		1.132	1.141	1.146	1.140		56.58	57.06	57.30	56.98				
	3	0.236	0.236	0.236	0.236			450.73	450.73	450.73	450.73		1.127	1.127	1.127	1.127		56.34	56.34	56.34	56.34				
	3	1	0.240	0.239	0.239	0.239		458.37	456.46	456.46	457.09		1.146	1.141	1.141	1.143		57.30	57.06	57.06	57.14				
	2	0.241	0.237	0.236	0.238			460.28	452.64	452.64	454.55		1.151	1.132	1.127	1.136		57.53	56.58	56.34	56.82				
	3	0.245	0.244	0.245	0.245			467.91	466.00	467.91	467.28		1.170	1.165	1.170	1.168		58.25	58.49	58.49	58.41				

ตารางที่ 53 ปริมาณสารตัวชี้วัดในคราบไข่ต้มของอาหารที่มีส่วนประกอบเป็นไข่ต้มอย่างเดียว (หน้า)

ห้อง Rep	Absorbance (620nm)						Carbohydrate conc. (μm/ml)						Total carbohydrate (g)						
	1	2	3	Ave	Ave	SD	1	2	3	Ave	Ave	SD	1	2	3	Ave	Ave	SD	
CC7	1	1	0.242	0.243	0.243	0.243	0.238	0.009	462.18	464.09	463.46	453.84	17.61	1.155	1.160	1.159	1.135	0.044	
	2	0.245	0.245	0.241	0.244	0.244	467.91	467.91	460.82	465.37	465.37	465.37	1.170	1.170	1.151	1.163	1.170	1.171	
	3	0.247	0.243	0.246	0.245	0.245	471.73	464.09	469.82	468.55	468.55	468.55	1.179	1.160	1.175	1.171	1.065	1.070	
	2	1	0.223	0.224	0.224	0.224	425.90	427.81	427.17	427.81	427.81	427.81	1.065	1.070	1.070	1.068	1.065	1.068	
	2	0.227	0.223	0.230	0.227	0.227	433.54	425.90	439.27	432.90	432.90	432.90	1.084	1.065	1.980	1.082	1.084	1.082	
	3	0.227	0.224	0.224	0.225	0.225	433.54	427.81	427.81	429.72	429.72	429.72	1.084	1.070	1.070	1.074	1.165	1.165	
CS7	3	1	0.244	0.246	0.244	0.245	466.00	469.82	466.00	467.28	467.28	467.28	1.165	1.175	1.165	1.168	1.160	1.160	
	2	0.243	0.243	0.241	0.242	0.242	464.09	464.09	460.28	462.82	462.82	462.82	1.160	1.160	1.151	1.157	1.165	1.165	
	3	0.244	0.246	0.244	0.245	0.245	466.00	469.82	466.00	467.28	467.28	467.28	1.165	1.175	1.165	1.168	1.165	1.165	
	1	1	0.258	0.257	0.255	0.257	0.254	0.006	492.74	490.83	487.01	490.20	485.24	12.40	1.232	1.227	1.218	1.225	
	2	0.265	0.266	0.267	0.266	0.266	506.11	508.02	509.93	508.02	508.02	508.02	1.265	1.270	1.275	1.270	1.237	1.237	
	3	0.259	0.259	0.253	0.257	0.257	494.56	494.65	483.19	490.83	490.83	490.83	1.237	1.237	1.208	1.227	1.198	1.198	
ST5	2	1	0.251	0.266	0.267	0.261	479.37	475.55	483.19	479.37	479.37	479.37	1.198	1.189	1.208	1.198	1.198	1.198	
	2	0.251	0.250	0.254	0.252	0.252	479.37	477.46	485.10	480.65	480.65	480.65	1.198	1.194	1.213	1.202	1.213	1.213	
	3	0.254	0.245	0.261	0.253	0.253	485.10	467.91	498.47	483.83	483.83	483.83	1.213	1.170	1.246	1.210	1.151	1.165	
	3	1	0.241	0.244	0.244	0.243	460.28	466.00	464.09	466.00	464.09	464.09	1.151	1.165	1.165	1.160	1.151	1.160	
	2	0.254	0.251	0.252	0.252	0.252	485.10	479.37	481.28	481.92	481.92	481.92	1.213	1.198	1.203	1.205	1.222	1.227	
	3	0.256	0.254	0.257	0.256	0.256	488.92	485.10	490.83	488.29	488.29	488.29	1.222	1.213	1.213	1.221	1.222	1.221	
ST6	1	1	0.248	0.246	0.246	0.247	0.236	0.007	473.64	469.82	469.82	471.10	451.01	13.22	1.184	1.175	1.175	1.128	0.033
	2	0.242	0.244	0.242	0.243	0.243	462.18	466.00	462.18	463.46	463.46	463.46	1.155	1.165	1.155	1.159	1.155	1.155	
	3	0.245	0.244	0.244	0.244	0.244	467.91	466.00	466.00	466.64	466.64	466.64	1.170	1.165	1.165	1.167	1.103	1.103	
	2	1	0.231	0.231	0.230	0.231	441.18	441.18	439.27	440.54	440.54	440.54	1.103	1.103	1.098	1.101	1.122	1.122	
	2	0.235	0.235	0.236	0.235	0.235	448.82	448.82	450.73	449.45	449.45	449.45	1.122	1.122	1.127	1.124	1.117	1.117	
	3	0.234	0.238	0.234	0.235	0.235	446.91	454.55	446.91	449.45	449.45	449.45	1.084	1.084	1.103	1.090	1.117	1.117	
ST7	3	1	0.227	0.227	0.231	0.228	433.54	433.54	441.18	436.08	436.08	436.08	1.112	1.112	1.141	1.122	1.112	1.112	
	2	0.233	0.233	0.239	0.235	0.235	445.00	445.00	456.46	448.82	448.82	448.82	1.084	1.084	0.012	1.084	1.112	1.112	
	3	0.227	0.227	0.227	0.227	0.227	433.54	433.54	433.54	433.54	433.54	433.54	1.112	1.112	1.141	1.122	1.112	1.112	
	1	1	0.210	0.207	0.205	0.207	0.204	0.006	401.07	395.34	391.52	395.98	389.19	10.74	1.003	0.988	0.979	0.990	
	2	0.211	0.204	0.209	0.208	0.208	402.98	398.61	399.16	397.25	397.25	397.25	1.007	0.974	0.998	0.993	1.037	1.037	
	3	0.205	0.205	0.203	0.204	0.204	391.52	391.52	387.70	390.25	390.25	390.25	0.979	0.979	0.969	0.976	0.94	0.94	
ST8	2	1	0.188	0.191	0.192	0.190	359.05	364.78	366.69	363.51	363.51	363.51	0.898	0.912	0.917	0.909	0.88	0.88	
	2	0.201	0.201	0.203	0.202	0.202	383.88	383.88	387.70	385.15	385.15	385.15	0.960	0.960	0.969	0.963	0.99	0.99	
	3	0.205	0.206	0.203	0.205	0.205	391.52	393.43	387.70	390.88	390.88	390.88	0.979	0.984	0.969	0.977	0.955	0.955	
	3	1	0.200	0.203	0.203	0.202	381.97	387.70	387.70	385.79	385.79	385.79	0.953	0.969	0.969	0.964	0.955	0.955	
	2	0.208	0.208	0.208	0.208	0.208	397.25	397.25	397.25	397.35	397.35	397.35	0.993	0.993	0.993	0.993	0.993	0.993	
	3	0.208	0.208	0.207	0.208	0.208	397.25	397.25	395.34	396.61	396.61	396.61	0.993	0.993	0.988	0.992	0.966	0.966	

ตารางที่ 53 ปริมาณสดตราบที่เป็นของที่ประกอบในตัวอย่างสูตรอาหารที่ส่งเสริมสุขภาพ (ต่อ)

8.) ปริมาณเกลือ (Salt content)

ตารางที่ 54 ปริมาณเกลือที่เป็นองค์ประกอบในตัวอย่างสูตรอาหารหนูสูริมชิริชิน

ตัวอย่าง	ปริมาตร KSCN (ml)			ปริมาณเกลือที่เป็นองค์ประกอบ (salt content)				
	R1	R2	R3	R1	R2	R3	Ave.	SD
COM	24.30	24.23	24.15	0.14	0.15	0.17	<b>0.15</b>	0.015
CN5	23.60	23.70	23.60	0.27	0.25	0.27	<b>0.27</b>	0.011
CN6	23.55	23.57	23.60	0.28	0.28	0.27	<b>0.28</b>	0.005
CN7	23.40	23.43	23.65	0.31	0.31	0.26	<b>0.29</b>	0.027
CC7	23.90	23.80	23.80	0.21	0.23	0.23	<b>0.23</b>	0.011
CS7	23.55	23.60	23.65	0.28	0.27	0.26	<b>0.27</b>	0.010
ST5	23.95	23.70	23.67	0.20	0.25	0.26	<b>0.24</b>	0.030
ST6	24.00	23.60	23.70	0.19	0.27	0.25	<b>0.24</b>	0.041
ST7	24.05	24.00	23.80	0.19	0.19	0.23	<b>0.20</b>	0.026
SS7	23.75	23.70	23.75	0.24	0.25	0.24	<b>0.25</b>	0.006
SA7	24.10	24.00	24.05	0.18	0.19	0.19	<b>0.19</b>	0.010

## 1.2. ข้อมูลผลการวิเคราะห์ทางจุลชีววิทยา

1.) ค่า Log จำนวนจุลินทรีย์ทั้งหมดในตัวอย่างอาหารหมูที่เก็บรักษาไว้ที่อุณหภูมิและระยะเวลาต่าง ๆ

ตารางที่ 55 ค่า Log ของจำนวนจุลินทรีย์ทั้งหมดในตัวอย่างอาหารหมูเมื่อเก็บรักษาไว้ที่อุณหภูมิและระยะเวลาต่าง ๆ

Temp. (°C)	Time (weeks)	Aerobic Plate Count (CFU/g)						Log TCP (Log A)					
		M	C	S	CNC	SNC	A	M	C	S	CNC	SNC	A
25	0	3200	1300	18550	650	221000	ND	3.505	3.114	4.268	2.813	5.344	ND
	1	ND	3850	ND	1000	185000	250	ND	3.585	ND	3.000	5.267	2.398
	2	54500	5500	58500	2000	204000	5000	4.736	3.740	4.767	3.301	5.310	3.699
	3	185000	7000	30500	ND	ND	ND	5.267	3.845	5.484	ND	ND	ND
	4	41500	ND	19850	1750	109500	2000	4.618	ND	4.298	3.243	5.491	3.301
	5	10000	3000	16000	2650	ND	200	4.000	3.477	4.204	3.423	ND	2.301
	6	6150	4500	10000	1250	575000	250	3.789	3.653	4.000	3.097	5.760	2.398
	8	ND	1000	16450	1050	ND	1850	ND	3.000	4.216	3.021	ND	3.267
	10	3000	ND	25000	1000	410000	ND	3.477	ND	4.398	3.000	5.613	ND
	12	ND	ND	30000	ND	ND	2300	ND	ND	4.477	ND	ND	3.362
35	0	3200	1300	18550	650	221000	ND	3.505	3.114	4.268	2.813	5.344	ND
	1	ND	2000	ND	ND	169000	ND	ND	3.301	ND	ND	5.228	ND
	2	10500	4500	14500	3000	248500	3000	4.021	3.653	4.161	3.477	5.395	3.477
	3	2000	4000	24750	ND	ND	1800	3.301	3.602	4.394	ND	ND	3.255
	4	2000	5500	32550	2050	418000	5000	3.301	3.740	4.513	3.312	5.621	3.699
	5	1000	5000	ND	ND	ND	ND	3.000	3.699	ND	ND	ND	ND
	6	8000	30000	124000	10650	4225000	3950	3.903	4.477	5.093	4.027	6.626	3.597
	8	100	30000	14500	14000	1155000	1500	2.000	4.477	4.161	4.146	6.063	3.176
	10	ND	13500	16500	12850	70000	50	ND	4.130	4.217	4.109	4.845	1.699
	12	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
55	0	3200	1300	18550	650	221000	ND	3.505	3.114	4.268	2.813	5.344	ND
	1	ND	1000	20650	800	ND	ND	ND	3.000	4.315	2.903	ND	ND
	2	4000	1500	18500	2500	352500	5500	3.602	3.176	4.267	3.398	5.547	3.740
	3	1000	3500	25500	ND	ND	ND	3.000	3.544	4.407	ND	ND	ND
	4	1000	9500	22050	9100	216000	2050	3.000	3.978	4.343	3.959	5.334	3.312
	5	ND	ND	8500	ND	ND	ND	ND	ND	3.929	ND	ND	ND
	6	40000	85000	5000	16000	235000	2450	4.602	4.929	3.699	4.204	5.371	3.389
	8	10000	29500	4000	17500	26500	2500	4.000	4.470	3.602	4.243	4.423	3.398
	10	ND	ND	7150	7050	10000	300	ND	ND	3.854	3.848	4.000	2.477
	12	ND	ND	15000	ND	ND	ND	ND	ND	4.176	ND	ND	ND

ตารางที่ 56 การเปลี่ยนแปลงจำนวนจุลทรรศ์ทางหนองคายตามอุณหภูมิ 25°C ที่ระบุต่อไปนี้

Sample	Time e	weeks)	0	1	2	3	4	5	6	8	10	12
M	bacteria	2900	3500	ND	ND	49000	60000	ND	260000	340000	ND	ND
	yeast	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	yeast	100	ND	1000	ND	ND	ND	ND	ND	ND	ND	ND
	mold	100	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
C	(CFU/g)	2900	3500	ND	ND	49000	60000	ND	360000	430000	ND	ND
	APC	Y&M	200	ND	1000	ND	ND	ND	ND	ND	3000	3000
	(colonies)										16000	16000
	bacteria	1300	ND	ND	2000	6000	5000	9000	2000	ND	ND	ND
S	yeast	ND	ND	2500	3200	ND	ND	1000	2000	ND	ND	ND
	yeast	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	mold	20	30	ND	ND	ND	ND	ND	ND	ND	ND	ND
	(CFU/g)	1300	ND	2500	5200	6000	5000	10000	4000	ND	ND	ND
CNC	Y&M	20	30	ND	ND	ND	ND	ND	ND	ND	ND	ND
	(colonies)										20000	20000
	bacteria	17700	19400	9100	8800	42000	75000	320000	270000	19800	19200	16000
	yeast	ND	ND	ND	ND	ND	ND	10000	10000	400	300	ND
SNC	yeast	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	mold	10	20	ND	ND	ND	ND	ND	ND	ND	ND	ND
	(CFU/g)	17700	19400	9100	8800	42000	75000	330000	280000	20200	19500	16000
	Y&M	10	20	ND	ND	ND	ND	ND	ND	ND	ND	ND
A	(colonies)										11000	11000
	bacteria	ND	650	700	ND	0	3000	273000	22000	1300	1100	100
	yeast	ND	ND	900	400	1000	ND	1000	1000	600	500	ND
	yeast	70	100	ND	700	ND	ND	ND	ND	ND	ND	ND
A	mold	ND	10	ND	ND	ND	ND	ND	ND	ND	ND	ND
	(CFU/g)	ND	650	1600	400	1000	3000	274000	223000	1900	1600	1200
	Y&M	70	110	ND	700	ND	ND	ND	ND	ND	ND	ND
	(colonies)										1200	1200
SNC	bacteria	TNTC	TNTC	180000	190000	ND	407000	ND	292000	327000	242000	209000
	yeast	ND	ND	ND	ND	1000	ND	300000	200000	ND	ND	ND
	yeast	10	10	ND	ND	100	ND	ND	ND	ND	ND	ND
	mold	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
A	(CFU/g)	ND	ND	180000	190000	ND	408000	300000	292000	327000	242000	209000
	Y&M	10	10	ND	ND	100	ND	ND	ND	ND	ND	ND
	(colonies)										560000	560000
	bacteria	ND	ND	500	ND	4000	ND	ND	ND	ND	ND	ND
A	yeast	ND	ND	ND	ND	1000	ND	1000	ND	ND	ND	ND
	yeast	ND	ND	300	ND	ND	ND	ND	ND	ND	ND	ND
	mold	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	(CFU/g)	ND	ND	500	ND	5000	ND	400	100	ND	ND	ND
A	Y&M	ND	ND	300	ND	ND	ND	ND	ND	ND	ND	ND
	(colonies)										2600	2600

ตารางที่ 57 การประเมินแนวโน้มจำนวนบุคลากรที่เข้าสู่ห้องทดลองและรักษา และรักษาระดับน้ำของห้องทดลองต่อวันต่อสัปดาห์ ของอาคารห้องน้ำที่รักษาที่อุณหภูมิ 35 °C ที่ระบุไว้เวลาต่างๆ

ตารางที่ 58 การประเมินแนวโน้มจำนวนทรัพย์ทั้งหมด และรายได้ของอาชญากรรมที่อุบัติใหม่ 55°C ที่ระบุระยะเวลา

## 2. การทดสอบฤทธิ์ของซิริซินในการยับยั้งการเกิดมะเร็งลำไส้ใหญ่

ตารางที่ 59 น้ำหนักเฉลี่ยของหนูทดลองทั้ง 5 กลุ่ม ในระยะเวลา 5 เดือน

เดือน	น้ำหนัก (กรัม)				
	กลุ่ม 1	กลุ่ม 2	กลุ่ม 3	กลุ่ม 4	กลุ่ม 5
1	237.38 ± 11.61	234.67 ± 30.53	228.54 ± 37.24	206.04 ± 25.24	229.00 ± 35.06
2	352.96 ± 18.86	336.92 ± 53.01	342.83 ± 42.14	339.54 ± 28.61	343.67 ± 45.30
3	425.88 ± 27.40	411.17 ± 35.07	423.04 ± 31.32	413.17 ± 26.64	405.83 ± 55.70
4	477.75 ± 29.36	455.83 ± 37.84	473.29 ± 29.85	458.85 ± 21.44	446.65 ± 50.64
5	502.75 ± 28.25	488.83 ± 40.81	498.08 ± 29.73	483.375 ± 24.84	486.83 ± 48.42

หมายเหตุ แสดงค่าเป็น Mean±SD

ตารางที่ 60 ปริมาณการกินอาหารของหนูทดลองทั้ง 5 กลุ่ม ในระยะเวลา 5 เดือน

เดือน	น้ำหนัก (กรัม)				
	กลุ่ม 1	กลุ่ม 2	กลุ่ม 3	กลุ่ม 4	กลุ่ม 5
1	10.73 ± 2.76	12.52 ± 2.40	11.84 ± 2.70	8.41 ± 3.00	12.37 ± 2.80
2	17.90 ± 2.00	18.88 ± 2.30	19.07 ± 1.90	17.51 ± 3.40	17.78 ± 4.40
3	19.37 ± 0.90	20.10 ± 1.60	19.51 ± 0.90	21.97 ± 3.10	19.90 ± 1.90
4	21.59 ± 2.70	22.21 ± 1.60	24.62 ± 4.00	22.11 ± 1.20	21.89 ± 0.50
5	18.21 ± 0.90	19.37 ± 2.70	22.38 ± 2.70	21.89 ± 2.70	21.21 ± 1.00

หมายเหตุ แสดงค่าเป็น Mean±SD

ตารางที่ 61 น้ำหนักเฉลี่ยของอวัยวะต่างๆ หนูทดลองทั้ง 5 กลุ่ม ในระยะเวลา 5 เดือน

อวัยวะ	น้ำหนัก (กรัม)				
	กลุ่ม 1	กลุ่ม 2	กลุ่ม 3	กลุ่ม 4	กลุ่ม 5
ตับ	15.47 ± 0.74	15.23 ± 0.61	14.77 ± 0.67	13.64 ± 0.21	13.82 ± 0.74
ลำไส้เล็ก	10.06 ± 0.48	10.33 ± 0.39	10.49 ± 0.54	10.06 ± 0.47	10.03 ± 0.54
หัวใจ	1.65 ± 0.10	1.64 ± 0.10	1.61 ± 0.04	1.55 ± 0.07	1.51 ± 0.07
ปอด	3.40 ± 0.17	3.25 ± 0.12	3.53 ± 0.11	3.02 ± 0.15	3.26 ± 0.16
ม้าม	1.06 ± 0.19	0.98 ± 0.11	1.13 ± 0.17	1.02 ± 0.04	1.00 ± 0.14
ลำไส้ใหญ่	1.70 ± 0.16	1.72 ± 0.30	1.96 ± 0.32	1.77 ± 0.19	1.58 ± 0.33

หมายเหตุ แสดงค่าเป็น Mean±SD

ตารางที่ 62 จำนวนเฉลี่ยของ ACF ต่อพื้นที่ (ตารางเซนติเมตร) ในลำไส้ใหญ่ของหนูทดลองในกลุ่มที่ได้รับ DMH ในระยะเวลา 5 เดือน

กลุ่มที่	G3	G4	G5
จำนวนเฉลี่ยของ ACF ต่อพื้นที่ ( $\text{ซม}^2$ )	$116.00 \pm 48.50$	$79.30 \pm 29.30$	$82.40 \pm 39.80$

หมายเหตุ แสดงค่าเป็น Mean $\pm$ SD

ตารางที่ 63 จำนวนเฉลี่ยของ ACF ต่อพื้นที่ (ตารางเซนติเมตร) ในส่วนต่างๆ ของลำไส้ใหญ่ของหนูทดลอง ในกลุ่มที่ได้รับ DMH ในระยะเวลา 5 เดือน

ส่วนของลำไส้ใหญ่	จำนวนเฉลี่ยของ ACF ต่อพื้นที่ ( $\text{ซม}^2$ )		
	กลุ่ม 3	กลุ่ม 4	กลุ่ม 5
ส่วนต้น	$3.90 \pm 5.30$	0.00	0.00
ส่วนกลาง	$38.00 \pm 27.50$	$21.00 \pm 18.50$	$28.40 \pm 29.80$
ส่วนปลาย	$73.40 \pm 27.00$	$57.80 \pm 26.40$	$53.50 \pm 22.80$

หมายเหตุ แสดงค่าเป็น Mean $\pm$ SD

ตารางที่ 64 จำนวน ACF ต่อพื้นที่ (ตารางเซนติเมตร) ในลำไส้ใหญ่ส่วนปลายของหนูทดลองในกลุ่มที่ได้รับ DMH ในระยะเวลา 5 เดือน แบ่งตามประเภทของ ACF (1 ถึง  $\geq 5$  crypt/ACF)

ประเภทของ ACF	จำนวน ACF ต่อพื้นที่ ( $\text{ซม}^2$ )		
	กลุ่ม 3	กลุ่ม 4	กลุ่ม 5
1 crypt/ACF	$6.49 \pm 6.27$	$6.06 \pm 7.69$	$5.84 \pm 6.96$
2 crypts/ACF	$12.01 \pm 8.70$	$12.34 \pm 10.48$	$11.26 \pm 7.19$
3 crypts/ACF	$9.74 \pm 6.49$	$7.58 \pm 6.49$	$9.52 \pm 7.47$
4 crypts/ACF	$7.47 \pm 7.14$	$3.68 \pm 1.73$	$5.41 \pm 4.89$
$\geq 5$ crypts/ACF	$37.66 \pm 10.12$	$28.79 \pm 15.79$	$21.43 \pm 9.03$

หมายเหตุ แสดงค่าเป็น Mean $\pm$ SD

ตารางที่ 65 พื้นที่เฉลี่ยของ ACF ที่แสดงขนาดของ ACF ประเภทที่มีจำนวน crypt ใน ACF  $\geq 5$  crypts ในคำ释迦ใหญ่ส่วนปลายของหนูทดลองในกลุ่มที่ได้รับ DMH ในระยะเวลา 5 เดือน

	พื้นที่เฉลี่ยของ ACF ( $\mu\text{m}^2$ )		
	กลุ่ม 3	กลุ่ม 4	กลุ่ม 5
คำ释迦ใหญ่ส่วนปลาย	$0.14 \pm 0.06$	$0.11 \pm 0.05$	$0.14 \pm 0.06$

หมายเหตุ แสดงค่าเป็น Mean $\pm$ SD

ตารางที่ 66 การบีบอ้มติด surface markers (ร้อยละของเซลล์) ของในเลือดของหนูทดลองทั้ง 5 กลุ่ม ที่ได้จากการเจาะเลือดที่หัวใจ เมื่อสิ้นสุดการทดลองที่ระยะเวลา 5 เดือน

กลุ่มการทดลองที่	Surface marker (%)					
	CD2	CD3	CD4	CD8a	CD11a	CD25
1	$63.59 \pm 7.96$	$63.59 \pm 7.96$	$44.45 \pm 4.90$	$40.73 \pm 8.98$	$96.37 \pm 1.60$	$21.63 \pm 3.38$
2	$66.71 \pm 17.18$	$66.71 \pm 17.18$	$50.66 \pm 24.36$	$25.25 \pm 13.75$	$83.58 \pm 34.14$	$18.33 \pm 12.89$
3	$61.93 \pm 6.69$	$61.93 \pm 6.69$	$40.83 \pm 8.20$	$28.60 \pm 6.46$	$98.32 \pm 0.51$	$23.18 \pm 3.66$
4	$76.09 \pm 12.86$	$76.09 \pm 12.86$	$54.47 \pm 14.40$	$25.72 \pm 4.88$	$98.74 \pm 0.85$	$32.31 \pm 3.84$
5	$66.16 \pm 24.21$	$66.16 \pm 24.21$	$41.30 \pm 26.49$	$30.25 \pm 18.75$	$94.35 \pm 7.06$	$21.86 \pm 6.82$

กลุ่มการทดลองที่	Surface marker				
	CD45	CD54	CD80	CD86	RT1B
1	$99.84 \pm 0.18$	$74.98 \pm 7.17$	$23.55 \pm 8.24$	$20.08 \pm 5.01$	$22.48 \pm 5.87$
2	$85.53 \pm 22.24$	$68.32 \pm 11.10$	$10.28 \pm 5.74$	$29.13 \pm 32.16$	$27.58 \pm 15.57$
3	$99.66 \pm 0.27$	$77.53 \pm 2.99$	$20.98 \pm 3.81$	$21.31 \pm 5.39$	$26.09 \pm 3.13$
4	$99.74 \pm 0.41$	$77.65 \pm 9.21$	$17.15 \pm 9.06$	$14.67 \pm 5.23$	$20.76 \pm 2.18$
5	$98.98 \pm 0.75$	$79.35 \pm 6.89$	$19.85 \pm 5.51$	$24.50 \pm 11.13$	$35.28 \pm 23.20$

หมายเหตุ แสดงค่าเป็น Mean $\pm$ SD

### 3. การทดสอบฤทธิ์ของซิริซินในการยับยั้งการดูดซึมโคเลสเตอรอล

ตารางที่ 67 ผลของซิริซิน 50  $\mu\text{g}/\text{ml}$  ต่อการนำเข้าของโคเลสเตอรอล ในเซลล์เพาะเลี้ยง Caco-2

Sample (50 $\mu\text{g}/\text{ml}$ )	Cholesterol uptake (% control)						Mean	SD.	SEM
	1	2	3	4	5	6			
PBS	100.00	100.00	100.00	100.00	100.00	100.00	100.00	0.00	0.00
EZE	50.07	57.47	60.64	56.56	67.49	61.09	58.89	5.78	2.36
BSA	84.19	76.73	129.86	110.79	104.34	89.20	99.18	19.65	8.02
sericin A	65.67	86.08	80.28	84.84	83.45	88.42	81.46	8.20	3.35
sericin B	82.16	99.42	74.03	92.17	71.78	62.62	80.36	13.67	5.58
sericin C	86.86	96.55	106.95	114.88	75.65	83.62	94.08	14.87	6.07

หมายเหตุ EZE = ezetimibe 100  $\mu\text{M}$

ตารางที่ 68 ผลของซิริซิน 100  $\mu\text{g}/\text{ml}$  ต่อการนำเข้าของโคเลสเตอรอล ในเซลล์เพาะเลี้ยง Caco-2

Sample (100 $\mu\text{g}/\text{ml}$ )	Cholesterol uptake (% control)					Mean	SD.	SEM
	1	2	3	4	5			
PBS	100.00	100.00	100.00	100.00	100.00	100.00	0.00	0.00
EZE			50.21	83.05		83.05	23.22	16.42
BSA			93.37	117.92		117.92	17.36	12.27
sericin A	56.14	119.34	74.30	109.48	92.58	90.37	25.69	11.49
sericin B	63.16	101.23		125.43	98.89	97.18	25.66	12.83
sericin C	66.08	124.28	113.19	134.12	121.58	111.85	26.65	11.92

หมายเหตุ EZE = ezetimibe 100  $\mu\text{M}$

ตารางที่ 69 ผลของซิริซิน 500  $\mu\text{g}/\text{ml}$  ต่อการนำเข้าของโคเลสเตอรอล ในเซลล์เพาะเลี้ยง Caco-2

Sample (500 $\mu\text{g}/\text{ml}$ )	Cholesterol uptake (% control)						Mean	SD.	SEM
	1	2	3	4	5	6			
PBS	100.00	100.00	100.00	100.00	100.00	100.00	100.00	0.00	0.00
EZE	60.64	56.56	67.49	61.09	50.07	57.47	64.29	4.51	2.26
BSA	121.25	106.65	103.78	119.01	77.50	98.32	111.40	8.74	4.37
sericin A	107.77	90.54	84.90	88.34	107.92	105.74	86.62	10.19	5.10
sericin B	113.28	97.54	101.87	95.46		84.47	98.66	7.96	3.98
sericin C	109.23	91.40	102.57	73.74		97.99	102.57	9.01	7.76

หมายเหตุ EZE = ezetimibe 100  $\mu\text{M}$

ตารางที่ 70 ผลของซิริซิน 1000 μg/ml ต่อการนำเข้าของโคเลสเตอรอล ในเซลล์เพาะเลี้ยง Caco-2

Sample (1000ug/ml)	Cholesterol uptake (% control)				Mean	SD.	SEM
	1	2	3	4			
PBS	100.00	100.00	100.00	100.00	100.00	0.00	0.00
EZE			83.05	66.39	74.72	11.78	8.33
BSA	111.81	114.93	110.66	110.66	110.66	2.02	1.01
sericin A	149.25	122.92	126.67	133.84	130.26	11.64	5.82
sericin B	128.78	115.68	122.62	108.03	115.33	8.94	4.47
sericin C	122.73	129.98	142.22	129.20	135.71	8.14	4.07

หมายเหตุ EZE = ezetimibe 100 μM

ตารางที่ 71 น้ำหนักของหนูทดลองที่ได้รับอาหารชนิดต่างๆ ในช่วงระยะเวลา 5 เดือน

ชนิดของอาหาร	น้ำหนักตัว (กรัม)					
	เริ่มต้น	เดือนที่ 1	เดือนที่ 2	เดือนที่ 3	เดือนที่ 4	เดือนที่ 5
Control diet	189.67±9.61	237.38±7.09	352.96±18.12	42588±26.39	496.83±27.72	502.75±27.71
Sericin diet	194.17±27.00	234.67±27.58	336.92±50.88	411.17±33.21	465.67±41.41	483.85±41.41
High cholesterol diet	175.33±23.21	345.83±17.74*	433.50±24.31*	487.67±24.95*	510.83±27.29	514.83±26.61
High cholesterol+ sericin diet	210.83±4.45*#	337.33±31.05*	414.67±17.48*	477.17±20.34*	491.50±19.67	519.50±17.92

หมายเหตุ แสดงค่าเป็น Mean±SD

\*p<0.05 เปรียบเทียบกับ control diet

# p<0.05 เปรียบเทียบระหว่างอาหารประเภทเดียวกันที่มีหรือไม่มีซิริซิน

ตารางที่ 72 การเปลี่ยนแปลงของน้ำหนักของหนูทดลองที่ได้รับอาหารชนิดต่างๆ ในแต่ละช่วงระยะเวลา 5 เดือน

ชนิดของอาหาร	น้ำหนักตัวที่เปลี่ยนแปลง (กรัม)				
	เดือนที่ 1	เดือนที่ 2	เดือนที่ 3	เดือนที่ 4	เดือนที่ 5
Control diet	47.71±11.19	163.29±9.47	236.21±18.59	307.17±19.99	313.08±23.08
Sericin diet	40.50±8.12	142.75±30.60	217.00±17.04	271.50±25.52*#	289.68±19.14
High cholesterol diet	170.50±16.65*	258.17±35.70*	312.33±32.64*	335.50±33.36	339.50±31.79
High cholesterol+ sericin diet	126.50±30.26*#	203.83±14.82*#	263.33±18.12*#	280.67±19.72*#	308.67±16.91

หมายเหตุ แสดงค่าเป็น Mean±SD

\*p<0.05 เปรียบเทียบกับ control diet

# p<0.05 เปรียบเทียบระหว่างอาหารประเภทเดียวกันที่มีหรือไม่มีซิริซิน

ตารางที่ 73 ปริมาณอาหารที่กินของหนูทดลองที่ได้รับอาหารชนิดต่างๆ ในช่วงระยะเวลา 5 เดือน

ชนิดของอาหาร	ปริมาณอาหาร (กรัม)				
	เดือนที่ 1	เดือนที่ 2	เดือนที่ 3	เดือนที่ 4	เดือนที่ 5
Control diet	10.73±2.58	17.90±2.00	19.37±0.95	21.59±2.67	18.21±0.92
Sericin diet	12.52±2.38	18.88±2.26	20.10±1.61	22.21±1.64	19.37±2.67
High cholesterol diet	16.43±0.27*	20.42±0.43*	17.55±0.65*	28.03±0.54*	30.00±0.00*
High cholesterol + sericin diet	15.25±0.83*#	19.92±0.20*#	23.09±0.40*#	27.13±0.05*#	30.00±0.00*

หมายเหตุ แสดงค่าเป็น Mean±SD

\* $p<0.05$  เปรียบเทียบกับ control diet

#  $p<0.05$  เปรียบเทียบระหว่างอาหารประเภทเดียวกันที่มีหรือไม่มีซิริซิน

ตารางที่ 74 ระดับของ total cholesterol ในเลือดของหนูทดลองที่ได้รับอาหารชนิดต่างๆ ในช่วงระยะเวลา 5 เดือน

ชนิดของอาหาร	Total cholesterol (mg/dl)				
	เริ่มต้น	เดือนที่ 2	เดือนที่ 3	เดือนที่ 4	เดือนที่ 5
Control diet	86.33±3.06	117.00±9.54	105.50±1.99	104.12±1.48	89.24±11.98
Sericin diet	85.67±3.79	114.00±13.11	138.47±3.35*#	116.22±3.20*#	82.80±5.60
High cholesterol diet	120.67±8.19*	134.83±18.71	128.56±10.40*	152.37±21.96*	156.50±24.36*
High cholesterol + sericin diet	104.17±11.02*#	118.51±63.95#	123.32±8.64*	141.21±13.13*	152.00±15.40*

หมายเหตุ แสดงค่าเป็น Mean±SD

\* $p<0.05$  เปรียบเทียบกับ control diet

#  $p<0.05$  เปรียบเทียบระหว่างอาหารประเภทเดียวกันที่มีหรือไม่มีซิริซิน



ตารางที่ 75 ระดับของ High density lipoprotein (HDL) ในเลือดของหนูทดลองที่ได้รับอาหารชนิดต่างๆ ในช่วงระยะเวลา 4 เดือน

ชนิดของอาหาร	HDL (mg/dl)				
	เริ่มต้น	เดือนที่ 2	เดือนที่ 3	เดือนที่ 4	เดือนที่ 5
Control diet	61.33±2.89	47.00±3.46	45.75±4.08	38.86±1.47	17.13±1.41
Sericin diet	65.67±4.16	38.67±3.06*#	51.39±11.34	35.47±2.80*#	18.85±3.47
High cholesterol diet	39.95±6.68*	17.23±2.53*	24.69±1.99*	50.74±18.23	27.34±12.80
High cholesterol + sericin diet	65.00±4.90#	19.46±1.63*#	21.81±1.97*#	52.21±11.69	32.35±5.79*

หมายเหตุ แสดงค่าเป็น Mean±SD

\* $p<0.05$  เปรียบเทียบกับ control diet

#  $p<0.05$  เปรียบเทียบระหว่างอาหารประเภทเดียวกันที่มีหรือไม่มีซิริซิน

ตารางที่ 76 ระดับของ Triglyceride ในเลือดของหนูทดลองที่ได้รับอาหารชนิดต่างๆ ในช่วงระยะเวลา 5 เดือน

ชนิดของอาหาร	HDL (mg/dl)				
	เริ่มต้น	เดือนที่ 2	เดือนที่ 3	เดือนที่ 4	เดือนที่ 5
Control diet	108.33±9.61	105.33±29.14	106.86±2.02	122.68±23.11	75.27±16.26
Sericin diet	82.33±12.50*#	65.00±19.16	86.08±0.34*#	88.69±8.28*#	64.04±14.20
High cholesterol diet	44.17±9.50*	208.34±50.26*	95.29±10.98	93.64±23.97*	75.33±14.83
High cholesterol + sericin diet	82.50±15.86*#	221.91±18.16*	72.36±10.10*#	103.56±22.13*	65.33±12.13

หมายเหตุ แสดงค่าเป็น Mean±SD

\* $p<0.05$  เปรียบเทียบกับ control diet

#  $p<0.05$  เปรียบเทียบระหว่างอาหารประเภทเดียวกันที่มีหรือไม่มีซิริซิน

ตารางที่ 77 ระดับของ total cholesterol ในเลือดของหนูทดลองที่ได้รับอาหารชนิดต่างๆ ในช่วง 14 วัน

Treatments	Serum total cholesterol (mg/dl)				
	0	4 days	8 days	11 days	14 days
High-cholesterol	80.23±12.35	108.35±17.91 <sup>#</sup>	151.99±6.24 <sup>#</sup>	164.62±20.46 <sup>#</sup>	177.68±20.05 <sup>#</sup>
+Sericin 10 mg/kg	98.20±9.77	116.76±6.31	127.60±6.71**	144.04±5.83	139.25±2.67**
+Sericin 100 mg/kg	92.67±8.70	112.82±9.29	131.30±8.32**	143.73±9.92	131.58±6.90**
+Sericin 1000 mg/kg	81.87±11.74	120.84±11.70	124.79±12.78**	133.33±10.17*	130.92±6.42**

หมายเหตุ <sup>#</sup> เปรียบเทียบระหว่างค่าในและวันกับวันที่ 0 (baseline)

\*เปรียบเทียบระหว่างกลุ่มทดลองกับกลุ่ม High-cholesterol ( $p<0.05$ )

\*\*เปรียบเทียบระหว่างกลุ่มทดลองกับกลุ่ม High-cholesterol ( $p<0.01$ )

ตารางที่ 78 ระดับของ non-HDL cholesterol ในเลือดของหนูทดลองที่ได้รับอาหารชนิดต่างๆ ในช่วง 14 วัน

Treatments	Serum non-HDL (mg/dl)				
	0	4 days	8 days	11 days	14 days
High-cholesterol	45.83±18.14	74.53±18.05 <sup>#</sup>	121.15±7.48 <sup>#</sup>	136.78±17.46 <sup>#</sup>	143.66±13.72 <sup>#</sup>
+Sericin 10 mg/kg	64.03±6.27	76.01±4.40	100.87±5.33**	113.32±6.74*	103.29±2.07**
+Sericin 100 mg/kg	56.00±10.65	71.28±8.84	103.20±6.32**	107.93±8.91*	91.97±7.74**
+Sericin 1000 mg/kg	41.43±5.85	77.79±11.81	99.38±11.07**	98.37±10.13**	93.18±7.49**

หมายเหตุ <sup>#</sup> เปรียบเทียบระหว่างค่าในและวันกับวันที่ 0 (baseline)

\*เปรียบเทียบระหว่างกลุ่มทดลองกับกลุ่ม High-cholesterol ( $p<0.05$ )

\*\*เปรียบเทียบระหว่างกลุ่มทดลองกับกลุ่ม High-cholesterol ( $p<0.01$ )

ตารางที่ 79 ระดับของ triglyceride ในเลือดของหนูทดลองที่ได้รับอาหารชนิดต่างๆ ในช่วง 14 วัน

Treatments	Serum triglyceride (mg/dl)				
	0	4 days	8 days	11 days	14 days
High-cholesterol	71.92±10.82	105.76±29.63 <sup>#</sup>	112.64±39.10	93.81±23.93	127.98±39.44 <sup>#</sup>
+Sericin 10 mg/kg	91.55±11.25	140.27±8.52*	129.31±31.79	100.93±8.67	122.26±18.57
+Sericin 100 mg/kg	53.94±18.37	86.06±27.82	100.47±31.18	143.29±29.64*	126.73±27.96
+Sericin 1000 mg/kg	44.80±12.73	96.67±24.12	91.91±15.66	125.40±29.61	142.66±7.89

หมายเหตุ \* เปรียบเทียบระหว่างค่าในและวันกับวันที่ 0 (baseline)

\* เปรียบเทียบระหว่างกลุ่มทดลองกับกลุ่ม High-cholesterol ( $p<0.05$ )

ตารางที่ 80 ระดับของ triglyceride ในเลือดของหนูทดลองที่ได้รับอาหารชนิดต่างๆ ในช่วง 14 วัน

Treatments	Serum HDL (mg/dl)				
	0	4 days	8 days	11 days	14 days
High-cholesterol	34.40±6.90	33.82±0.97	30.84±2.57	27.85±8.96	34.02±6.83
+Sericin 10 mg/kg	34.17±4.34	40.76±3.62**	26.73±1.82*	30.71±2.31	35.97±1.12
+Sericin 100 mg/kg	36.97±2.44	41.54±2.87**	28.10±2.44	35.81±2.28	39.61±2.76
+Sericin 1000 mg/kg	40.44±7.74	43.06±1.01**	25.40±1.95**	34.96±1.88	37.74±2.05

หมายเหตุ \* เปรียบเทียบระหว่างกลุ่มทดลองกับกลุ่ม High-cholesterol ( $p<0.05$ )

\*\* เปรียบเทียบระหว่างกลุ่มทดลองกับกลุ่ม High-cholesterol ( $p<0.01$ )

## 4. การพัฒนาอาหารเสริมชิรซิน

### 4.1. การวิเคราะห์หาปริมาณ % Drug content

ตารางที่ 81 ตารางแสดง % drug content

ตัวรับ	LOT	Sericin B ในเม็ดยา ตามทฤษฎี (mg)	Absorbance 275 nm	Sericin B (mg/ml)	Sericin B ที่มีอยู่จริงในเม็ดยา (mg)	% drug content	Mean	SD
A1	1	350.0	0.5520	0.70	351.71	100.49	100.55	0.46
		350.0	0.5550	0.71	353.62	101.03		
		350.0	0.5500	0.70	350.44	100.13		
		350.0	0.5623	0.72	358.28	102.37		
		350.0	0.5630	0.72	358.71	102.49		
	2	350.0	0.5943	0.76	378.63	108.18	108.24	0.52
		350.0	0.5977	0.76	380.75	108.79		
		350.0	0.5920	0.75	377.15	107.76		
		350.0	0.6013	0.77	383.08	109.45		
		350.0	0.5947	0.76	378.84	108.24		
	3	350.0	0.6187	0.79	394.10	112.60	110.30	2.17
		350.0	0.5950	0.76	379.05	108.30		
		350.0	0.6043	0.77	384.99	110.00		
		350.0	0.6200	0.79	394.95	112.84		
		350.0	0.5937	0.76	378.21	108.06		
A2	1	350.0	0.5830	0.74	371.42	106.12	106.65	1.13
		350.0	0.5817	0.74	370.58	105.88		
		350.0	0.5930	0.76	377.78	107.94		
		350.0	0.5973	0.76	380.54	108.73		
		350.0	0.6017	0.77	383.29	109.51		
	2	350.0	0.6060	0.77	386.05	110.30	109.47	0.72
		350.0	0.5990	0.76	381.60	109.03		
		350.0	0.5993	0.76	381.81	109.09		
		350.0	0.6083	0.78	387.53	110.72		
		350.0	0.6070	0.77	386.68	110.48		
	3	350.0	0.6057	0.77	385.84	110.24	110.54	0.30
		350.0	0.6090	0.78	387.96	110.84		
		350.0	0.6073	0.77	386.90	110.54		
		350.0	0.5943	0.76	378.63	108.18		
		350.0	0.6077	0.77	387.11	110.60		

ตารางที่ 81 ตารางแสดง % drug content (ต่อ)

ลำดับ	LOT	sericin B ในเม็ดยา ตามทฤษฎี (mg)	Absorbance 275 nm	sericin B (mg/ml)	sericin B ที่มีอยู่จริงในเม็ดยา (mg)	% drug content	Mean	SD
A3	1	350.0	0.5823	0.74	371.00	106.00	103.56	4.18
		350.0	0.5423	0.69	345.56	98.73		
		350.0	0.5820	0.74	370.79	105.94		
		350.0	0.6053	0.77	385.62	110.18		
		350.0	0.6053	0.77	385.62	110.18		
	2	350.0	0.5703	0.73	363.37	103.82	106.73	2.88
		350.0	0.5867	0.75	373.75	106.79		
		350.0	0.6020	0.77	383.51	109.57		
		350.0	0.6003	0.76	382.45	109.27		
		350.0	0.6080	0.77	387.32	110.66		
	3	350.0	0.6077	0.77	387.11	110.60	108.30	2.24
		350.0	0.5943	0.76	378.63	108.18		
		350.0	0.5830	0.74	371.42	106.12		
		350.0	0.5877	0.75	374.39	106.97		
		350.0	0.5860	0.75	373.33	106.67		
B2	1	350.0	0.5517	0.70	351.50	100.43	99.74	0.89
		350.0	0.5423	0.69	345.56	98.73		
		350.0	0.5497	0.70	350.23	100.06		
		350.0	0.5640	0.72	359.34	102.67		
		350.0	0.5927	0.76	377.57	107.88		
	2	350.0	0.5673	0.72	361.46	103.27	104.28	1.60
		350.0	0.5830	0.74	371.42	106.12		
		350.0	0.5682	0.72	362.03	103.44		
		350.0	0.5493	0.70	350.01	100.00		
		350.0	0.5930	0.76	377.78	107.94		
	3	350.0	0.5363	0.68	341.75	97.64	101.30	5.61
		350.0	0.5920	0.75	377.15	107.76		
		350.0	0.5410	0.69	344.72	98.49		
		350.0	0.6090	0.78	387.96	110.84		
		350.0	0.6067	0.77	386.47	110.42		

#### 4.2. การพัฒนาคำรับยาเม็ด sericin B เมื่องตันด้วยเทคนิค Wet granulation

พัฒนาคำรับยาเม็ด Sericin B Protein โดยการเปลี่ยนแปลงปัจจัยในกระบวนการผลิตดังตาราง  
ข้างล่างและมีวิธีการเตรียมดังนี้

1. ชั่ง explatab 1.5%, Avicel PH 101 และ Cericin B แล้วให้เข้ากันโดยใช้วิธีผสมแบบ Geometric dilution (เริ่มผสมจากสารที่มีปริมาณน้อยไปหาสารที่มีปริมาณมาก) โดยทำการ ผสมสารในโกร่ง
2. เมื่อผสมเสร็จแล้วให้ spray น้ำปริมาณ ~ 4-8 ml (สำหรับเตรียมทำยาอม 6 เม็ด) ให้เป็นก้อนหมวด
3. นำไปผ่านแรร์ No. 14 แล้วอบแห้งที่ 60 °C นาน 4 ชั่วโมง เมื่อแกรนูลแห้งจากนั้นจึงนำไปผ่านแรร์ No.16
4. ซึ่งน้ำหนักของแกรนูลที่ได้ จากนั้นเติม explatab อีก 1.5% และ Aerosil 200 0.2% (โดยคำนวณจากน้ำหนักแกรนูลที่ได้หลังผ่านการอบ) โดยทำการผสมแบบ Geometric dilution ในถุง
5. เมื่อผสมเข้ากันดีแล้วจากนั้นจึงนำสารมาซั่งก่อนนำไปทำการตอกเม็ดด้วยเครื่อง Hydrolic pressure (ใช้แรงตอก 1 ตัน ค้างนาน 1 นาที)
6. ทำการเปลี่ยนแปลงปัจจัยตามตารางข้างล่างกำหนด

ตารางที่ 82 แสดงตัววินิจฉัยของยาตัวรับเม็ดยา Diluent, ชนิด Diluent กับ % Explotab

ตัวรับ	ปัจจัยที่เปลี่ยนแปลง	Ingredient					Result
		Sericin B (mg)	Avicel PH101 (mg)	Lactose (mg)	Explotab (%)	Aerosil 200 Isopropanol ๙๘ : ๒	
A 1	Avicel PH 101	500	100	-	3%	0.2%	0 : 100 0.408 > 30
A 2		500	150	-	3%	0.2%	0 : 100 0.415 > 30
A 3		500	200	-	3%	0.2%	0 : 100 0.465 > 30
A 4	Lactose	500	-	100	3%	0.2%	0 : 100 1 ㎜ hold 1 นาที 0.415 > 30
A 5		500	-	200	3%	0.2%	0 : 100 0.482 > 30
A 6		500	-	300	3%	0.2%	0 : 100 0.551 > 30
A 7		500	-	200	4%	0.2%	0 : 100 0.503 > 30
A 8	Explotab (%)	500	-	200	5%	0.2%	0 : 100 0.504 > 30
A 9		500	-	200	6%	0.2%	0 : 100 0.504 > 30

ตารางที่ 83 ผลิตภัณฑ์ป้องกันเชื้อราและปรุงรักษา Diluent ที่มี Diluent กับ % Explotab

ตัวอย่าง	ไขข้อที่ ญี่ปุ่นสัมภาระ	Ingredient				Result			
		Sericin B (mg)	Avicel PH101 (mg)	Corn starch(mg)	Explotab (%)	Aerosil 200 (%)	Isopropanol : น้ำ		
B1	Avicel PH 101	400	200	-	3%	0.2%	0 : 100	hold 1 นาที	0.398 > 30
B2		400	300	-	3%	0.2%	0 : 100	hold 1 นาที	0.449 > 30
B3		400	400	-	3%	0.2%	0 : 100	1 ตัน	0.514 > 30
B4		400	300	-	4%	0.2%	0 : 100	hold 1 นาที	0.444 > 30
B5	Explotab (%)	400	300	-	5%	0.2%	0 : 100	hold 1 นาที	0.445 > 30
B6		400	300	-	6%	0.2%	0 : 100	hold 1 นาที	0.429 > 30
B7	Avicel PH 101 ผสม	400	-	300	3%	0.2%	0 : 100	1 ตัน	0.456 > 30
B8	Corn starch	400	150	150	3%	0.2%	0 : 100	hold 30 วินาที	0.456 > 30
B9		400	-	400	3%	0.2%	0 : 100	hold 30 วินาที	0.535 > 30
B10		400	200	200	3%	0.2%	0 : 100	hold 30 วินาที	0.535 > 30
B11		400	-	300	6%	0.2%	0 : 100	1 ตัน	0.492 > 30
B12	Explotab (%)	400	-	300	12%	0.2%	0 : 100	hold 30 วินาที	0.505 > 30
B13	มาด้า	400	-	300	15%	0.2%	0 : 100	hold 30 วินาที	0.509 > 30
B14	Corn starch	400	-	400	6%	0.2%	0 : 100	1 ตัน	0.569 > 30
B15		400	-	400	9%	0.2%	0 : 100	hold 1 นาที	0.552 > 30
B16		400	-	400	12%	0.2%	0 : 100	hold 1 นาที	0.549 > 30

ตารางที่ 84 แสดงส่วนประกอบของยาทาร์นีโนบิลีบันแบล็คบรูนยา Sericin กับ Diluent

ตัวรับ	ปูดจักษ์ บีดชั่นเบล็ง	Ingredient					ความหนา (mm)	เด็กตัวไม่น้ำ (นาฬิก)	Result
		Sericin B (mg)	Avicel PH101 (mg)	Corn starch (mg)	Explotab	Aerosil 200 Isopropanol : น้ำ			
C1		350	-	350	9%	0.2%	0 : 100	1 ต่อน hold 30 วินาที	0.479 > 30
C2	Sericin B ผลิต	350	-	450	9%	0.2%	0 : 100	1 ต่อน hold 1 นาที	0.547 > 30
C3	Corn starch	300	-	400	9%	0.2%	0 : 100	1 ต่อน hold 30 วินาที	0.477 22
C4		300	-	500	9%	0.2%	0 : 100	1 ต่อน hold 1 นาที	0.553 > 30

ตราที่ 85 แสดงถึงรากของอนุกรม  $\{I_{n+1}\}$  ที่ได้จากการหักตัวค่า  $I_n$  ออกจากตัวค่า  $I_{n+1}$  ทั้งนี้โดยทันที

ດំរើន	ប្រចាំថ្ងៃ ប្រតិបត្តិមុនបែង	Ingredient					Result	
		Sericin B (mg)	Avicel PH101 (mg)	Corn starch(mg)	Explotab (%)	Aerosil 200 Isopropanol : ឬ	ការអេអង (mm)	ដំឡើងអេអង (មាតិ)
D1		300	-	400	9%	0.2%	25 : 75	0.547
D2	Isopropanol : ឬ	300	-	400	9%	0.2%	50 : 50	0.569
D3		300	-	400	9%	0.2%	75 : 25	0.549
D4		300	-	400	9%	0.2%	100 : 0	0.552

ตารางที่ 86 ผลสัมฤทธิ์ทางเคมีของตัวรับเมื่อเปรียบเทียบยาปฏิชีวนะ Sericin B กับ อัตราส่วน Isopropanol: น้ำ

ตัวรับ	ปัจจัย ปฏิเสธยาปฏิชีวนะ	Ingredient				Result			
		Sericin B (mg)	Corn starch(mg)	Explotab (%)	Aerosil 200 (%)	Isopropanol น้ำ : น้ำ	แรงดึงดูด	ความหนา (mm)	แตกต่างหน้า ทางกายภาพ (นาที)
E1	Sericin B แตะ	350	400	9%	0.2%	50 : 50	1 ตัน hold	0.513	15
E2	Isopropanol	350	400	9%	0.2%	75 : 25	30 วินาที	0.511	15
E3	น้ำ : น้ำ	400	400	9%	0.2%	50 : 50	1 ตัน	0.528	23
E4		400	400	9%	0.2%	75 : 25	hold 1 นาที	0.520	19

หมายเหตุ เมื่อตัดจากตัวรับที่เปรียบเทียบ Sericin B และ Isopropanol: น้ำ มีร้อยละต่ำกว่า 10%



ตารางที่ 87 เมตริกส่วนประกอบของตัวรับเม็ดถ่านยาปฏิชินแบบเจลกับปริมาณ Diluent กับ % Explotab

ตัวรับ	ปัจจัยที่ เปลี่ยนแปลง	Ingredient						Result
		Sericin B (mg)	Avicel PH101 (mg)	Corn starch(mg)	Explotab	Aerosil 200	Isopropanol : น้ำ	
F1	Avicel PH 101 ผลิต	350	100	300	9%	0.2%	50 : 50	0.516 12
F2	Corn starch	350	200	200	9%	0.2%	50 : 50	0.501 12
F3		350	300	100	9%	0.2%	50 : 50	hold 30 วันทึบ 0.537 7
F4		350	400	0	9%	0.2%	50 : 50	0.533 10
F5		350	300	100	0%	0.2%	50 : 50	1 ตัน 0.485 6
F6	Exploratab (%)	350	300	100	3%	0.2%	50 : 50	hold 30 วันทึบ 0.507 7
F7		350	300	100	6%	0.2%	50 : 50	0.530 4
F8	Avicel PH 101 ผลิต	350	262.5	87.5	0%	0.2%	50 : 50	1 ตัน hold 0.464 8
F9	Corn starch	350	225	75	0%	0.2%	50 : 50	30 วันทึบ 0.407 19

หมายเหตุ เม็ดยาตัวรับที่เปลี่ยนแปลง Avicel PH 101 ผลิต Corn starch น้ำอ้อยแตกบริเวณๆ อบ

